



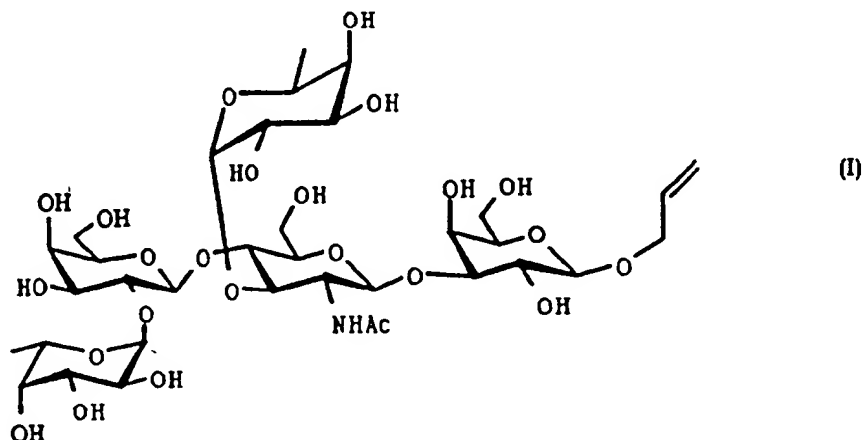
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(54) Title: SYNTHESIS OF GLYCOCONJUGATES OF THE LEWIS Y EPITOPE AND USES THEREOF

(57) Abstract

The present invention provides a method of synthesizing an allyl pentasaccharide having structure (I) as well as related oligosaccharide ceramides and other glycoconjugates useful as vaccines for inducing antibodies to epithelial cancer cells in an adjuvant therapy therefor, and in a method for preventing recurrence of epithelial cancer.



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**SYNTHESIS OF GLYCOCONJUGATES OF THE
LEWIS Y EPITOPE AND USES THEREOF**

5

This application is a continuation-in-part of U.S. Serial No. 08/430,355, filed April 28, 1995, which was a continuation-in-part of U.S. Serial No. 08/213,053, filed March 15, 1994, the contents of which are hereby incorporated by reference into this application.

This invention was made with government support under grants GM-15240-02, GM-16291-01, HL-25848-14 and AI-16943 from the National Institutes of Health. Accordingly, the U.S. Government has certain rights in the invention.

Background of the Invention

Throughout this application, citations for various publications are provided within parentheses in the text. The disclosures of these publications are hereby incorporated in their entirety by reference into this application in order to more fully describe the state of the art to which this invention pertains.

The function of carbohydrates as structural materials and as energy storage units in biological systems is well recognized. By contrast, the role of carbohydrates as signaling molecules in the context of biological processes has only recently been appreciated. (M.L. Phillips, E. Nudelman, F.C.A. Gaeta, M. Perez, A.K. Singhal, S. Hakomori, J.C. Paulson, *Science*, 1990, 250, 1130; M.J. Polley, M.L. Phillips, E. Wagner, E. Nudelman, A.K. Singhal, S. Hakomori, J.C. Paulson, *Proc. Natl. Acad. Sci. USA*, 1991, 88, 6224; T. Taki, Y. Hirabayashi, H. Ishikawa, S. Kon, Y. Tanaka, M. Matsumoto, *J. Biol. Chem.*, 1986, 261, 3075; Y. Hirabayashi, A. Hyogo, T. Nakao, K. Tsuchiya, Y. Suzuki, M. Matsumoto, K. Kon, S. Ando, *ibid.*, 1990, 265, 8144; O. Hindsgaul, T. Norberg,

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J. Le Pendu, R. U. Lemieux, *Carbohydr. Res.*, **1982**, 109, 109; U. Spohr, R.U. Lemieux, *ibid.*, **1988**, 174, 211)

The elucidation of the scope of carbohydrate involvement in mediating cellular interaction is an important area of inquiry in contemporary biomedical research. The carbohydrate molecules, carrying detailed structural information, tend to exist as glycoconjugates (cf. glycoproteins and glycolipids) rather than as free entities. Given the complexities often associated with isolating the conjugates in homogeneous form and the difficulties in retrieving intact carbohydrates from these naturally occurring conjugates, the applicability of synthetic approaches is apparent. (For recent reviews of glycosylation see: Paulsen, H., *Angew. Chem. Int. Ed. Engl.*, **1982**, 21, 155; Schmidt, R.R., *Angew. Chem. Int. Ed. Engl.*, **1986**, 25, 212; Schmidt, R.R., *Comprehensive Organic Synthesis*, Vol. 6, Chapter 1(2), Pergamon Press, Oxford, **1991**; Schmidt, R.R., *Carbohydrates, Synthetic Methods and Applications in Medicinal Chemistry*, Part I, Chapter 4, VCH Publishers, Weinheim, New York, **1992**. For the use of glycals as glycosyl donors in glycoside synthesis, see Lemieux, R.U., *Can. J. Chem.*, **1964**, 42, 1417; Lemieux, R.U., Faser-Reid, B., *Can. J. Chem.*, **1965**, 43, 1460; Lemieux, R.U., Morgan, A.R., *Can. J. Chem.*, **1965**, 43, 2190; Thiem, J., Karl, H., Schwentner, J., *Synthesis*, **1978**, 696; Thiem, J. Ossowski, P., *Carbohydr. Chem.*, **1984**, 3, 287; Thiem, J., Prahst, A., Wendt, T. *Liebigs Ann. Chem.*, **1986**, 1044; Thiem, J. in *Trends in Synthetic Carbohydrate Chemistry*, Horton, D., Hawkins, L.D., McGarvey, G.L., eds., ACS Symposium Series #386, American Chemical Society, Washington, D.C., **1989**, Chapter 8.)

The carbohydrate domains of the blood group substances contained in both glycoproteins and glycolipids are distributed in erythrocytes, epithelial cells and various secretions. The early focus on these systems centered on

their central role in determining blood group specificities. (R.R. Race and R. Sanger, *Blood Groups in Man*, 6th ed., Blackwell, Oxford, 1975) However, it is recognized that such determinants are broadly implicated in cell adhesion and binding phenomena. (For example, see M.L. Phillips, E. Nudelman, F.C.A. Gaeta, M. Perez, A.K. Singhal, S. Hakomori, J.C. Paulson, *Science*, 1990, 250, 1130.) Moreover, ensembles related to the blood group substances in conjugated form are encountered as markers for the onset of various tumors. (K.O. Lloyd, *Am. J. Clinical Path.*, 1987, 87, 129; K.O. Lloyd, *Cancer Biol.*, 1991, 2, 421) Carbohydrate-based tumor antigenic factors might find applications at the diagnostic level, as resources in drug delivery or ideally in immunotherapy. (Toyokuni, T., Dean, B., Cai, S., Boivin, D., Hakomori, S., and Singhal, A.K., *J. Am. Chem Soc.*, 1994, 116, 395; Dranoff, G., Jaffee, E., Lazenby, A., Golumbek, P., Levitsky, H., Brose, K., Jackson, V., Hamada, H., Paardoll, D., Mulligan, R., *Proc. Natl. Acad. Sci. USA*, 1993, 90, 3539; Tao, M-H., Levy, R., *Nature*, 1993, 362, 755; Boon, T., *Int. J. Cancer*, 1993, 54, 177; Livingston, P.O., *Curr. Opin. Immunol.*, 1992, 4, 624; Hakomori, S., *Annu. Rev. Immunol.*, 1984, 2, 103; K. Shigeta, et al., *J. Biol. Chem.*, 1987, 262, 1358)

The use of synthetic carbohydrate conjugates to elicit antibodies was first demonstrated by Gobel and Avery in 1929. (Goebel, W.F., and Avery, O.T., *J. Exp. Med.*, 1929, 50, 521; Avery, O.T., and Goebel, W.F., *J. Exp. Med.*, 1929, 50, 533.) Carbohydrates were linked to carrier proteins via the benzenediazonium glycosides. Immunization of rabbits with the synthetic antigens generated polyclonal antibodies. Other workers (Allen, P.Z., and Goldstein, I.J., *Biochemistry*, 1967, 6, 3029; Rude, E., and Delius, M.M., *Carbohydr. Res.*, 1968, 8, 219; Himmelsbach, K., et al., *Eur. J. Immunol.*, 1971, 1, 106; Fielder, R.J., et al., *J. Immunol.*, 1970, 105, 265)

developed similar techniques for conjugation of carbohydrates to protein carriers. Most of them suffered by introducing an antigenic determinant in the linker itself, resulting in generation of polyclonal antibodies.

5 Kabat (Arakatsu, Y., et al., *J. Immunol.*, 1966, 97, 858), and Gray (Gray, G.R., *Arch. Biochem. Biophys.* 1974, 163, 426) developed conjugation methods that relied on oxidative or reductive coupling, respectively, of free reducing oligosaccharides. The main disadvantage of

10 these techniques, however, is that the integrity of the reducing end of the oligosaccharide was compromised. In 1975 Lemieux described the use an 8-carbomethoxy-1-octanol linker (Lemieux, R.U., et al., *J. Am. Chem. Soc.*, 1975, 97, 4076) which alleviated the problem of linker

15 antigenicity and left the entire oligosaccharide intact. Equally effective in producing glycoconjugates was the allyl glycoside method described by Bernstein and Hall. (Bernstein, M.A., and Hall, L.D., *Carbohydr. Res.*, 1980, 78, C1.) In this technique the allyl glycoside of the

20 deblocked sugar is ozonized followed by a reductive workup. The resultant aldehyde is then reductively coupled to a protein carrier with sodium cyanoborohydride.

25 In the mid-70's and early 80's Lemieux and his collaborators made contributions to antibody production stimulated by synthetic glycoconjugates (Lemieux, R.U., et al., *J. Am. Chem. Soc.*, 1975, 97, 4076) and to conformational issues (Lemieux, R.U., et al., *Can. J.*

30 *Chem.*, 1979, 58, 631; Spohr, U., et al., *Can. J. Chem.*, 1985, 64, 2644; Vondonselaar, M., et al., *J. Biol. Chem.*, 1987, 262, 10848) important in the interactions of the blood group determinants (and analogues thereof) with the carbohydrate binding proteins known as lectins. More

35 recently, workers at Bristol-Meyers Squibb reported the X-ray crystal structure of the Lewis^x epitope complexed with the antibody BR96. (Jeffrey, P.D., et al., *Nature*

Structural Biol., 1995, 2, 466.) Two main components appear to govern recognition between carbohydrates and most antibodies. The first is multiple hydrogen bonding between the sugar hydroxyls and the amino acid residues of Asp, Asn, Glu, Gln, and Arg. The second major interaction is stacking between the sugar-ring faces and aromatic side chains, which occurs most frequently with tryptophan. In the complex with BR96 the most significant interactions involve the latter; additional hydrogen bonding occurs between the sugar hydroxyls and the indole nitrogens. Most antibody binding sites can support about 6 linear carbohydrate residues in a groove or cavity shaped binding site.

Glycoconjugates would be used, ideally, in direct immunotherapy or the monoclonal antibodies generated from vaccinations could be used to specifically target known chemotherapeutic agents to tumor sites. The immune response to carbohydrates is generally not strong, resulting mainly in production of IgM type antibodies. IgM antibodies are capable of complement fixation. Complement is a family of enzymes that can lyse cells to which antibodies are bound. The response to carbohydrate antigens normally does not enlist the use of T-cells which would aid in the body's rejection of the tumor. While the probability of complete tumor rejection as a result of vaccination with a conjugate is unlikely, such treatments will boost immune surveillance and recurrence of new tumor colonies can be reduced. (Dennis, J., *Oxford Glycosystems Glyconews Second*, 1992; Lloyd, K.O., in *Specific Immunotherapy of Cancer with Vaccines*, 1993, New York Academy of Sciences, 50-58.) Toyokuni and Singhal have described a synthetic glycoconjugate (Toyokuni, T., et al., *J. Am. Chem. Soc.*, 1994, 116, 395) that stimulated a measurable IgG titer, a result which is significant since an IgG response is generally associated with enlistment of helper T cells.

The use of immunoconjugates has shown promise in the reduction of large tumor masses. The workers at Bristol-Meyers Squibb (Trail, P.A., et al., *Science*, 1993, 261, 212) have described the attachment of the known
5 chemotherapeutic drug doxorubicin to the antibody BR96. BR96 is an anti-Lewis^Y antibody which has been shown to bind to human breast, lung and colon carcinomas. Athymic mice that have had human cancers (L2987-lung, RCA-colon, and MCF7-breast carcinomas) xenografted subcutaneously
10 were treated with the drug-antibody conjugate (BR96-DOX). The result was complete regression of the tumor mass in 78% of the mice treated. BR96 is efficiently internalized by cellular lysosomes and endosomes following attachment to the cell surface. The change in pH upon
15 internalization results in cleavage of the labile hydrazone thereby targeting the drug specifically to the desired site.

Many of the blood group determinant structures can also
20 occur in normal tissues. Antigen expression in normal cells and cancer cells can have subtle distributional differences. In the case of Le^Y, (which does appear in normal tissues) the expression of the determinant in tumor cells tends to be in the form of mucins which are
25 secreted. Mucins are glycoproteins with a high content of the amino acids serine and threonine. It is through the hydroxyl functionality of these amino acids that Lewis^Y is linked. Thus, in terms of generating competent antibodies against tumor cells expressing the Le^Y antigen
30 it is important that the antibody recognize the mucin structure.

Structurally, the blood group determinants fall into two basic categories known as type I and type II. Type I is
35 characterized by a backbone comprised of a galactose 1-3b linked to N-acetyl glucosamine while type II contains, instead, a 1-4b linkage between the same building blocks

- (cf. *N*-acetyl lactosamine). The position and extent of α -fucosylation of these backbone structures gives rise to the Lewis-type and H-type specificities. Thus, monofucosylation at the C₄-hydroxyl of the *N*-acetyl glucosamine (Type I series) constitutes the Le^a type, whereas fucosylation of the C₃-hydroxyl of this sugar (Type II series) constitutes the Le^x determinant. Additional fucosylation of Le^a and Le^x types at the C₂-hydroxyl of the galactose sector specifies the Le^b and Le^y types, respectively. The Le^y determinant is expressed in human colonic and liver adenocarcinomas. (Leverly, S.B., et al., *Carbohydr. Res.*, 1986, 151, 311; Kim, Y.S., *J. Cellular Biochem. Suppl.*, 16G 1992, 96; Kaizu, T., et al., *J. Biol. Chem.*, 1986, 261, 11254; Leverly, S.B., et al., *Carbohydr. Res.* 1986, 151, 311; Hakomori, S., et al., *J. Biol. Chem.*, 1984, 259, 4672; Fukushi, Y., et al., *ibid.*, 1984, 259, 4681; Fukushi, Y., et al., *ibid.*, 1984, 259, 10511.)
- 20 The presence of an α -monofucosyl branch, solely at the C₂-hydroxyl in the galactose moiety in the backbone, constitutes the H-type specificity (Types I and II). Further permutation of the H-types by substitution of α -linked galactose or α -linked *N*-acetylgalactosamine at its C₃-hydroxyl group provides the molecular basis of the familiar serological blood group classifications A, B, and O. (Lowe, J.B., The Molecular Basis of Blood Diseases, Stamatoyannopoulos, et. al., eds., W.B. Saunders Co., Philadelphia, PA, 1994, 293.)
- 30 Several issues merit consideration in contemplating the synthesis of such blood group substances and their neoglycoconjugates. For purposes of synthetic economy it would be helpful to gain relief from elaborate protecting group manipulations common to traditional syntheses of complex branched carbohydrates. Another issue involves fashioning a determinant linked to a protein carrier. It

is only in the context of such conjugates that the determinants are able to galvanize B-cell response and complement fixation. In crafting such constructs, it is beneficial to incorporate appropriate spacer units
5 between the carbohydrate determinant and the carrier. (Stroud, M.R., et al., *Biochemistry*, 1994, 33, 10672; Yuen, C.-T., et al., *J. Biochem.*, 1994, 269, 1595; Stroud, M.R., et al., *J. Biol. Chem.*, 1991, 266, 8439.)

10 The present invention provides new strategies and protocols for oligosaccharide synthesis. The object is to simplify such constructions such that relatively complex domains can be assembled with high stereo-
15 specificity. Major advances in glycoconjugate synthesis require the attainment of a high degree of convergence and relief from the burdens associated with the manipulation of blocking groups. Another requirement is that of delivering the carbohydrate determinant with appropriate provision for conjugation to carrier proteins
20 or lipids. (Bernstein, M.A., and Hall, L.D., *Carbohydr. Res.*, 1980, 78, C1; Lemieux, R.U., *Chem. Soc. Rev.*, 1978, 7, 423; R.U. Lemieux, et al., *J. Am. Chem. Soc.*, 1975, 97, 4076) This is a critical condition if the synthetically derived carbohydrates are to be
25 incorporated into carriers suitable for biological application.

Antigens which are selective or ideally specific for cancer cells could prove useful in fostering active
30 immunity. (Hakomori, S., *Cancer Res.*, 1985, 45, 2405-2414; Feizi, T., *Cancer Surveys*, 1985, 4, 245-269) Novel carbohydrate patterns are often presented by transformed cells as either cell surface glycoproteins or as membrane-anchored glycolipids. In principle, well chosen
35 synthetic glycoconjugates which stimulate antibody production could confer active immunity against cancers which present equivalent structure types on their cell

surfaces. (Dennis, J., *Oxford Glycosystems Glyconews* Second, 1992; Lloyd, K. O., in *Specific Immunotherapy of Cancer with Vaccines*, 1993, New York Academy of Sciences pp. 50-58) Chances for successful therapy improve with increasing restriction of the antigen to the target cell. A glycosphingolipid was isolated by Hakomori and collaborators from the breast cancer cell line MCF-7 and immunocharacterized by monoclonal antibody MB1. (Bremer, E. G., et al., *J. Biol. Chem.*, 1984, 259, 14773-14777; Menard, S., et al., *Cancer Res.*, 1983, 43, 1295-1300) The novel glycosphingolipid structure 1b (Figure 8) was proposed for this breast tumor-associated antigen on the basis of methylation and enzymatic degradation protocols. A ¹H NMR spectrum consistent with but not definitive for the proposed structure was obtained from trace amounts of isolated antigen. While individual sectors of the proposed structure were not unknown, the full structure was first described based on studies on the breast cancer line. It should be noted that MB1 also binds to normal human mammary gland tissue and ovarian cancer cell lines. Therefore, 1b as a total entity is likely not restricted to the transformed breast cells. Alternatively, smaller subsections of 1b are adequate for antibody recognition and binding. (The synthesis of the DEF fragment of 1b has been reported, and has been shown to bind to MB1: Lay, L., et al., *Helv. Chim. Acta*, 1994, 77, 509-514.)

The compounds prepared by processes described herein are antigens useful in adjuvant therapies as vaccines capable of inducing antibodies immunoreactive with epithelial carcinomas, for example, human colon, lung and ovarian tumors. Such adjuvant therapies have potential to reduce the rate of recurrence of cancer and increase survival rates after surgery. Clinical trials on 122 patients surgically treated for AJCC stage III melanoma who were treated with vaccines prepared from melanoma differen-

5 tiation antigen GM2 (another tumor antigen which like
MBrl is a cell surface carbohydrate) demonstrated in
patients (lacking the antibody prior to immunization) a
highly significant increase in disease-free interval
(P.O. Livingston, et al., J. Clin Oncol., 12, 1036
(1994)).

10 The present invention provides a method of synthesizing
Le^Y-related antigens as well as artificial protein-
conjugates of the oligosaccharide which might be more
immunogenic than the smaller glycolipid. The antigen
contains a novel array of features including the α -
linkage between the B and the C entities, as well as the
15 β -linked ring D gal-NAC residue. (For the synthesis of
a related structure (SSEA-3) which lacks the fucose
residue see: Nunomura, S.; Ogawa, T., *Tetrahedron Lett.*,
1988, 29, 5681-5684.) The present invention also
provides a total synthesis of 1b, rigorous proof that the
Hakomori antigen does, in fact, correspond to 1b and the
20 synthesis of a bioconjugatable version of 1b. The
conciseness of the synthesis reflects the efficiency of
glycal assembly methods augmented by a powerful method
for sulfonamidoglycosylation (see, e.g., the
transformation of 14b-15b, Figure 10).

25

Brief Description of the Figures

Figure 1 shows glycal assembly leading to neoglycoproteins.

5 Figure 2 shows the synthesis of 4a. Reagents: a) TBDPSCl, imidazole/DMF 84%; b) carbonyldiimidazole, cat. imidazole, THF (65%) c) 5a, di-tert-butylpyridine, AgClO₄, SnCl₂, ether (51%); PhSO₂NH₂, I(sym-coll)₂ClO₄ (94%).

10 Figure 3 shows the synthesis of 8a. Reagents: a) 9a, AgBF₄, 4A mol. sieves, THF (75%); b) i. TBAF, THF; ii. Na/NH₃; iii Ac₂O, pyr. c) i. 3,3-dimethioxirane; allyl alcohol, ZnCl₂ (72%); ii. NaOMe, MeOH (quant.).

15 Figure 4 shows a strategy for the solid-phase of oligosaccharides using the glycal assembly method.

Figure 5 shows the application of the solid-support method to the assembly of 1,2-branching patterns of
20 complex carbohydrates.

Figure 6 shows the synthesis of a tetrasaccharide having H-type 2 blood group specificity. Reagents: (a) 1. 3,3-dimethyldioxirane, CH₂Cl₂; 2. 8, ZnCl₂, THF; (b) 10, 25 Sn(OTf)₂, di-tert-butylpyridine, THF; (c) TBAF, AcOH, THF; (d) TIPSCl, imidazole, DMF; (e) I(coll)₂ClO₄, PhSO₂NH₂, CH₂Cl₂; (f) 15, AgBF₄, 4A M.S., THF; (g) 1. TBAF, AcOH, THF; 2. Na/NH₃; 3. Ac₂O, pyridine.

30 Figure 7a and 7b show the synthesis of a Le^b hexa-saccharide in bioconjugatable form. Reagents: (a) 1. 3,3-dimethyldioxirane, CH₂Cl₂; 2. 19, ZnCl₂, THF; (b) 10, Sn(OTf)₂ di-tert-butylpyridine, THF; (c) TBAF, AcOH, THF; (d) TIPSCl, imidazole, DMF; (e) I(coll)₂ClO₄, PhSO₂NH₂, 35 CH₂Cl₂; (f) 24, AgBF₄, 4A M.S., THF; (g) 1. TBAF, AcOH, THF; 2. Na/NH₃; 3. Ac₂O, pyridine; (h) 1. 3,3-dimethyl-dioxirane, CH₂Cl₂; 2. allyl alcohol, ZnCl₂; 3. NaOMe, MeOH.

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Figure 8 shows the structure of the MBrl antigen and a reaction pathway to a trisaccharide intermediate.

Reagents: a. n-Bu₂SnO, PMBCl, TBABr, PhH, 70%; b. NaH, BnBr, DMF, 95%; c. (i) 3,3-dimethyldioxirane, CH₂Cl₂; (ii) TBAF, THF; (iii) NaH, BnBr, DMF, 40% (three steps); d. NaH, BnBr, DMF, 80%; e. (i) TBAF, THF; (ii) NaOMe, MeOH, 93% (two steps); f. (n-Bu₃Sn)₂O, BnBr, TBABr, PhH, 90%; g. SnCl₂, AgClO₄, 2,6-di-butylpyridine, 4 Å mol. sieves, Et₂O, 40% & (4.5:1 &:B); h. DDQ, CH₂Cl₂, H₂O, 84%.

Figure 9 shows a reaction pathway to a trisaccharide intermediate.

Reagents: a. (i) 3,3-dimethyldioxirane, CH₂Cl₂; (ii) 10a, ZnCl₂, THF, 87%; b. SnCl₂, AgClO₄, Et₂O, 47%; c. I(coll)₂ClO₄, PhSO₂NH₂, 4 Å mol. sieves, 47%.

Figure 10(a) shows a reaction pathway to the hexasaccharide MBrl antigen.

Reagents: a. EtSH, LiHMDS, DMF, 75%. B. 8b (0.5 equiv), MeOTf, 4 Å Mol. sieves, 70-85% B, (10:1 B &); c. (i) 3,3-dimethyldioxirane, CH₂Cl₂ (ii) 17b (5 equiv), Zn(OTf)₂, 20%; d. Ac₂O, Et₃N, DMAP, CH₂Cl₂ 95%; e. Lindlar's cat., H₂ palmitic anhydride, EtOAc, 90%; f. (i) TBAF, THF; (ii) NaOMe, MeOH, 94%; g. (i) Na, NH₃, THF; (ii) Ac₂O, Et₃N, DMAP, CH₂Cl₂, 80% h. NaOMe, MeOH, quant.

Figure 10(b) shows a reaction pathway to the allyl glycoside.

Reagents: a. TBAF, THF, 94%; b. (i) Na, NH₃, THF; (ii) Ac₂O, Et₃N, DMAP, THF, DMF, 85%; c. (i) 3,3-dimethyldioxirane, CH₂Cl₂, (ii) allyl alcohol, 65% (+ 29% of α-manno isomer); d. NaOMe, MeOH, quant.

Figure 11 shows a reaction pathway to intermediates for preparing the hexasaccharide antigen MBrl.

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Figure 12 shows a reaction pathway to the hexasaccharide antigen MBr1 by a 4+2 synthetic approach.

5 Figure 13(a) shows the proposed mode of action for inflammatory response.

Figure 13(b) shows the structure of SLe^x.

10 Figure 13(c) shows the structure of sulfated E-selectin ligands.

Figure 14(a) shows a reaction pathway to prepare the Lubineau sulfated Le^a.

15 Figure 14(b) shows a reaction pathway to prepare the Nicolau sulfated Le^a.

20 Figure 15(a) shows compounds 17d, 18d and sulfated Le^a 19d.

Figure 15(b) shows the preparation of sulfated Le^x glycal 21d.

25 Figure 15(c) shows the preparation of trisaccharide intermediates 26d and 27d.

Figure 16(a) shows the preparation of disaccharide intermediates 30d and 31d.

30 Figure 16(b) shows the preparation of trisaccharide Le^a glycal 32d.

35 Figure 16(c) shows the preparation of trisaccharide intermediate 35d.

Figure 17 shows the preparation of sulfated trisaccharide 19d.

Figure 18(a) shows the preparation of TIPS- and TBDPS-protected tetrasaccharides 22c, 23c and 24c.

5 Figure 18(b) shows the preparation of TBDPS-protected tetrasaccharide intermediate 25c.

Figure 18(c) shows the preparation of TBDPS-protected tetrasaccharide ceramide intermediate 28c.

10 Figure 18(d) shows a model reduction reaction to prepare tetrasaccharide ceramides.

15 Figure 19(a) shows a reduction reaction to prepare tetrasaccharide ceramide intermediate 31c.

Figure 19(b) shows a reaction pathway to prepare tetrasaccharide ceramide 32c.

20 Figure 20(a) shows the reactivity of compound 17c with αLe^Y (s193) and αLe^b (T218) control.

Figure 20(b) shows the the enzyme-linked immunosorbant assay used to measure antibody titer.

25 Figure 21(a) shows in panels A, B and C the measured titers of total antibody Ig, IgM and IgG type antibodies, respectively, in five mice immunized with conjugate 17c; in panels D, E and F controls where mice were immunized with BSA carrier alone.

30 Figure 21(b) shows in panels G, H and I the measured titer of Ig, IgM and IgG type antibodies with the Le^Y mucin structure as the test antigen, respectively, mice immunized with conjugate 17c; in panels J, K and L show
35 results of immunization with ceramide conjugate 32b.

Figure 22(a) shows the preparation of tetrasaccharide intermediate 11c.

5 Figure 22(b) shows the preparation of O-allyl pentasaccharide 15c.

10 Figure 22(c) shows the preparation of a Le^y-BSA glycoconjugate 17c by reductive amination of pentasaccharide aldehyde intermediate 16c.

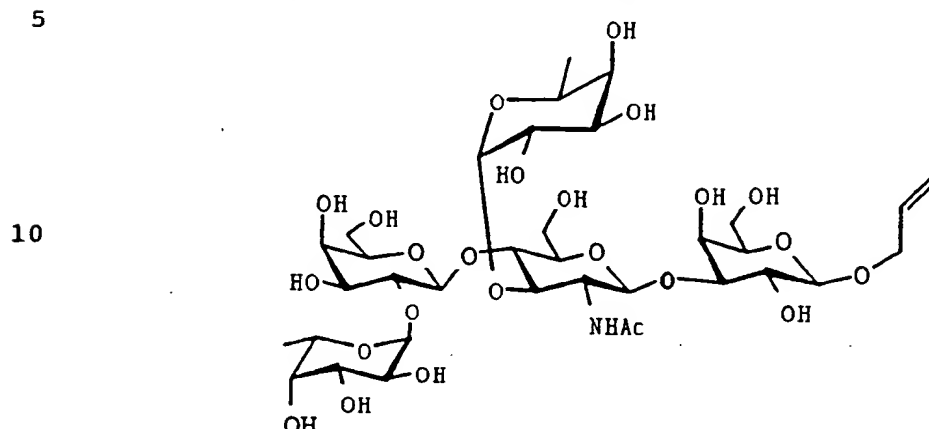
Figure 23(a) shows the preparation of protected hexasaccharide glycal 19c.

15 Figure 23(b) shows three methods of elaborating tetrasaccharide iodosulfonamide 11c.

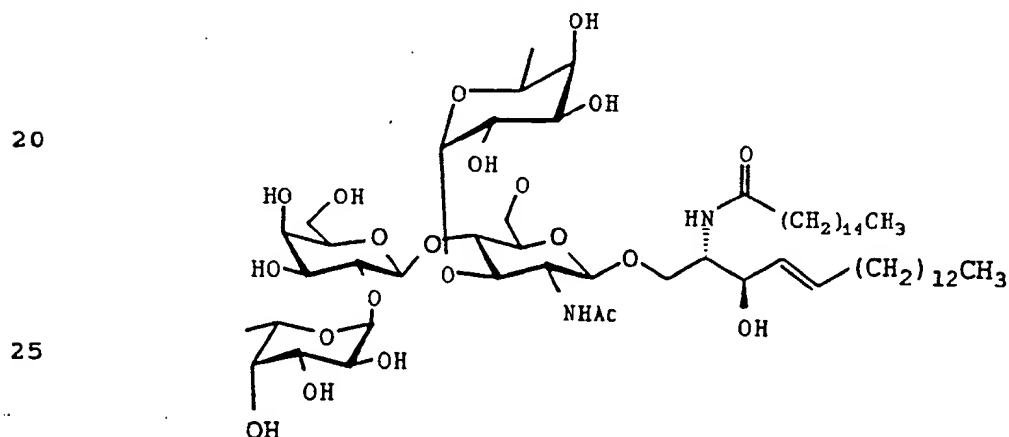
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Summary of the Invention

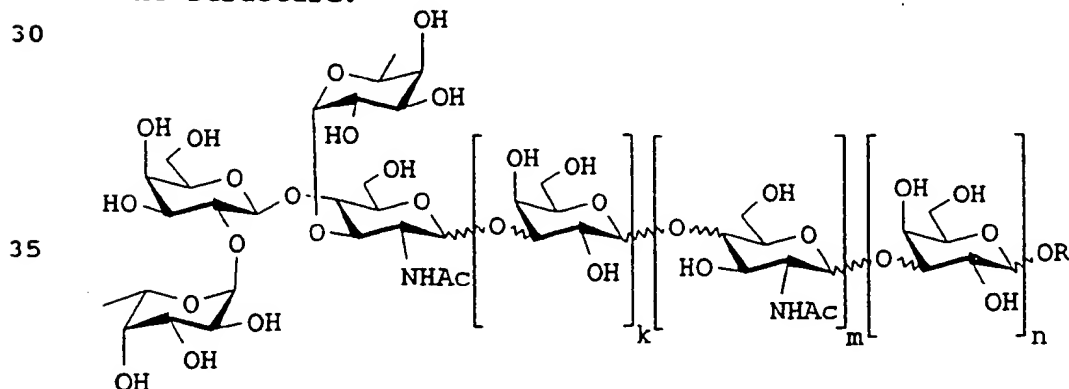
The present invention provides a method of synthesizing an allyl pentasaccharide having the structure:



The present invention also provides a method of synthesizing a ceramide having the structure:



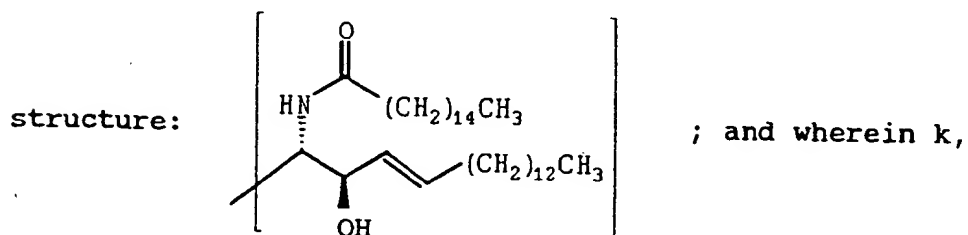
The present invention further provides a compound having the structure:



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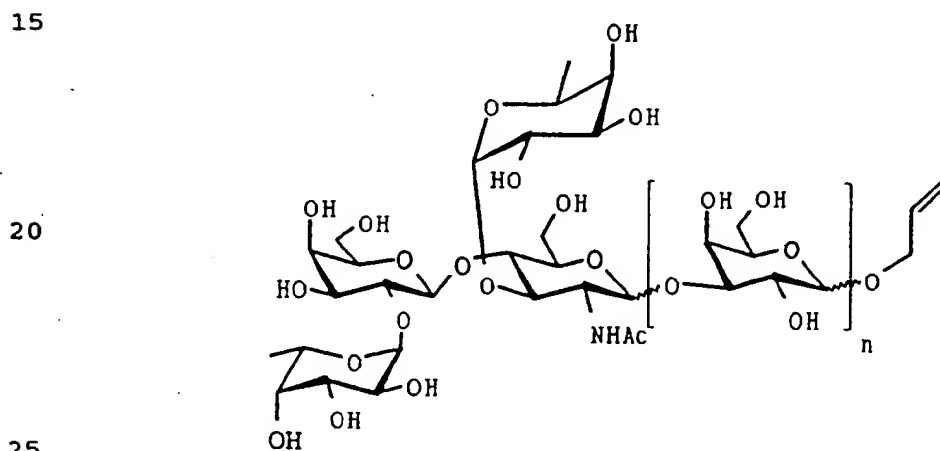
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wherein R is H, substituted or unsubstituted alkyl, aryl or allyl, or an amino acyl moiety, an amino acyl residue of a peptide, an amino acyl residue of a protein, which amino acyl moiety or residue bears an ω -amino group or an ω -(C=O)- group, which group is linked to O via a polymethylene chain having the structure $-(CH_2)_r-$, where r is an integer between about 1 and 9, or a moiety having the



m and n are independently 0, 1, 2 or 3.

The present invention also provides a compound having the structure:

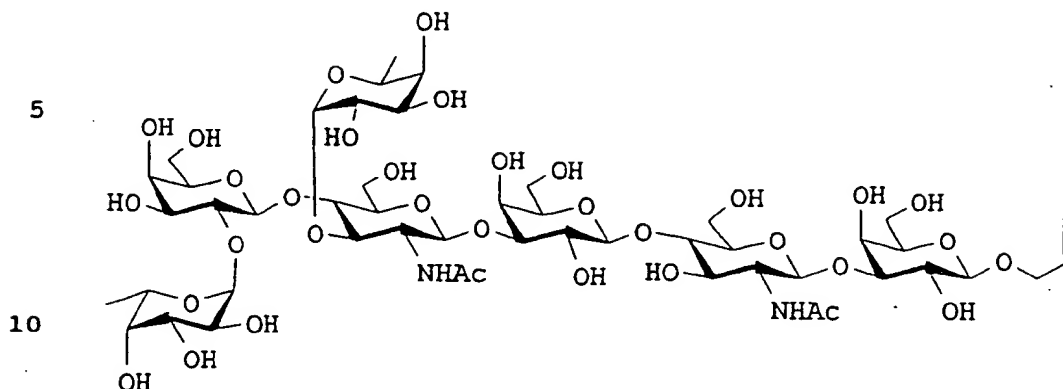


wherein n is 0, 1, 2, 3 or 4.

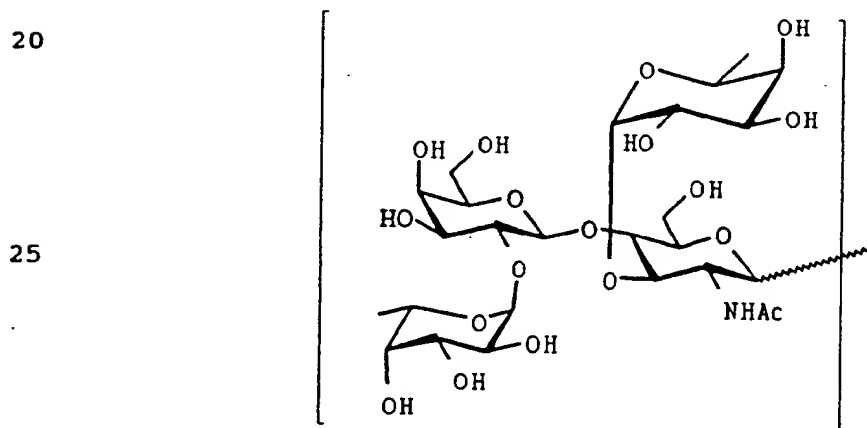
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The present invention also provides a compound having the structure:



15 In addition, the present invention provides a method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with epithelial tumor cells, which comprises administering to the subject an amount of a compound which contains a Le^x tetrasaccharide determinant having the structure:



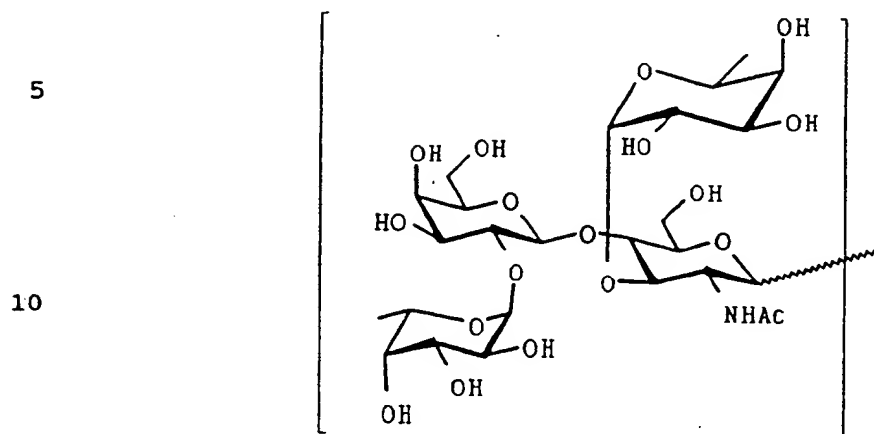
30 effective to induce the antibodies.

The present invention provides a method of preventing recurrence of epithelial cancer in a subject which comprises vaccinating the subject with a compound which

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contains a Le^x tetrasaccharide determinant having the structure:



effective to induce the antibodies.

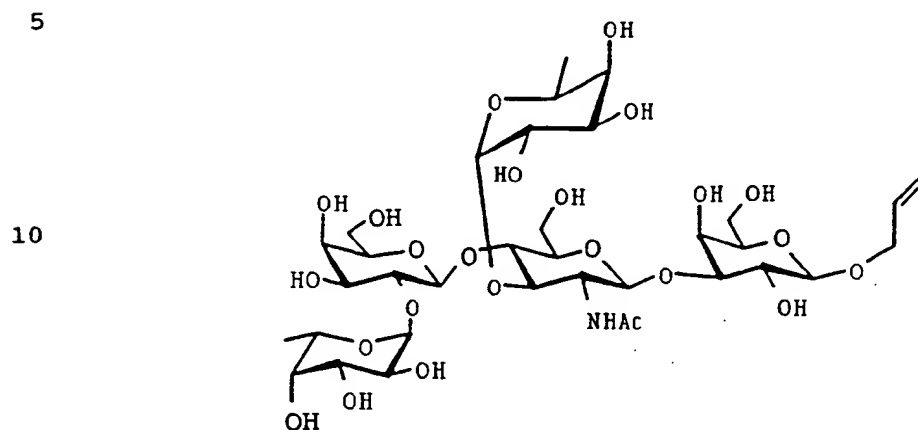
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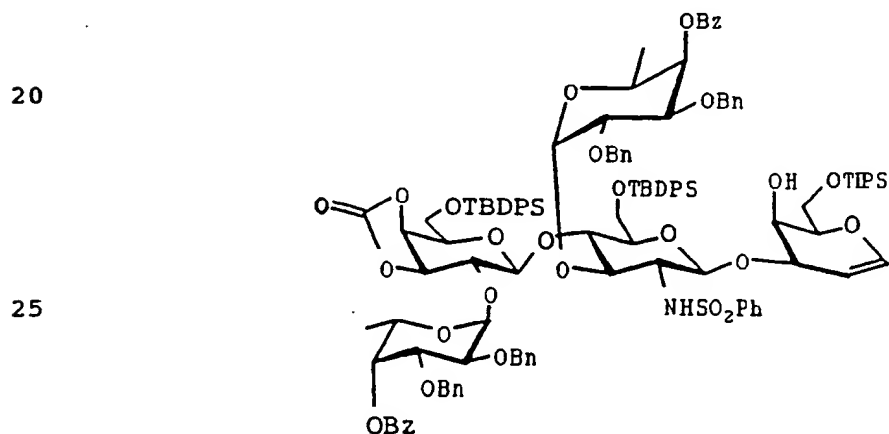
Detailed Description of the Invention

The present invention provides a method of synthesizing an allyl pentasaccharide having the structure:



which comprises:

(a) (i) de-silylating a compound having the structure:

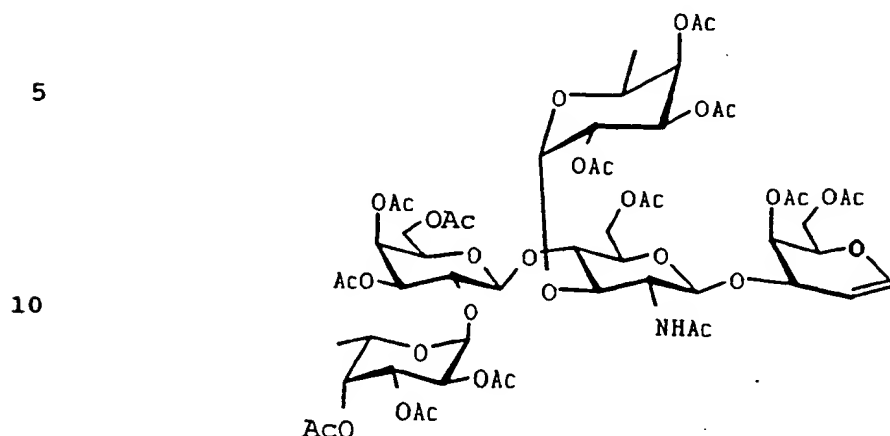


30 with $R^1R^2R^3R^4NF$ wherein R^1 , R^2 , R^3 , and R^4 are independently a linear or branched chain alkyl, aralkyl or aryl, to form an N-sulfonamide pentasaccharide; (ii) cleaving the N-sulfonamide pentasaccharide formed in step (a)(i) to form a deprotected pentasaccharide; and (iii) acetylating

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the deprotected pentasaccharide formed in step (b) (ii) to form a peracetate having the structure:



(b) (i) treating the peracetate formed in step (a) (iii) with an epoxidizing agent to form an epoxide peracetate; (ii) reacting the epoxide peracetate formed in step (b) (i) with allyl alcohol to form an allylglycoside peracetate; and (iii) cleaving the allylglycoside peracetate with an alkoxide salt to form the allyl pentasaccharide. In one embodiment, the present invention provides the method wherein R^1 , R^2 , R^3 , and R^4 in step (a) (i) are n-butyl. In another embodiment, the present invention provides a method wherein the cleaving step (a) (ii) is performed with Na/NH_3 . In another embodiment, the present invention provides a method wherein the epoxidizing agent of step (b) (i) is 3,3-dimethyldioxirane. In another embodiment, the present invention provides a method wherein the alkoxide salt of step (b) (iii) is NaOMe.

Step (a) (i) may be carried out using a fluoride salt such as tetra-n-butylammonium fluoride in a suitable nonaqueous dipolar solvent, such as THF. Cleaving step (a) (ii) may be effected using a reducing metal in liquid ammonia with a proton donor such as methanol or ethanol. Peracetylation step (a) (iii) is performed using acetyl chloride or acetic anhydride in the presence of an organic

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base such as pyridine. Epoxidation step (b)(i) is effected using an epoxidizing agent such as peracetic acid, m-chloroperbenzoic acid or trifluoroacetic acid, but preferably with 3,3-dimethyldioxirane. Ring-opening

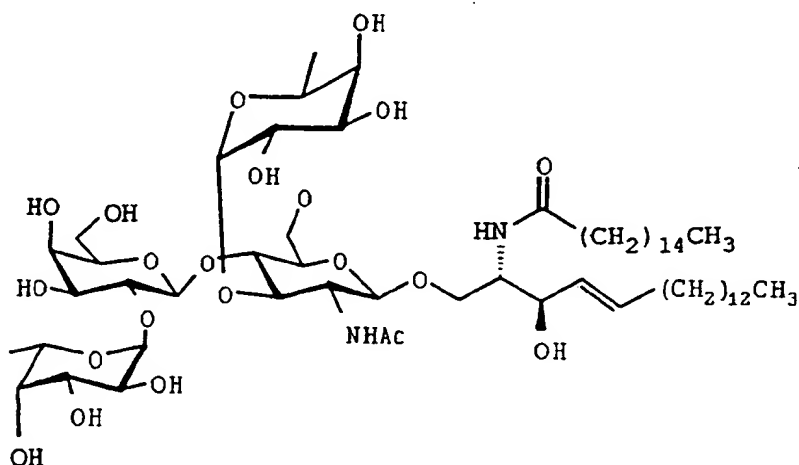
5 step (b)(ii) is carried out with allyl alcohol in the presence of a Lewis acid catalyst such as ZnCl_2 . Saponification step (b)(iii) is effected using a metal alkoxide such as sodium, lithium or potassium methoxide or ethoxide in the presence of the corresponding alcohol.

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The present invention also provides a method of synthesizing a ceramide having the structure:

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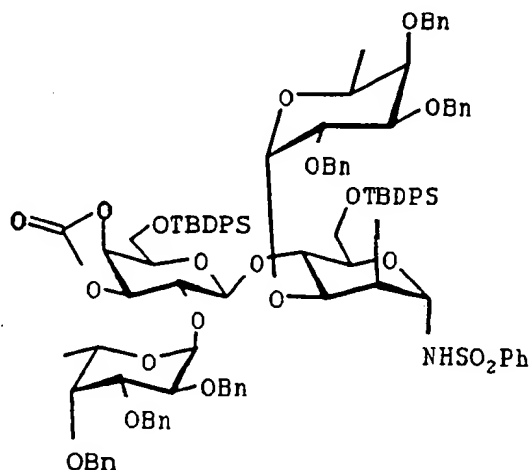
25

which comprises:

(a) treating a compound having the structure:

30

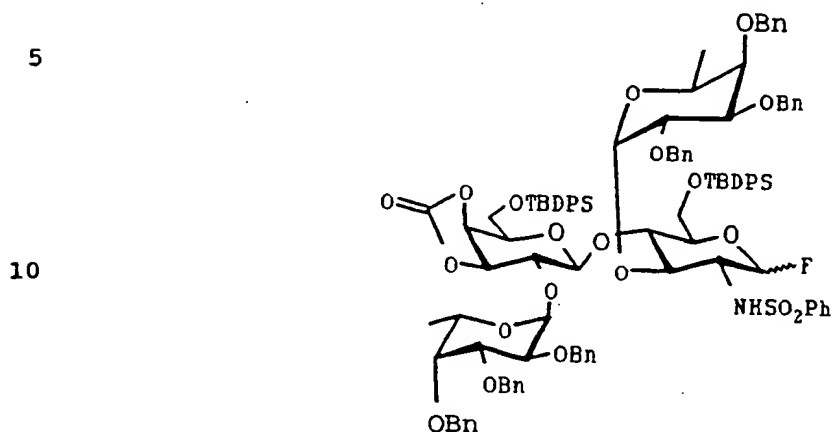
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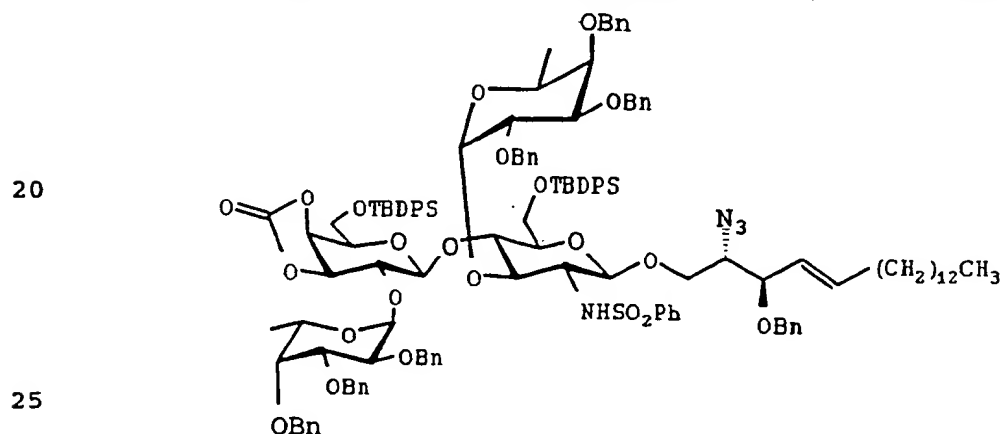
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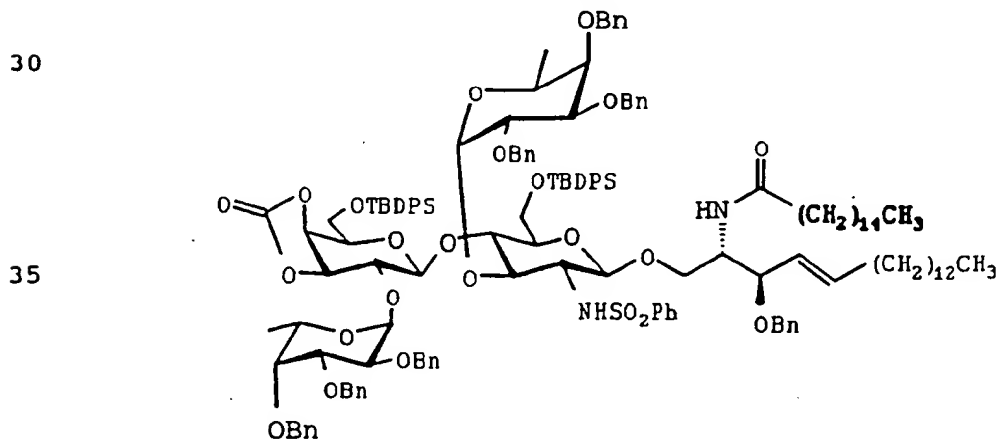
with aqueous silver(I) followed by reacting with diethylaminosulfur trifluoride to form a compound having the structure:



(b) treating the compound formed in step (a) with azidosphingosine to form a compound having the structure:



(c) reducing the compound formed in step (b) to form a compound having the structure:



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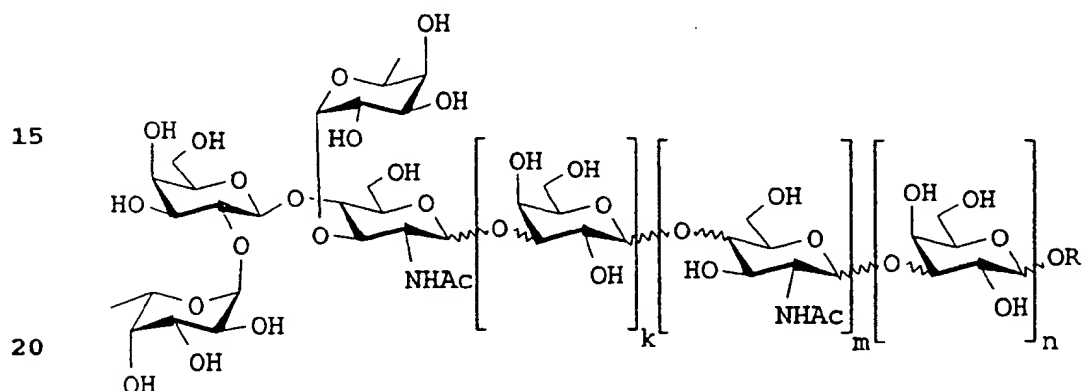
and (d) (i) de-silylating the compound formed in step (c) with $R^1R^2R^3R^4NF$ wherein R^1 , R^2 , R^3 , and R^4 are independently a linear or branched chain alkyl, aralkyl or aryl; (ii) reductively cleaving the compound formed in
5 step (d)(i) to form a polyalcohol tetrasaccharide; (iii) peracetylating the polyalcohol tetrasaccharide to form a peracetate tetrasaccharide; and (iv) saponifying the peracetate tetrasaccharide with a metal alkoxide to form the ceramide. In one embodiment, the present invention
10 provides a method wherein silver(I) in step (a) is silver carbonate. In another embodiment, the present invention provides a method wherein step (b) is performed in the presence of zirconocene dichloride and silver triflate. In another embodiment, the present invention provides a
15 method wherein the reducing step (c) is performed using hydrogen gas and palmitic anhydride in the presence of Lindlar's catalyst. In yet another embodiment, the present invention provides a method wherein R^1 , R^2 , R^3 , and R^4 in step (d)(i) are n-butyl and step (d)(ii) is performed using Na/NH_3 .
20

Treating step (a) is effected using a silver salt, such as silver carbonate, in the presence of a nonnucleophilic base, such as triethylamine, in a mixed aqueous and
25 dipolar solvent such as THF. After aqueous solvents are removed, the mixture is treated in situ at low temperatures, between -60° and 0° , preferably at about -30° , with diethylaminosulfur trifluoride (DAST) under suitable conditions. Coupling step (b) is carried out
30 using a mixed metal system such as Cp_2ZrCl_2 and silver triflate in an organic solvent such as dichloromethane. Reducing step (c) is effected using a noble metal catalyst such as Lindlar's catalyst and a hydrogen atmosphere at ambient to 100 psi. De-silylating step
35 (d)(i) may be carried out using a fluoride salt such as tetra-n-butylammonium fluoride in a suitable nonaqueous dipolar solvent, such as THF. Cleaving step (d)(ii) may

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be effected using a reducing metal in liquid ammonia with a proton donor such as methanol or ethanol. Peracetylation step (d)(iii) is performed using acetyl chloride or acetic anhydride in the presence of an organic base such as pyridine. Saponification step (d)(iv) is effected using a metal alkoxide such as sodium, lithium or potassium methoxide or ethoxide in the presence of the corresponding alcohol.

The present invention provides a compound having the structure:

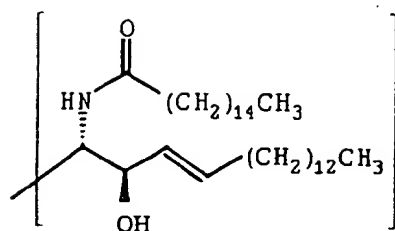


wherein R is H, substituted or unsubstituted alkyl, aryl or allyl, or an amino acyl moiety, an amino acyl residue of a peptide, an amino acyl residue of a protein, which amino acyl moiety or residue bears an ω -amino group or an ω -(C=O)- group, which group is linked to O via a polymethylene chain having the structure $-(CH_2)_r-$, where r is an integer between about 1 and 9, or a moiety having the

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structure:



; and wherein k,

m and n are independently 0, 1, 2 or 3.

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The chemical structure shows a branched oligosaccharide. It consists of a central chain of glucose units linked by 1-4 glycosidic bonds. The chain is branched at the second glucose unit from the left, which is linked to a side chain of two more glucose units. The repeating units are labeled k, m, and n. The unit k is a glucose unit with an NHAc group at the 2-position. The unit m is a glucose unit with an NHAc group at the 2-position. The unit n is a glucose unit with an NHAc group at the 2-position. The chain ends with a long-chain fatty acid derivative, specifically 1-(2-hydroxy-3-(12-hydroxydodecyl)propyl)pyrrolidine-2-carboxylic acid, which is linked to the terminal glucose unit via its hydroxyl group.

15

20

Chemical structure of compound 15, showing a complex glycoside. The structure includes a glucose unit linked to a mannose unit, which is further linked to a chiral auxiliary. The auxiliary is a 1,2-diol derivative of a long-chain alcohol, specifically $\text{NHCO}(\text{CH}_2)_{14}\text{CH}_3$ and $(\text{CH}_2)_{12}\text{CH}_3$. The glucose unit has hydroxyl groups at C2, C3, and C6. The mannose unit has hydroxyl groups at C2, C3, and C6. The auxiliary has a hydroxyl group at C2 and a long alkyl chain at C1.

30

35

The chemical structure shows a branched polysaccharide. It features a central chain of glucose units in Haworth projection. One unit has an 'NHAc' group at the C2 position. A branch is attached via an oxygen atom to a glucose unit. The main chain continues with a repeating unit in brackets, subscripted with 'n'. The chain terminates in a vinyl group (-CH=CH₂). Various hydroxyl groups are shown at different positions on the rings.

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wherein n is 0, 1, 2, 3 or 4. In one embodiment, the present invention provides a compound wherein n is 1. In another embodiment, the present invention provides a compound wherein n is 2.

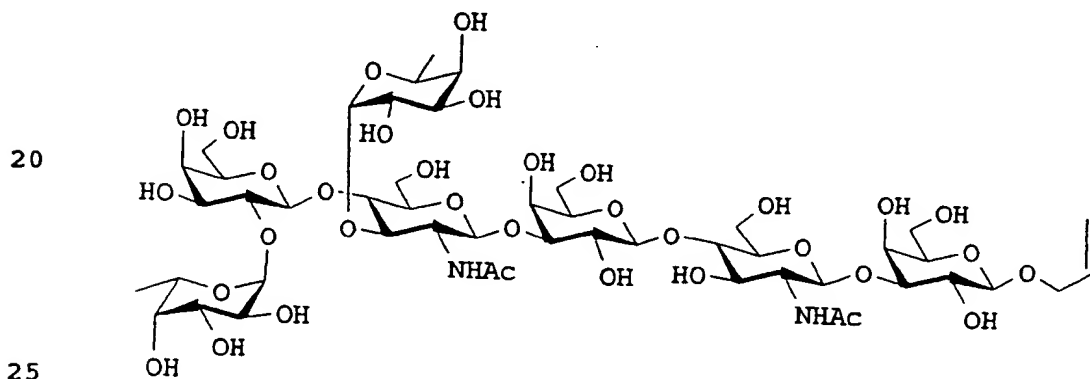
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In general, each O-allyl Le^x containing oligosaccharide may be linked to a carrier protein by a two-step process. Ozonolysis affords an aldehyde which is then reductively aminated by the free surface ϵ -amines of the carrier protein, using a reducing agent such as sodium cyanoborohydride. The product is a Le^x-carrier protein adduct useful for inducing antibodies as disclosed herein.

10

The present invention also provides a compound having the structure:

15



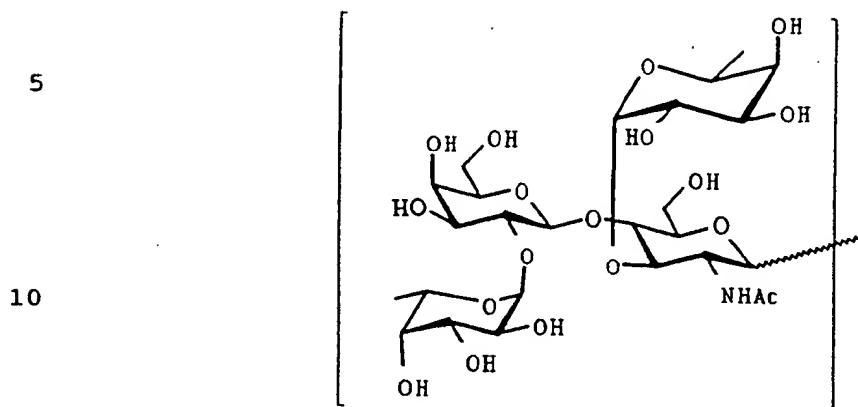
The present invention provides several therapeutic uses for the compounds disclosed herein. Accordingly, the present invention provides a method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with epithelial tumor cells, which comprises administering to the subject an

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amount of a compound which contains a Le^x tetrasaccharide determinant having the structure:



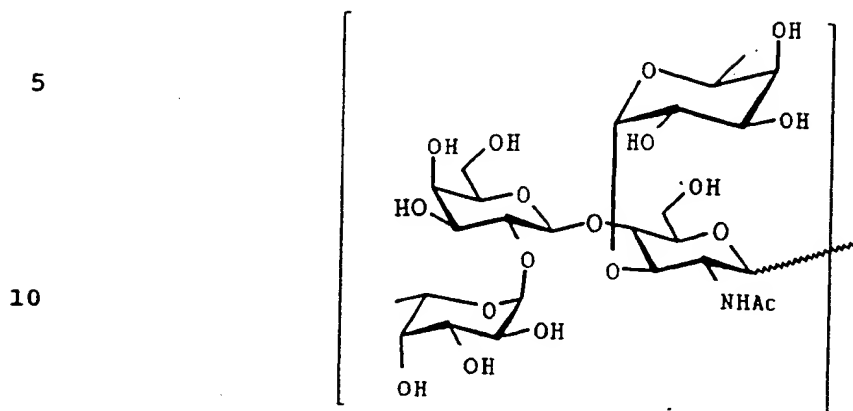
effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the compound is bound to a suitable carrier protein. In a certain embodiment, the present invention provides a method wherein the carrier protein is bovine serum albumin, polylysine, or KLH. In another certain embodiment, the present invention provides a method which further comprises coadministering an immunological adjuvant. In another embodiment, the present invention provides a method wherein the adjuvant is bacteria or liposomes. Specifically, the invention provides a method wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21. In various embodiments, the present invention may be practiced using any of the compounds disclosed hereinabove. In a further embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.

35 The present invention also provides a method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with colon tumor cells, which comprises administering to the subject an amount of

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a compound which contains a Le^x tetrasaccharide determinant having the structure:



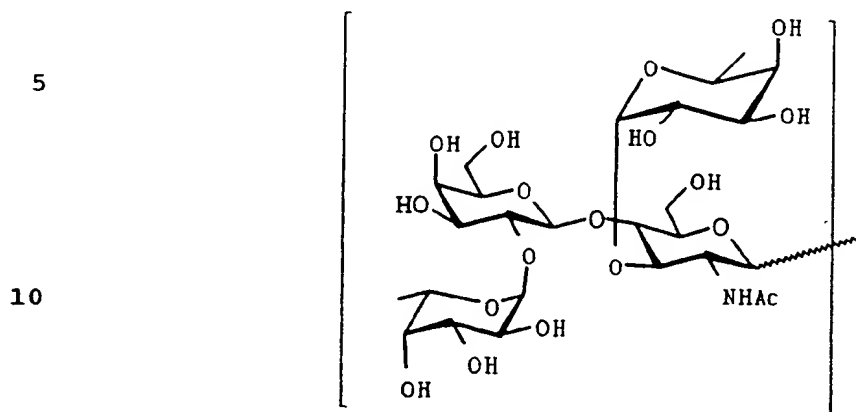
effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the compound is bound to a suitable carrier protein. In a certain embodiment, the present invention provides a method wherein the carrier protein is bovine serum albumin, polylysine, or KLH. In another certain embodiment, the present invention provides a method which further comprises coadministering an immunological adjuvant. In another embodiment, the present invention provides a method wherein the adjuvant is bacteria or liposomes. Specifically, the invention provides a method wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21. In various embodiments, the present invention may be practiced using any of the compounds disclosed hereinabove. In a further embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.

The present further provides a method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with ovarian tumor cells, which comprises administering to the subject an amount of

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a compound which contains a Le^x tetrasaccharide determinant having the structure:



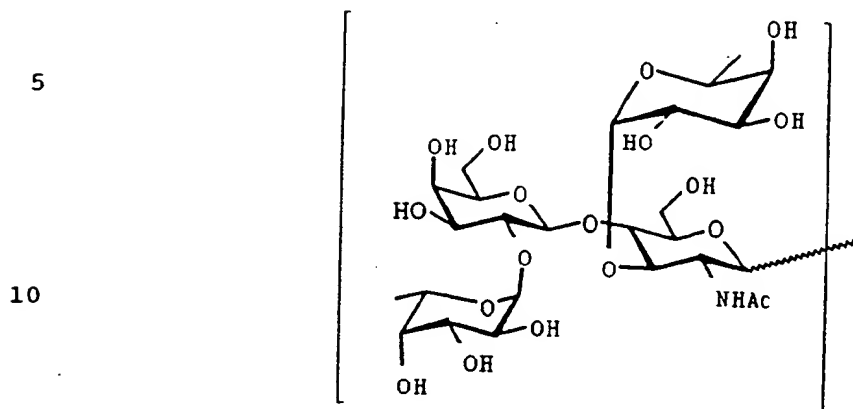
effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the compound is bound to a suitable carrier protein. In a certain embodiment, the present invention provides a method wherein the carrier protein is bovine serum albumin, polylysine, or KLH. In another certain embodiment, the present invention provides a method which further comprises coadministering an immunological adjuvant. In another embodiment, the present invention provides a method wherein the adjuvant is bacteria or liposomes. Specifically, the invention provides a method wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21. In various embodiments, the present invention may be practiced using any of the compounds disclosed hereinabove. In a further embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.

The present invention provides a method of preventing recurrence of epithelial cancer in a subject which comprises vaccinating the subject with a compound which

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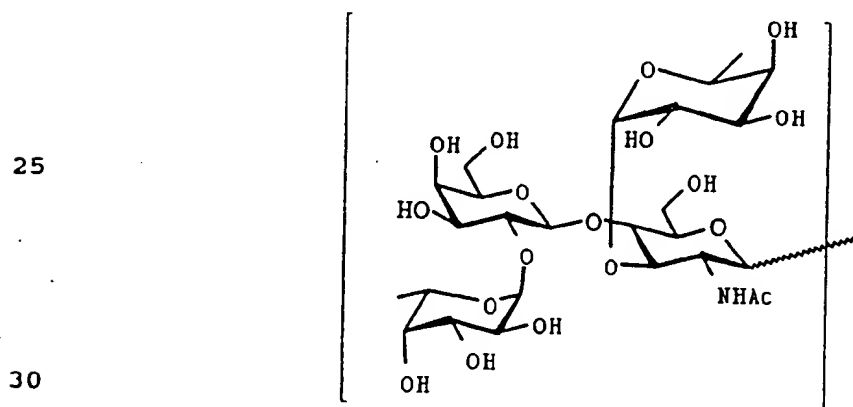
contains a Le^x tetrasaccharide determinant having the structure:



effective to induce the antibodies.

15 In particular, the present invention provides a method of preventing recurrence of colon cancer in a subject which comprises vaccinating the subject with a compound which contains a Le^x tetrasaccharide determinant having the structure:

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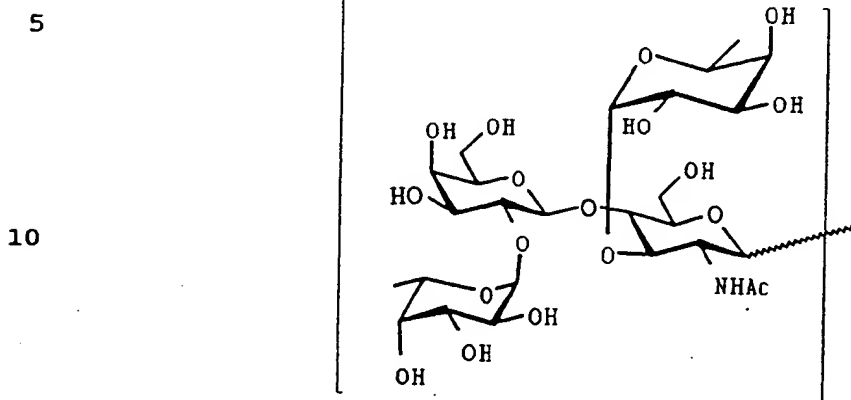
effective to induce the antibodies. In various embodiments, the present invention may be practiced using any of the compounds disclosed hereinabove.

35 The present invention also provides a method of preventing recurrence of ovarian cancer in a subject

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which comprises vaccinating the subject with a compound which contains a Le^x tetrasaccharide determinant having the structure:

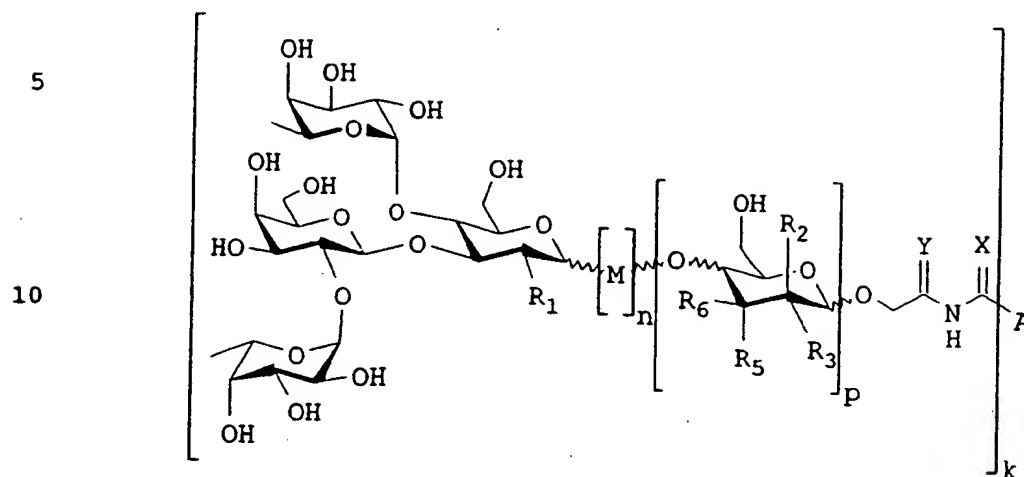


effective to induce the antibodies. In various embodiments, the present invention provides a method of treating epithelial tumors, including colon, lung, ovarian, and prostate, wherein the compound is bound to a suitable carrier protein. In various embodiments, the present invention provides a method wherein the carrier protein is bovine serum albumin, polylysine, or KLH. In other embodiments, the present invention provides a method which further comprises coadministering an immunological adjuvant. In certain embodiments, the present invention provides a method wherein the adjuvant is bacteria or liposomes. In specific embodiments, the present invention provides a method wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21. In certain embodiments, the present invention may be practiced using any of the compounds disclosed hereinabove.

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The present invention provides a compound having the structure:



wherein A is selected from the group consisting of (i) an amino acid bearing an ω -amino group or an ω -(C=O)- group, (ii) an amino acid residue of a peptide, which residue bears an ω -amino group or an ω -(C=O)- group, and (iii) an amino acid residue of a protein, which residue bears an ω -amino group or an ω -(C=O)- group; wherein R_1 is H, OH, NH_2 , or NHR_4 , where R_4 is SO_2Ph , a linear or branched chain alkyl or acyl group, or an aryl group; wherein M has the structure:



35 wherein n is an integer from 0 to 18, and where n is greater than 1, each M is independently the same or different; wherein p is either 0 or 1; wherein R_2 , R_3 , R_5

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and R_6 are independently the same or different, and are H or OH, with the proviso that geminal R_2 and R_3 are not both OH, and geminal R_5 and R_6 are not both OH; wherein each wavy line between a carbon atom and an oxygen atom denotes an R or S configuration at the carbon atom; wherein X and Y are independently the same or different and are H_2 or O; and wherein k is an integer greater than or equal to 1, with the proviso that when A is an amino acid bearing an ω -amino group or an ω -(C=O)- group, k is equal to 1.

In one embodiment, the present invention provides the compound disclosed hereinabove wherein A is lysine or a lysine residue.

In another embodiment, the present invention provides the compound disclosed hereinabove wherein A is glutamic acid or a glutamic acid residue.

In another embodiment, the present invention provides the compound disclosed hereinabove wherein A is aspartic acid or an aspartic acid residue.

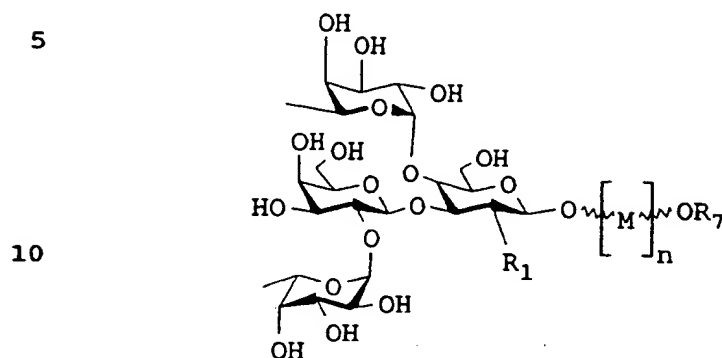
The invention also provides the compound disclosed hereinabove wherein A is an amino acid residue of a globular protein. In one embodiment, the invention provides the compound wherein the globular protein is selected from the group consisting of bovine serum albumin and human serum albumin.

In one embodiment, the invention provides the compound disclosed hereinabove wherein k is 1.

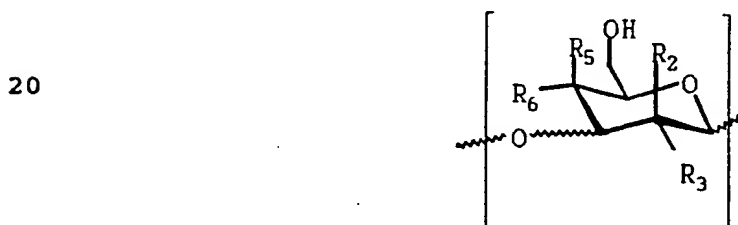
In another embodiment, the invention provides the compound disclosed hereinabove wherein n and p are both equal to 0.

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The invention provides a compound having the structure:



15 wherein R_1 is H, OH, NH_2 or NHR_4 , where R_4 is SO_2Ph , a linear or branched chain alkyl or acyl group, or an aryl group; wherein M has the structure:



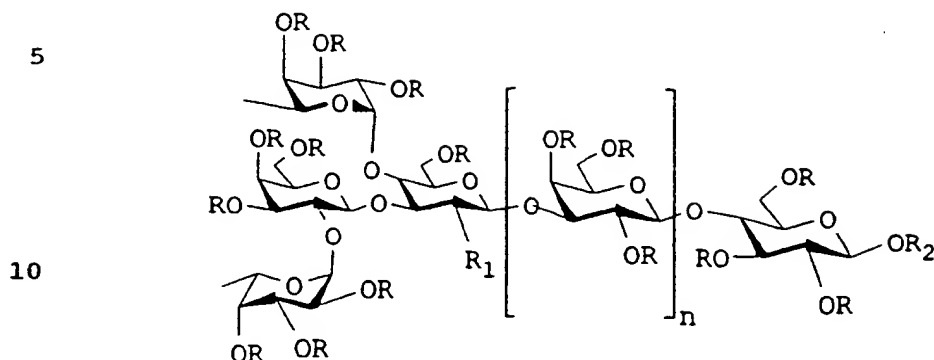
25 wherein n is an integer from 0 to 18, and where n is greater than 1, each M is independently the same or different; wherein R_2 , R_3 , R_4 and R_6 are independently the same or different and are H or OH, with the proviso that geminal R_2 and R_3 are not both OH, and geminal R_4 and R_6 are not both OH; wherein each wavy line between a carbon atom and an oxygen atom denotes an R or S configuration at the carbon atom; and wherein R_7 is a substituted or

30 unsubstituted allyl group.

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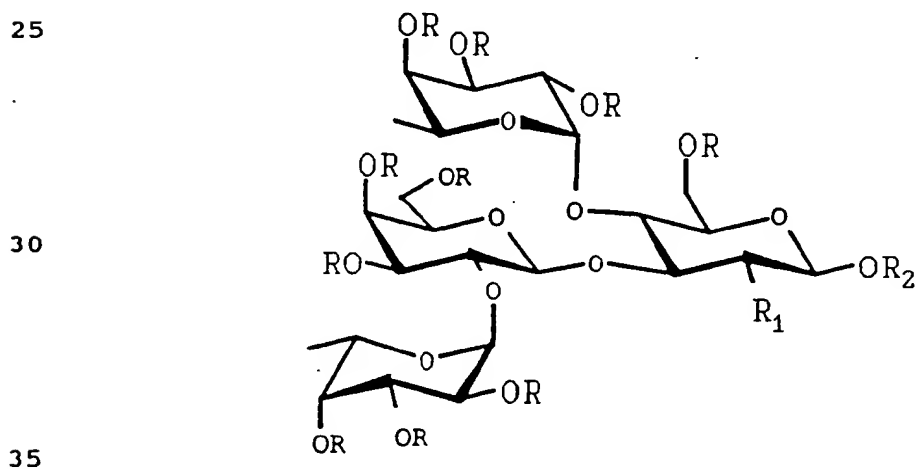
The invention also provides a compound having the structure:



15 wherein n is an integer from 1 to 18; wherein R is H or a linear or branched chain acyl group; wherein R_1 is H , OH , NH_2 or NHR_4 , where R_4 is SO_2Ph , a linear or branched chain alkyl or acyl group, or an aryl group; and wherein R_2 is a substituted or unsubstituted allyl group. In one embodiment, the invention provides the compound wherein

20 n is 1.

The invention further provides a compound having the structure:



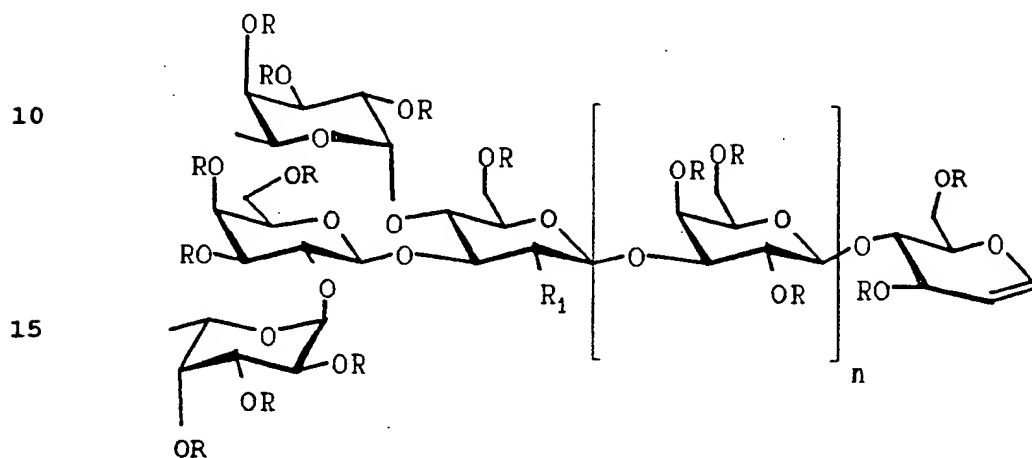
wherein R is H or a linear or branched chain acyl group; wherein R_1 is H , OH , NH_2 or NHR_4 , where R_4 is SO_2Ph , a

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linear or branched chain alkyl or acyl group, or an aryl group; and wherein R₂ is a substituted or unsubstituted allyl group.

5 The invention also provides a compound having the structure:

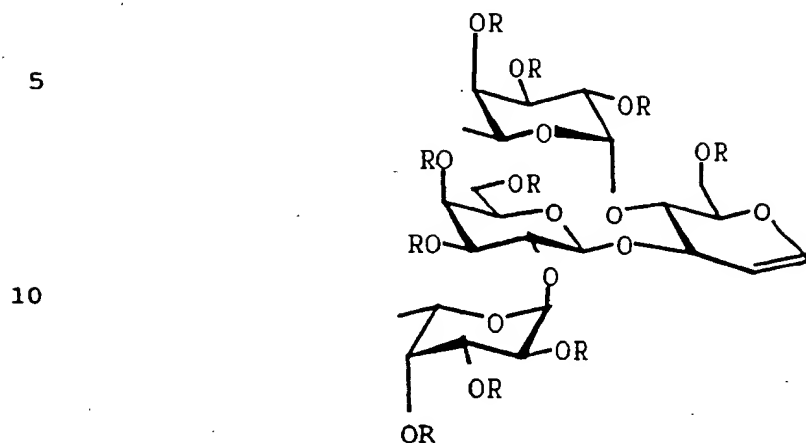


wherein R is H or a linear or branched chain acyl group;
20 wherein R₁ is H, OH, NH₂ or NHR₄, where R₄ is SO₂Ph, a
linear or branched chain alkyl or acyl group, or an aryl
group; wherein R₂ is a substituted or unsubstituted allyl
group; and wherein n is an integer from 1 to 18. In one
embodiment, the invention provides the compound wherein
25 n is 1.

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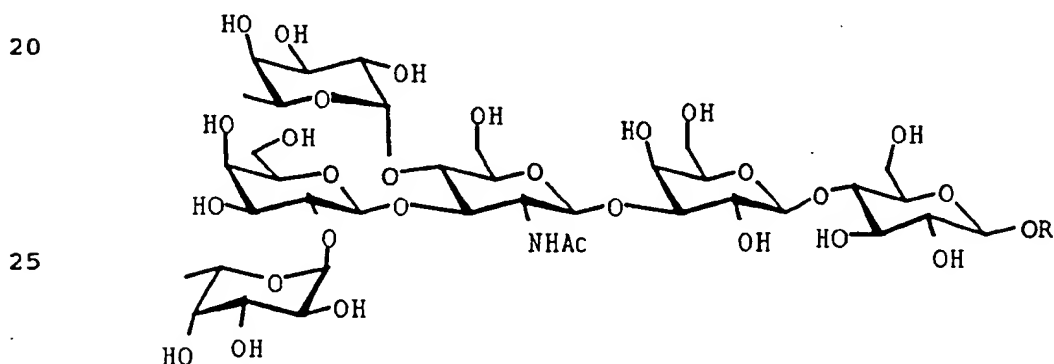
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The invention also provides a compound having the structure:



wherein R is H or a linear or branched chain acyl group.

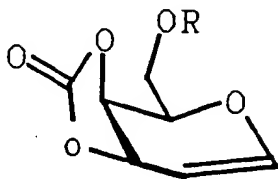
The invention also provides a process for synthesizing a compound having the structure:



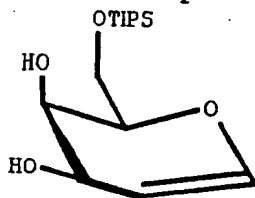
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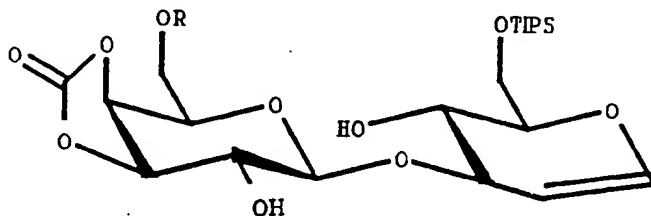
wherein R is a substituted or substituted allyl group,
which comprises the steps of (a) synthesizing a compound
having the structure:



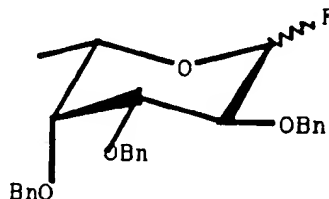
wherein R is a trialkylsilyl, aryldialkylsilyl,
alkyldiarylsilyl or triarylsilyl group; (b) reacting the
compound of step (a) with a compound having structure:



under suitable conditions to form a compound having the
structure:



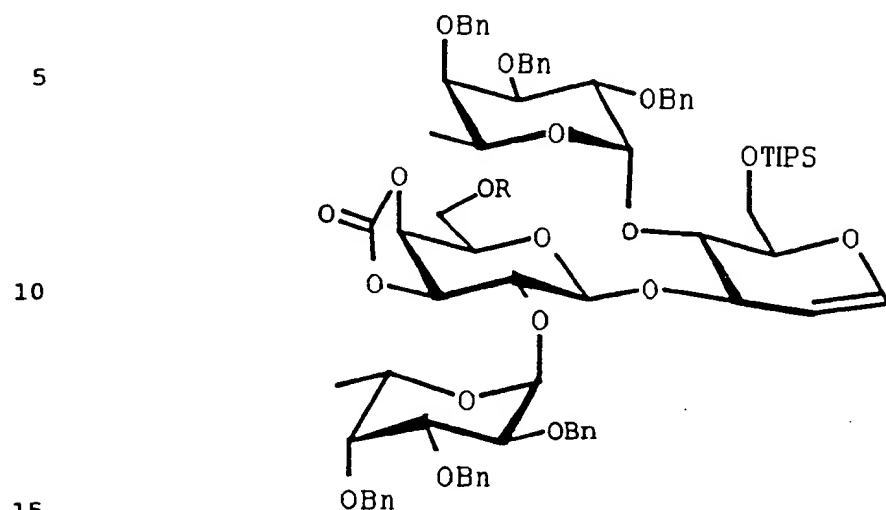
wherein R is a trialkylsilyl, aryldialkylsilyl,
alkyldiarylsilyl or triarylsilyl group; (c) reacting the
compound formed in step (b) with a compound having the
structure:



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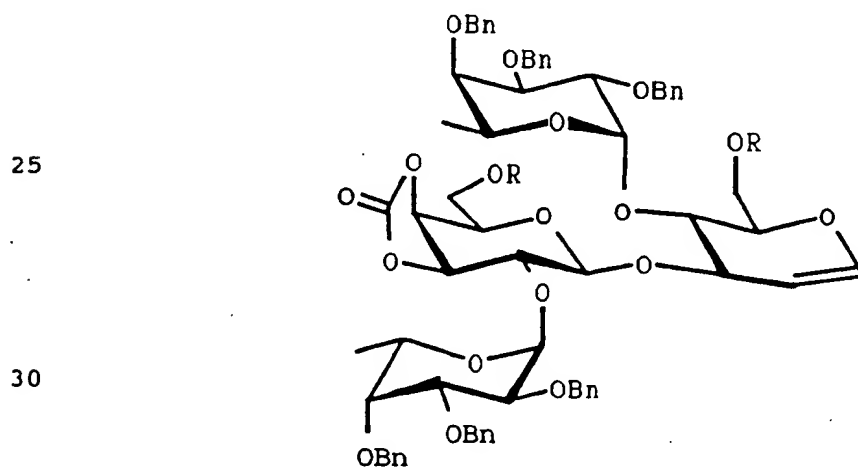
-41-

under suitable conditions to form a compound having the structure:



wherein R is a trialkylsilyl, aryldialkylsilyl, alkyl diarylsilyl or triarylsilyl group; (d) deprotecting and re-protecting the compound formed in step (c) under suitable conditions to form a compound having the structure:

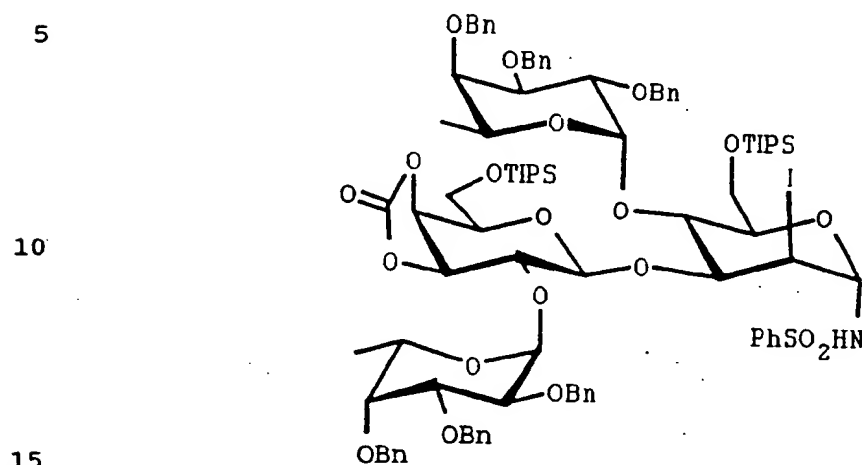
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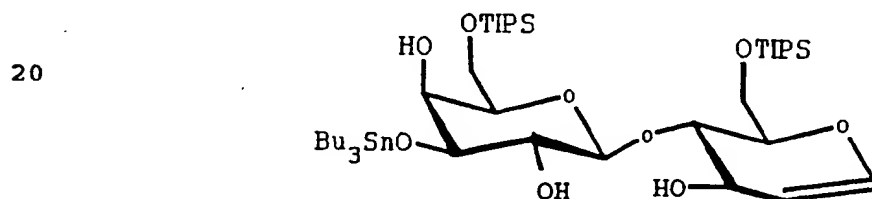
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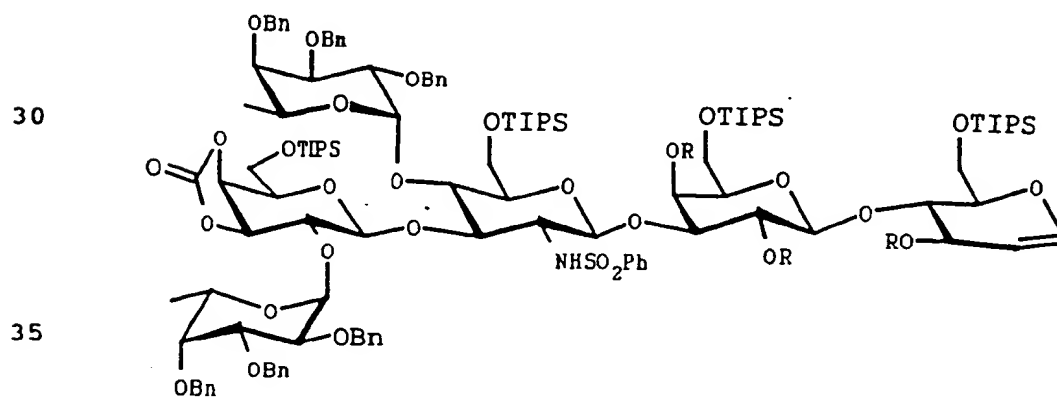
wherein R is TIPS; (e) iodosulfonamidating the compound formed in step (d) under suitable conditions to form a compound having the structure:



(f) reacting the compound formed in step (e) with a compound having the structure:



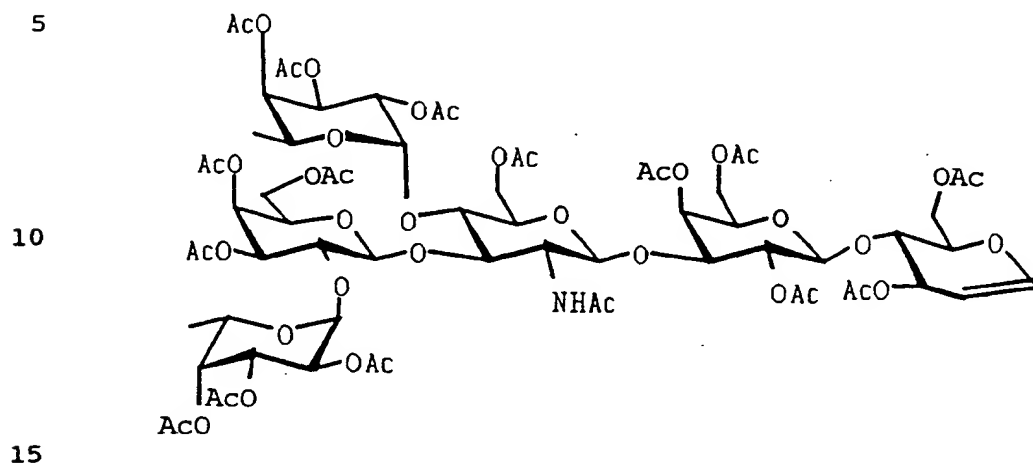
under suitable conditions to form a compound having the structure:



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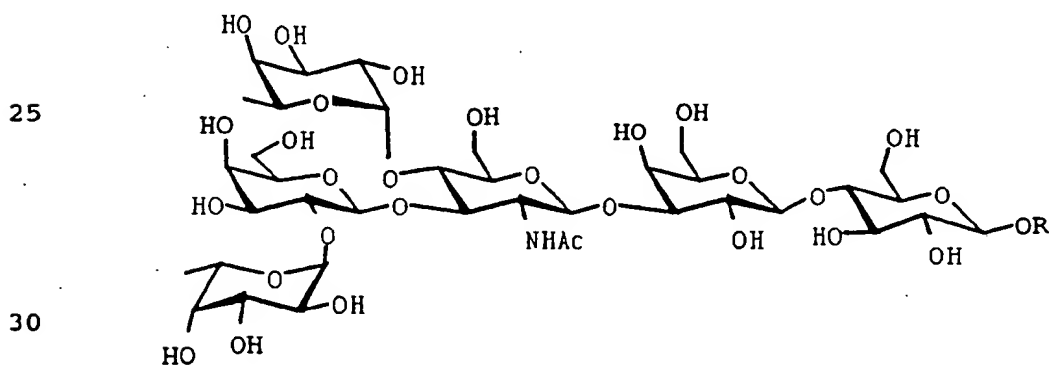
-43-

wherein R is H; (g) deprotecting and peracetylating the compound formed in step (f) under suitable conditions to form a compound having the structure:



(h) epoxidizing the compound formed in step (g) under suitable conditions to form an epoxide thereof and reacting the epoxide under suitable conditions to form a compound having the structure:

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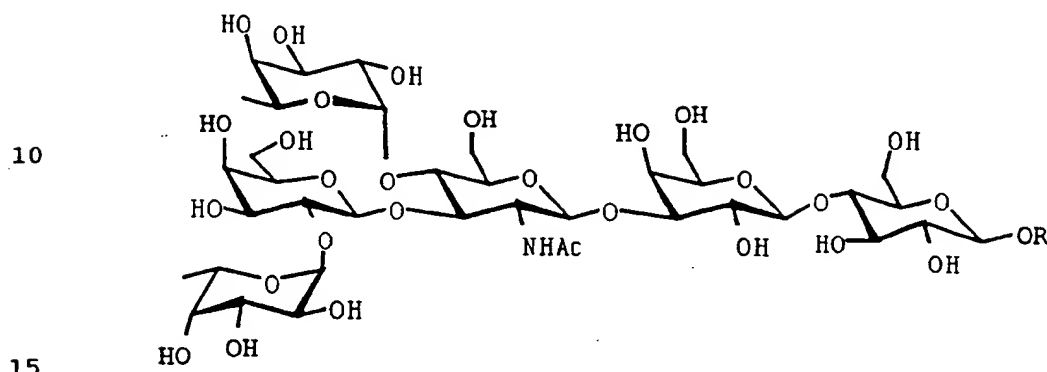
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wherein R is a substituted or unsubstituted allyl group;
and (i) treating the compound formed in step (h) under
suitable conditions to form a compound having the
structure:

5



20 wherein R is a substituted or unsubstituted allyl group.
In the above process the suitable conditions necessary
for the various reactions and treatments may be found in
the Experimental Details section which follows
hereinafter. However, it is within the confines of the
25 present invention that the specific reagents and solvents
provided as well as the specific conditions necessary for
reaction or treatment may be substituted with other
suitable reactants, solvents and conditions well known to
those skilled in the art.

30

The allyl compound may be conjugated to a peptide or
protein via amine or carboxylic acid side chain. In
practicing the invention, a bioconjugate is prepared
35 according to the protocol of Bernstein and Hall
(Carbohydr. Res. 1980, 78, C1). The allyl group is
ozonolyzed to form either an aldehyde or carboxylic acid,

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which is condensed to a terminal amine to form, respectively, an imine or an amide. The imine is reduced with sodium borohydride to the amine. Alternatively, the aldehyde is reductively aminated using procedures known in the art to form an amine which is reacted with a side-chain terminal carboxylic acid to form an amide conjugate.

The invention provides a pharmaceutical composition which comprises a therapeutically effective amount of the compound disclosed hereinabove and a pharmaceutically acceptable carrier.

Pharmaceutically acceptable carriers are well known to those skilled in the art and include, but are not limited to, 0.01-0.1M and preferably 0.05M phosphate buffer or 0.8% saline. Additionally, such pharmaceutically acceptable carriers may be aqueous or non-aqueous solutions, suspensions, and emulsions. Examples of non-aqueous solvents are propylene glycol, polyethylene glycol, vegetable oils such as olive oil, and injectable organic esters such as ethyl oleate. Aqueous carriers include water, alcoholic/aqueous solutions, emulsions or suspensions, including saline and buffered media. Parenteral vehicles include sodium chloride solution, Ringer's dextrose, dextrose and sodium chloride, lactated Ringer's or fixed oils. Intravenous vehicles include fluid and nutrient replenishers, electrolyte replenishers such as those based on Ringer's dextrose, and the like. Preservatives and other additives may also be present, such as, for example, antimicrobials, antioxidants, chelating agents, inert gases and the like.

The invention further provides a method for treating a subject afflicted with a disorder caused by Helicobacter pylori which comprises administering to the subject a therapeutically effective amount of the pharmaceutical

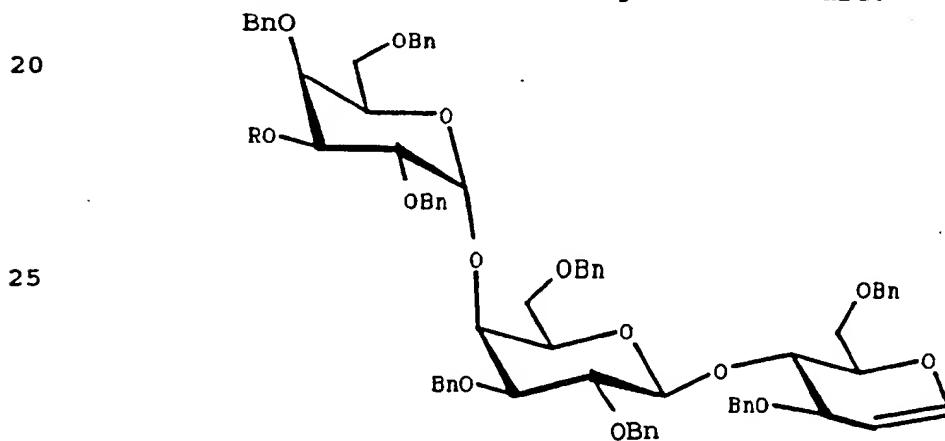
-46-

composition disclosed hereinabove so as to treat the subject afflicted with the disorder.

5 In one embodiment, the invention provides a method of treating a subject afflicted with gastric or duodenal ulcer. In another embodiment, the invention provides a method of treating a subject afflicted with gastric adenocarcinoma.

10 In addition, the invention provides a method for inhibiting the adhesion of Helicobacter pylori to gastric epithelium in a subject which comprises administering to the subject an amount of the compound disclosed hereinabove effective to inhibit the adhesion of
15 Helicobacter pylori to gastric epithelium in the subject.

The present invention also provides a process for synthesizing a compound having the structure:



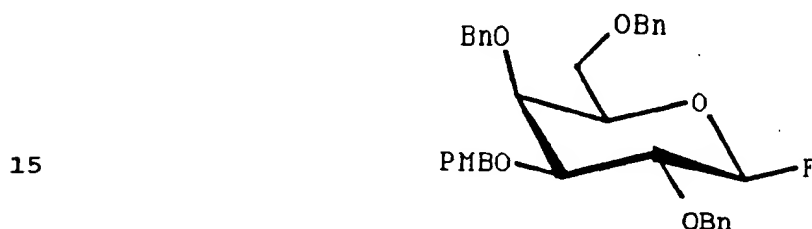
30 wherein R is H which comprises: (a) (i) reacting a compound having the structure:



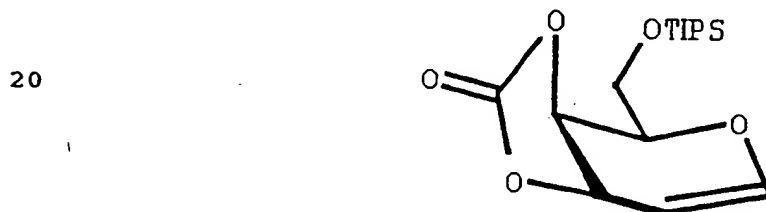
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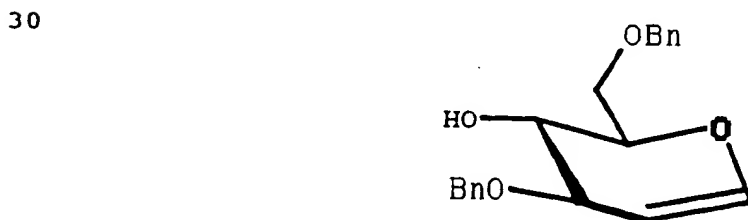
with an epoxidizing agent under suitable conditions to form an epoxide; (ii) cleaving the epoxide formed in step (a)(i) under suitable conditions with R_4NF wherein each R is independently the same or different and is a linear or branched chain alkyl, aralkyl or aryl group to form a fluoroalcohol; and (iii) alkylating the fluoroalcohol formed in step (b)(ii) under suitable conditions with a non-nucleophilic base and an organic halide having the formula $C_6H_5CH_2X$ wherein X is Br, Cl, I or F to form a compound having the structure:



(b) (i) synthesizing a compound having the structure:



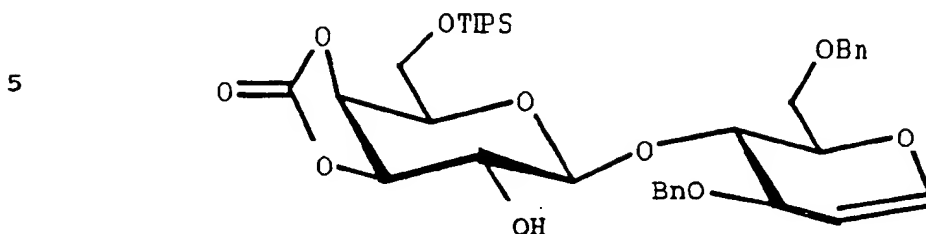
25 (c) (i) treating the compound formed in step (b) with an epoxidizing agent under suitable conditions to form an epoxide; and (ii) coupling the epoxide formed in step (c)(i) with a compound having the structure:



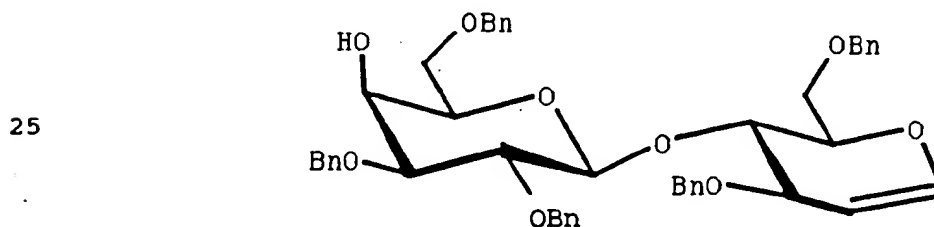
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under suitable conditions to form a compound having the structure:



- (d) (i) alkylating the compound formed in step (c)(ii) under suitable conditions with a non-nucleophilic base and an organic halide having the formula $C_6H_5CH_2X$ wherein X is Br, Cl, I or F; and (ii) de-silylating the compound formed in step (d)(i) under suitable conditions with R_4NF wherein each R is independently the same or different and is a linear or branched chain alkyl, aralkyl or aryl group; (iii) treating the compound formed in step (d)(ii) under suitable conditions with a metal alkoxide to form a deprotected disaccharide; and (iv) alkylating the disaccharide formed in step (d)(iii) under suitable conditions to form a selectively deprotected disaccharide having the structure:
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- 15
- 20

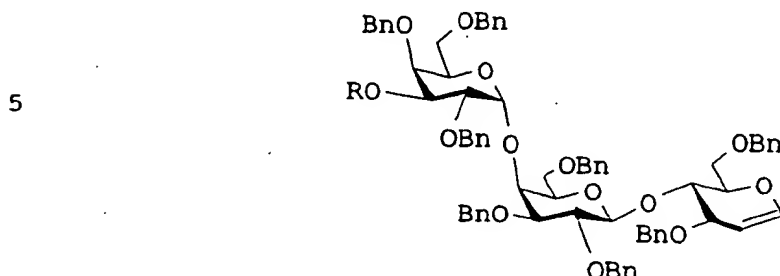


- (e) (i) coupling the selectively deprotected disaccharide formed in step (d)(iv) with the compound formed in step (a)(iii) under suitable conditions to form a protected trisaccharide; and (ii) de-protecting the protected
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trisaccharide formed in step (e)(i) under suitable conditions to form a trisaccharide having the structure:



10 wherein R is H. In step (a) reaction (i) may be carried out using a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide. A preferred agent is 3,3-dimethyldioxirane. Non-nucleophilic, inert solvents may be used, such as dichloromethane. Reaction (a)(ii) may be performed using organic ammonium fluoride salts, including tetrabutylammonium fluoride, in a range of solvents, including ethereal solvents, preferably in tetrahydrofuran. Step (iii) may be performed using a non-nucleophilic base such as sodium hydride in a non-nucleophilic solvent such as DMF. In step (b) the compound shown may be prepared as described herein. Step (c)(i) may be carried out using a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane. Coupling step (c)(ii) may be carried out using a metal catalyst, such as zinc chloride, in an inert solvent, such as THF. Step (d)(i) is carried out using a non-nucleophilic base such as sodium hydride in a non-nucleophilic solvent such as DMF. In step (d)(ii) de-silylation is effected using an organic ammonium fluoride salt, including tetrabutylammonium fluoride, in a range of solvents, including ethereal solvents, preferably in tetrahydrofuran. The carbonate ester is cleaved using a metal alkoxide, such as sodium methoxide, in an alcoholic

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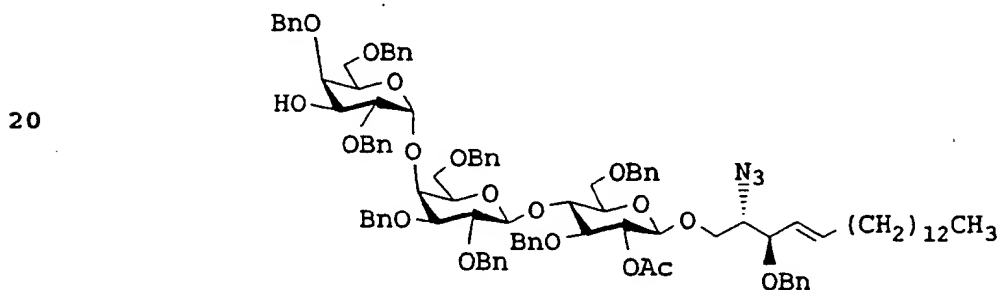
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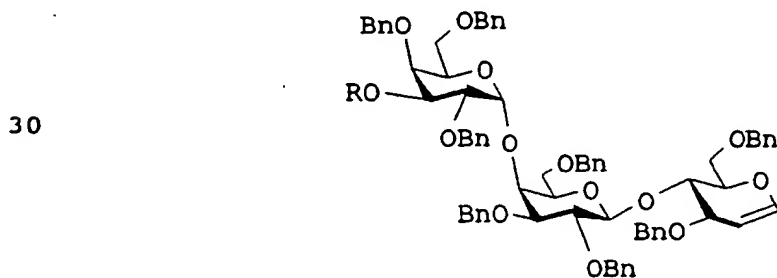
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medium such as methanol. Step (d)(iv) is selectively performed using a metal oxide, such as $(n\text{-Bu}_3\text{Sn})_2\text{O}$, in the presence of an organic ammonium bromide, such as tetra-*n*-butylammonium bromide, in an inert solvent such as benzene. Step (e) is a coupling performed in the presence of a metal halide salt, such as SnCl_4 , in the presence of silver perchlorate and 2,6-di-*t*-butylpyridine, in a solvent, such as ether, containing molecular sieves. Oxidative removal of PMB is performed with an oxidizing agent such as DDQ in an inert solvent system, which may preferably be heterogeneous, for example, using water/dichloromethane.

The present invention also provides a process for synthesizing a trisaccharide ceramide having the structure:



which comprises: (a) synthesizing a trisaccharide having the structure:

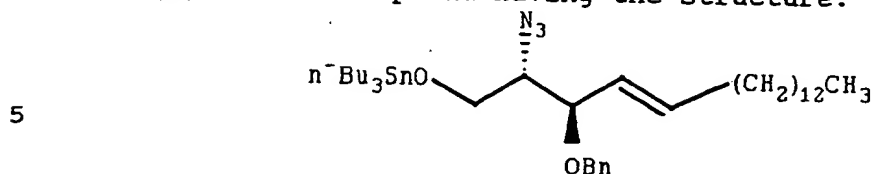


wherein R is PMB; (b) (i) reacting the trisaccharide formed in step (a) with an epoxidizing agent under suitable conditions to form a trisaccharide epoxide; and

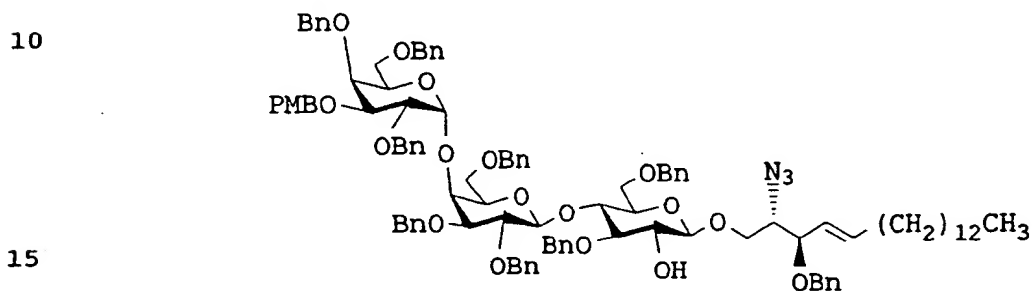
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(ii) reacting the trisaccharide epoxide formed in step (b)(i) with a compound having the structure:



under suitable conditions to form a protected trisaccharide ceramide having the structure:



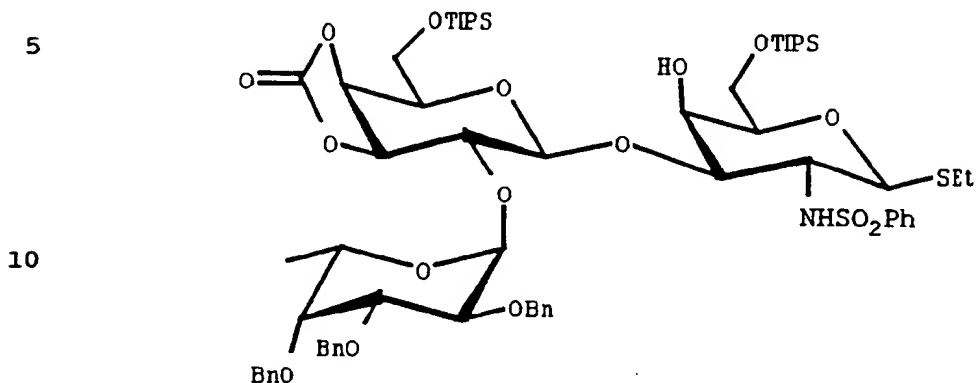
(c) (i) acylating the ceramide formed in step (b)(ii) under suitable conditions; and (ii) selectively de-protecting the compound formed in step (c)(i) under suitable conditions to form the trisaccharide ceramide.

In step (a) the trisaccharide may be synthesized as described herein. Step (b)(i) is performed using a variety of epoxidizing agents including peracetic acid, *m*-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane. Coupling step (b)(ii) may be carried out using a tributyltin ether of the ceramide precursor and a metal catalyst, such as zinc chloride, in an inert solvent, such as THF. In step (c)(i) acylation is performed using a linear or branched chain alkyl anhydride preferably acetic anhydride or halide in the presence of triethylamine and DMAP in an inert organic solvent such as dichloromethane. The PMB protecting group is removed oxidatively, preferably as described above.

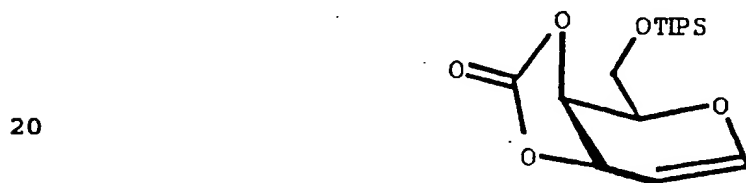
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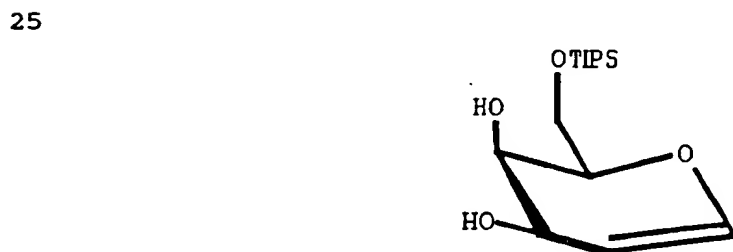
The present invention further provides a process for synthesizing a mercaptotrisaccharide having the structure:



which comprises: (a) (i) synthesizing a compound having the structure:



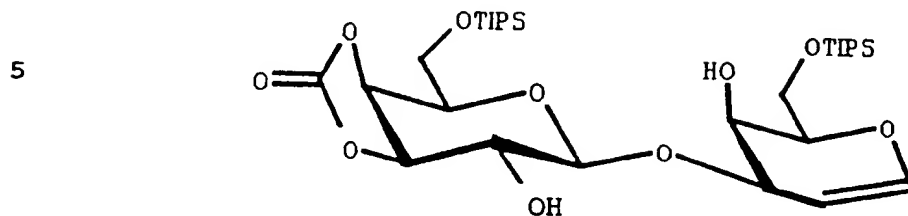
(ii) coupling the compound of step (a)(i) with a compound having structure:



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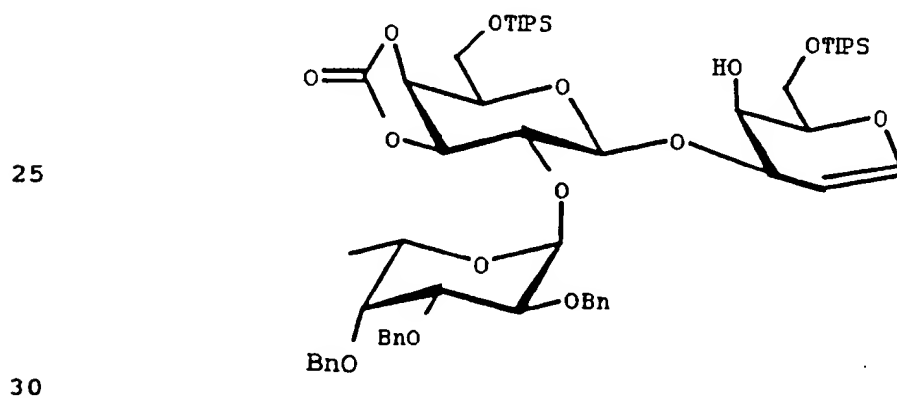
under suitable conditions to form a disaccharide having the structure:



10 (b) coupling the disaccharide formed in step (a)(ii) with a compound having the structure:



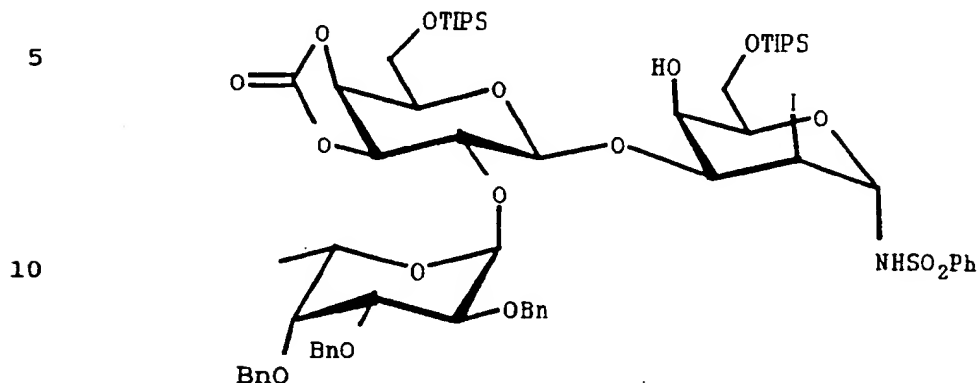
20 under suitable conditions to form a trisaccharide having the structure:



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(c) iodosulfonamidating the trisaccharide formed in step (b) under suitable conditions to form a iodosulfonamide having the structure:



and (d) reacting the iodosulfonamide formed in step (c) under suitable conditions with a thiolate to form the mercaptotrisaccharide.

20 Step (a)(ii) is performed by reacting the compound of step (a)(i), which may be obtained as described herein or otherwise, with a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane, followed by coupling with the diol monosaccharide of step (a)(ii) which may be carried out using a metal catalyst, such as zinc chloride, in an inert solvent, such as THF. Coupling with the

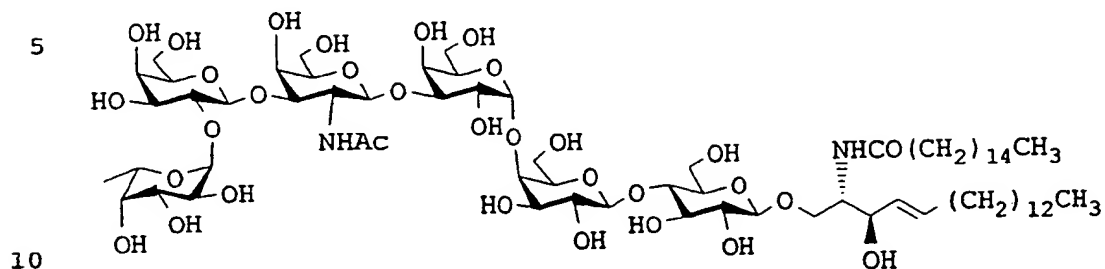
25 fluorosugar is carried out in step (b) in the presence of a metal halide salt, such as SnCl_2 , in the presence of silver perchlorate and 2,6-di-t-butylpyridine, in a solvent, such as ether, containing molecular sieves. Step (c) is performed using $\text{I}(\text{coll})_2$ perchlorate and

30 PhSO_2NH_2 in the presence of molecular sieves. Step (d) is carried out using alkyl thiol and a base such as LiHMDS in an inert solvent as DMF.

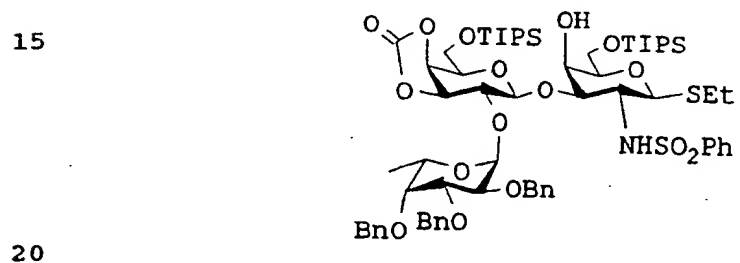
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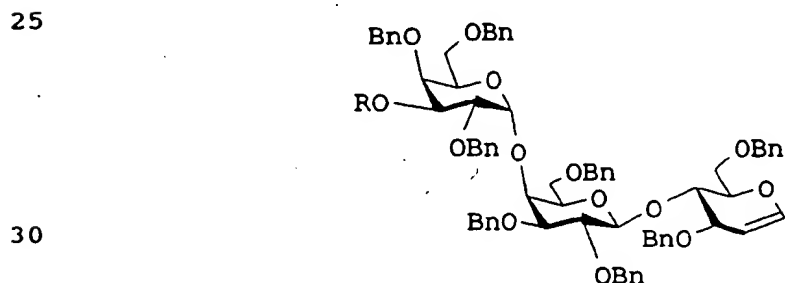
The present invention also provides a process of synthesizing a hexasaccharide ceramide having the structure:



which comprises: (a) coupling a compound having the structure:



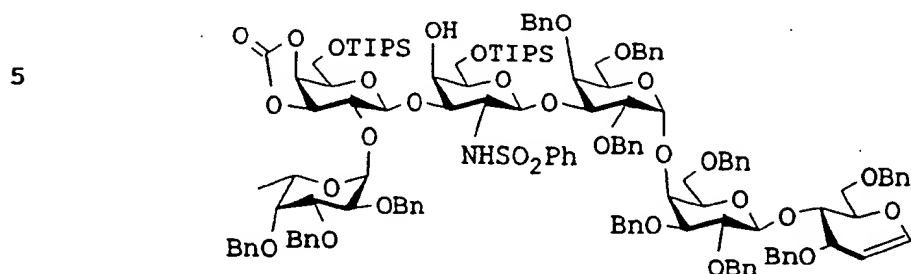
with a compound having the structure:



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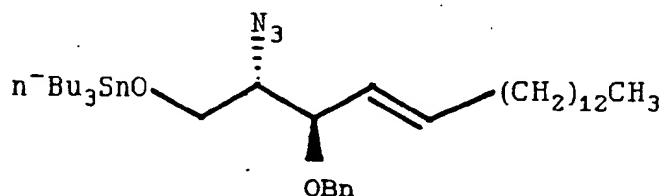
under suitable conditions to form a compound having the structure:



10

(b) (i) reacting the compound formed in step (a) with an epoxidizing agent under suitable conditions to form a hexasaccharide epoxide; and (ii) reacting the hexasaccharide epoxide with a stannyl ether having the structure:

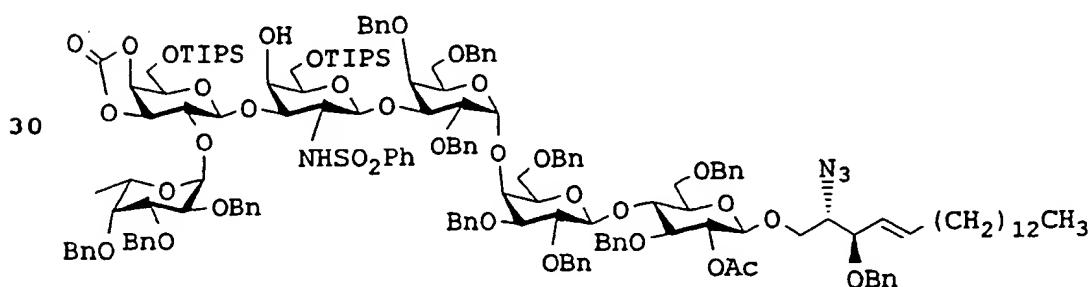
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under suitable conditions to form a hexasaccharide alcohol; (c) acylating the hexasaccharide alcohol formed in step (b)(ii) under suitable conditions to form a hexasaccharide acetate having the structure:

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(d) reductively acylating the hexasaccharide acetate formed in step (c) under suitable conditions in the

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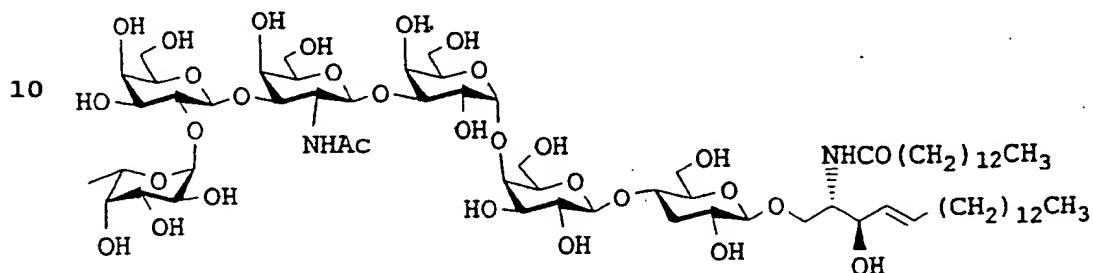
presence of palmitic anhydride to form a hexasaccharide ceramide; (e) desilylating and partially deprotecting the hexasaccharide ceramide under suitable conditions to form a partially deprotected hexasaccharide ceramide; (f) (i) reducing the partially deprotected hexasaccharide ceramide under suitable conditions to form a deprotected hexasaccharide ceramide acetate; and (ii) acylating the deprotected hexasaccharide ceramide acetate under suitable conditions to form a hexasaccharide ceramide peracetate; and (g) saponifying the hexasaccharide ceramide peracetate under suitable conditions to form the hexasaccharide ceramide.

Step (a) is performed using triflate esters, such as methyl triflate, in the presence of molecular sieves in an inert solvent. Step (b)(i) is carried out using a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane. Step (b)(ii) is performed using a stannyl ether of the ceramide precursor, preferably the tri-n-butyl stannylether, in the presence of a metal salt, such as Zn triflate, in an inert solvent, such as THF. Step (c) is carried out using acetic anhydride in the presence of a base such as triethylamine and DMAP. Step (d) is carried out using a noble metal catalyst such as Lindlar's catalyst and hydrogen gas in the presence of palmitic anhydride in an inert solvent such as ethyl acetate. Desilylation step (e) is effected using organic ammonium fluoride salts, such as tetra-n-butylammonium fluoride in THF. The carbonate ester is cleaved using a metal alkoxide such as NaOMe in an alcohol such as methanol. In step (f)(i) reduction is performed using a metal such as lithium or sodium in liquid ammonia and an inert solvent such as THF. Step (f)(ii) is carried out using acetic anhydride in the presence of a base such as Et₃N and DMAP

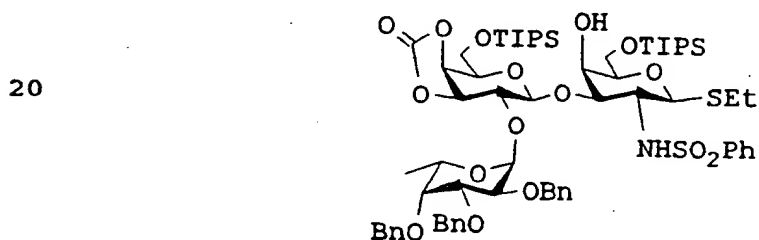
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in an inert solvent such as dichloromethane. The peracetate is saponified using a metal alkoxide such as sodium methoxide in an alcohol such as methanol.

- 5 The present invention also provides a process of synthesizing a hexasaccharide ceramide having the structure:

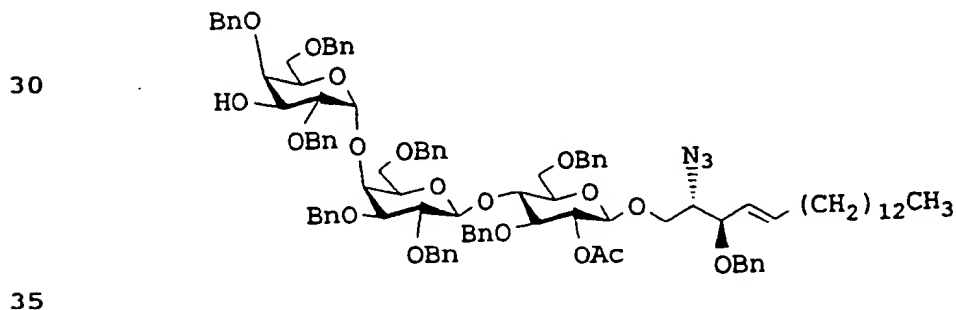


- 15 which comprises: (a) coupling a compound having the structure:

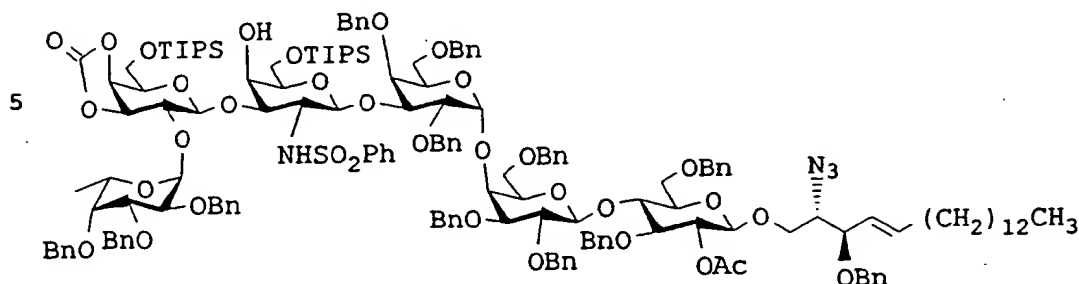


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with a compound having the structure:



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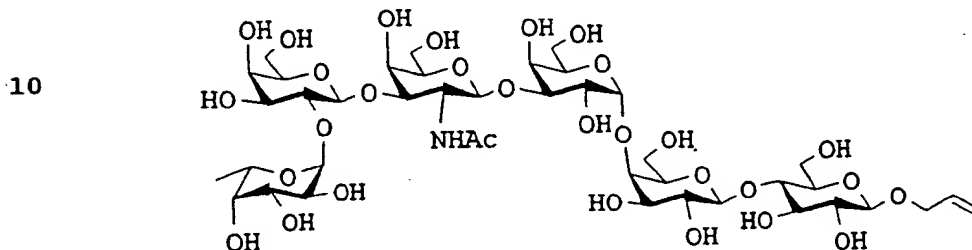
10 and (b) (i) reducing the hexasaccharide formed in step
(a) under suitable conditions in the presence of palmitic
anhydride to form a palmitoyl amide; (ii) desilylating
the palmitoyl amide with R_4NF wherein each R is
independently the same or different and is a linear or
15 branched chain alkyl, aralkyl or aryl group under
suitable conditions to form a partially deprotected
hexasaccharide; (iii) de-protecting the hexasaccharide
formed in step (b)(ii) under suitable conditions to form
a deprotected hexasaccharide; (iv) acylating the
20 hexasaccharide formed in step (b)(iii) under suitable
conditions to form a hexasaccharide ceramide peracetate;
and (v) saponifying the hexasaccharide ceramide
peracetate under suitable conditions to form the
hexasaccharide ceramide.

Step (a) is performed using triflate esters, such as methyl triflate, in the presence of molecular sieves in an inert solvent. Step (b)(i) is carried out using using a noble metal catalyst such as Lindlar's catalyst and hydrogen gas in the presence of palmitic anhydride in an inert solvent such as ethyl acetate. Step (b)(ii) is performed using organic ammonium fluoride salts, such as tetra-n-butylammonium fluoride in THF. In step (b)(iii) reduction is performed using a metal such as lithium or sodium in liquid ammonia and an inert solvent such as THF. Step (b)(iv) is carried out using acetic anhydride in the presence of a base such as Et₃N and DMAP in an

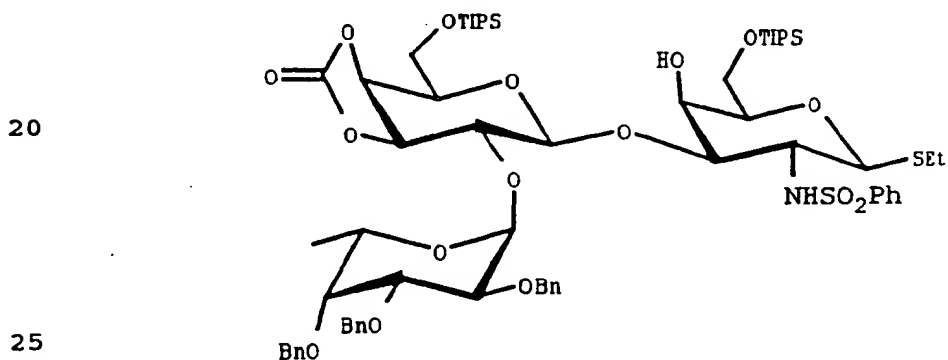
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inert solvent such as dichloromethane. In step (v) the peracetate carbonate is saponified using a metal alkoxide such as sodium methoxide in an alcohol such as methanol.

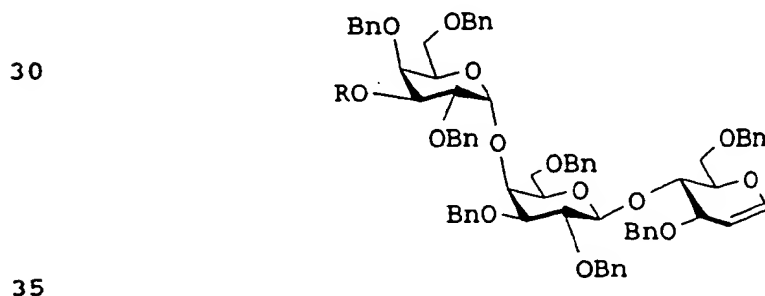
- 5 The present invention also provides a process of synthesizing an allyl hexasaccharide having the structure:



- 15 which comprises: (a) coupling a compound having the structure:



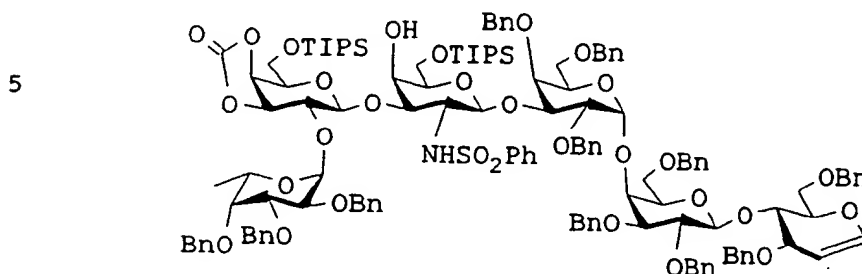
with a compound having the structure:



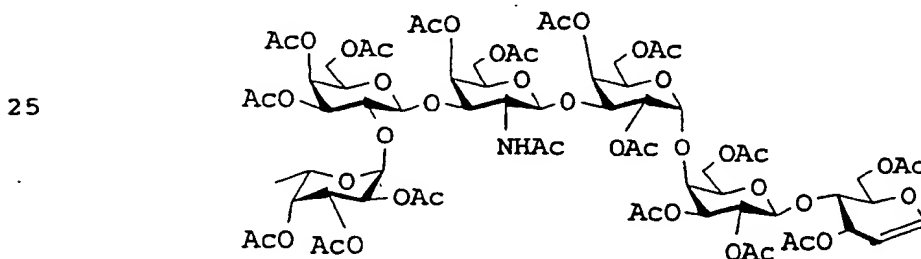
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wherein R is H under suitable conditions to form a hexasaccharide having the structure:



- (b) (i) desilylating the compound formed in step (a) with R_4NF wherein each R is independently the same or different and is a linear or branched chain alkyl, aralkyl or aryl group under suitable conditions to form a partially deprotected hexasaccharide; (ii) de-protecting the hexasaccharide formed in step (b)(i) under suitable conditions to form a deprotected hexasaccharide; and (iii) peracylating the compound formed in step (b)(ii) under suitable conditions to form a hexasaccharide peracetate having the structure:
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- (c) (i) reacting the hexasaccharide peracetate formed in step (b)(iii) with an epoxidizing agent under suitable conditions to form an hexasaccharide epoxide peracetate; (ii) treating the hexasaccharide epoxide peracetate formed in step (c)(i) with allyl alcohol under suitable conditions to form an allyl hexasaccharide peracetate; and (iii) saponifying the allyl hexasaccharide peracetate
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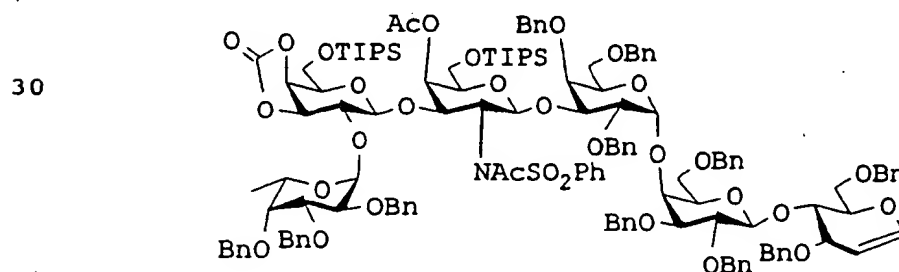
-62-

under suitable conditions to form the allyl hexasaccharide.

Step (a) is performed using triflate esters, such as methyl triflate, in the presence of molecular sieves in an inert solvent. Step (b)(i) is carried out using organic ammonium fluoride salts, such as tetra-n-butylammonium fluoride in THF. Step (b)(ii) is performed using a metal alkoxide such as sodium methoxide in an alcohol such as methanol, followed by reduction performed using a metal such as lithium or preferably sodium in liquid ammonia and an inert solvent such as THF. Step (b)(iii) is carried out using acetic anhydride in the presence of a base such as Et₃N and DMAP in an inert solvent such as dichloromethane. In step (c)(i) is carried out using a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane. Step (c)(ii) is carried out using allyl alcohol in an inert solvent. Step (c)(iii) the peracetate carbonate is saponified using a metal alkoxide such as sodium methoxide in an alcohol such as methanol.

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The present invention provides a process of synthesizing a hexasaccharide having the structure:

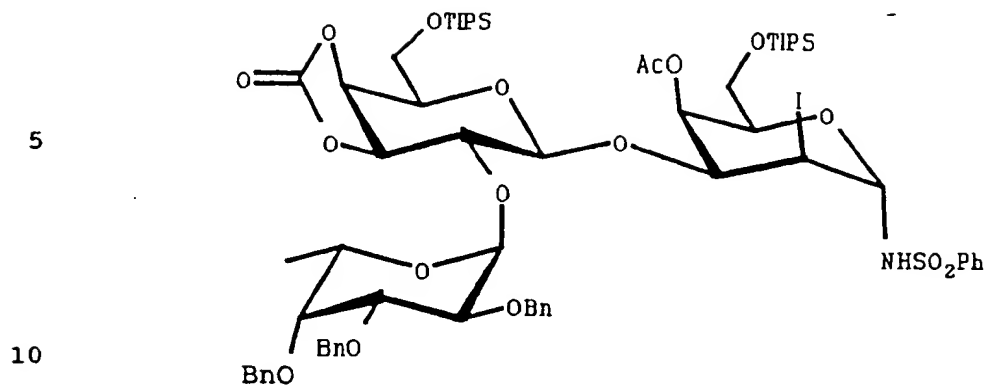


which comprises: (a) coupling a compound having the

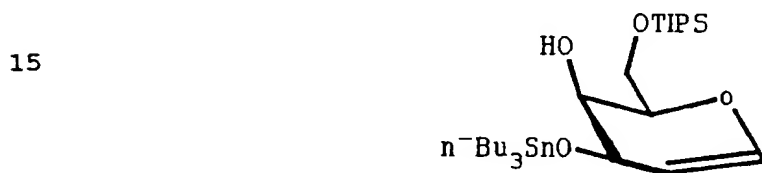
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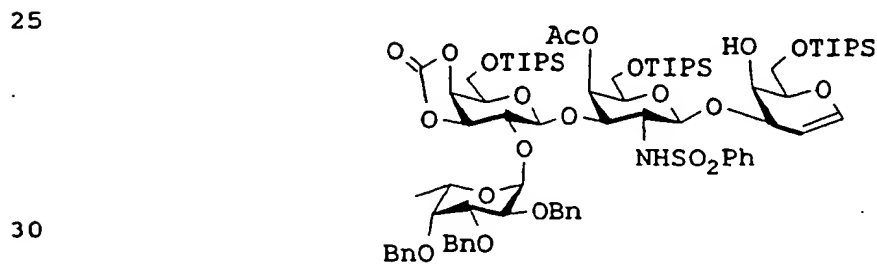
structure:



with a compound having the structure:



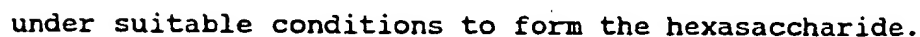
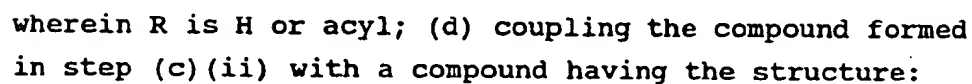
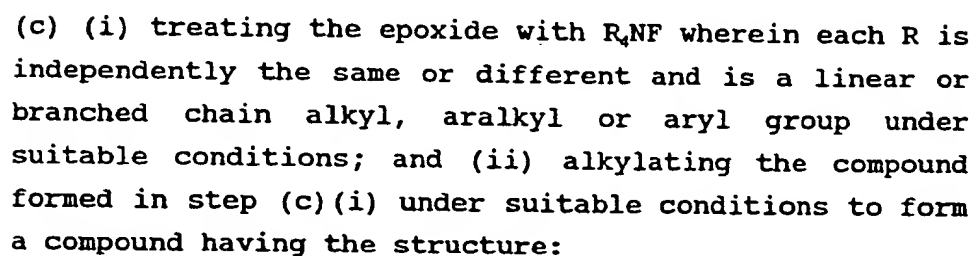
under suitable conditions to form a compound having the structure:



(b) (i) acylating the compound formed in step (a) under suitable conditions; and (ii) reacting the compound formed in step (b)(i) with an epoxidizing agent under

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suitable conditions to form an epoxide having the structure:



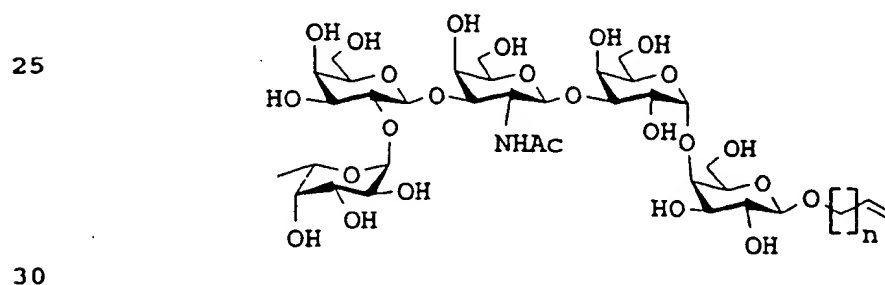
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Step (a) is performed using a metal catalyst such as silver tetrafluoroborate in an inert solvent. Step (b)(i) is carried out using acetic anhydride in the presence of a base such as Et₃N and DMAP in an inert solvent such as dichloromethane. Step (b)(ii) is carried out using a variety of epoxidizing agents including peracetic acid, m-chlorobenzoic acid, trifluoroacetic acid, and hydrogen peroxide, 3,3-dimethyldioxirane being preferred, in non-nucleophilic, inert solvents, such as dichloromethane. Step (c)(i) is effected with organic ammonium fluoride salts, such as tetra-n-butylammonium fluoride in THF. Step (c)(ii) is performed using a non-nucleophilic base such as sodium hydride in an inert solvent. Step (d) is performed using a metal salt catalyst such as tin dichloride in the presence of silver perchlorate in an inert solvent such as di-*t*-butylpyridine. Further transformations provide deprotected products or conjugates with proteins or other carriers.

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The present invention further provides a compound having the structure:



wherein n is an integer between about 0 and about 9.

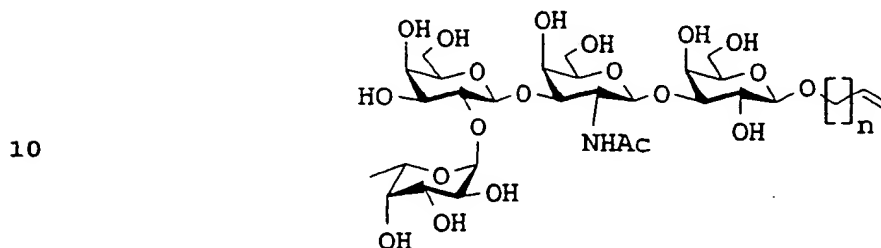
The allyl glycoside shown is prepared using the glycal coupling methods taught herein, and may be bound to protein carriers using general reactions described herein or by standard methods in the art. For example, the allyl glycoside may be prepared by coupling compound 9b

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disclosed herein with a suitably protected 8b, followed by coupling with 12b, then coupling with allyl alcohol and an appropriate deprotection sequence.

- 5 The present invention also provides a compound having the structure:

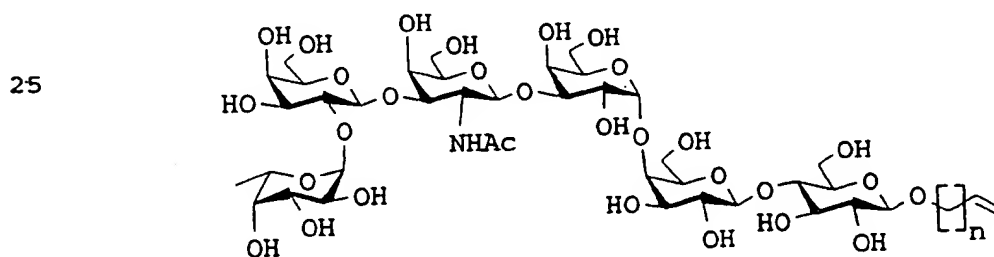


wherein n is an integer between about 0 and about 9.

- 15 The allyl glycoside shown is prepared using the glycal coupling methods, allylation and a deprotection sequence as taught herein (see Fig. 12), and may be bound to protein carriers using general reactions described herein or by standard methods in the art.

20

The present invention also provides a compound having the structure:



- 30 wherein n is an integer between about 0 and about 9.

- 35 The allyl glycosides shown are prepared using the glycal coupling methods taught herein, and may be bound to protein carriers using general reactions described herein or by standard methods in the art.

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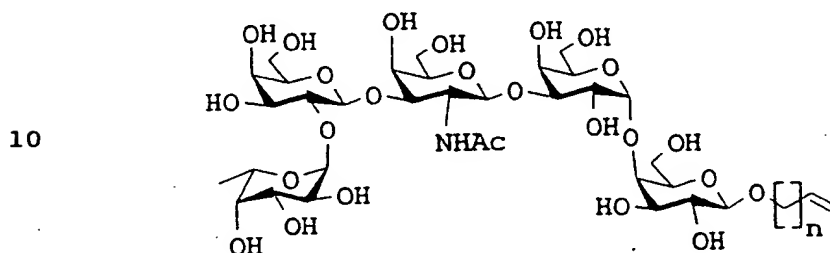


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The present invention also provides a method of inducing antibodies in a subject, wherein the antibodies are immunoreactive with human breast tumor cells, which comprises administering to the subject an amount of the compound having the structure:



15 wherein n is an integer between about 0 and about 9 either alone or bound to a suitable immunological adjuvant effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the antibodies induced are MBrl antibodies. In another embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease. In another embodiment, the present invention provides a method wherein the adjuvant is a protein carrier, bacteria or liposomes. In yet another embodiment, the present invention provides wherein the adjuvant is bacille Calmette-Guerin.

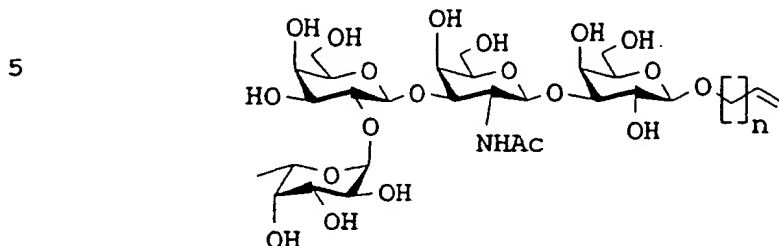
30 The present invention provides a method of preventing recurrence of breast cancer in a subject which comprises vaccinating the subject with the compound shown hereinabove either alone or bound to a suitable immunological carrier, adjuvant or vehicle.

35 The present invention also provides a method of inducing antibodies in a subject, wherein the antibodies are immunoreactive with human breast tumor cells, which

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comprises administering to the subject an amount of the compound having the structure:



15 wherein n is an integer between about 0 and about 9 either alone or bound to a suitable immunological adjuvant effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the antibodies induced are MBr1 antibodies. In another embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease. In another embodiment, the present invention provides a method wherein the adjuvant is a protein carrier, bacteria or liposomes. In yet another embodiment, the present invention provides wherein the adjuvant is bacille Calmette-Guerin.

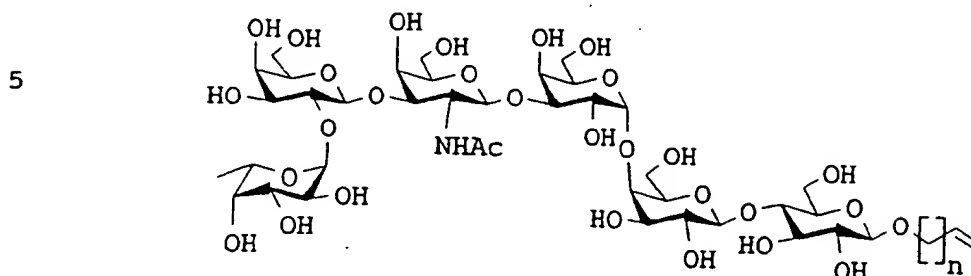
25 The present invention also provides a method of preventing recurrence of breast cancer in a subject which comprises vaccinating the subject with the compound shown hereinabove either alone or bound to a suitable immunological carrier, adjuvant or vehicle.

30 The present invention additionally provides a method of inducing antibodies in a subject, wherein the antibodies are immunoreactive with human breast tumor cells, which

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comprises administering to the subject an amount of the compound having the structure:



wherein n is an integer between about 0 and about 9 either alone or bound to a suitable immunological adjuvant effective to induce the antibodies. In one embodiment, the present invention provides a method wherein the antibodies induced are MBrl antibodies. In another embodiment, the present invention provides a method wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease. In another embodiment, the present invention provides a method wherein the adjuvant is a protein carrier, bacteria or liposomes. In yet another embodiment, the present invention provides wherein the adjuvant is bacille Calmette-Guerin.

25 The present invention also provides a method of preventing recurrence of breast cancer in a subject which comprises vaccinating the subject with the compound shown hereinabove either alone or bound to a suitable immunological carrier, adjuvant or vehicle.

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Experimental DetailsGeneral Procedures

- 5 All air- and moisture-sensitive reactions were performed in a flame-dried apparatus under an argon atmosphere unless otherwise noted. Air-sensitive liquids and solutions were transferred via syringe or canula. Wherever possible, reactions were monitored by thin-layer chromatography (TLC). Gross solvent removal was performed in vacuum under aspirator vacuum on a Buchi rotary evaporator, and trace solvent was removed on a high vacuum pump at 0.1-0.5 mmHg.
- 10
- 15 Melting points (mp) were uncorrected and performed in soft glass capillary tubes using an Electrothermal series IA9100 digital melting point apparatus.

- Infrared spectra (IR) were recorded using a Perkin-Elmer 1600 series Fourier-Transform instrument. Samples were prepared as neat films on NaCl plates unless otherwise noted. Absorption bands are reported in wavenumbers (cm^{-1}).
- 20
- Only relevant, assignable bands are reported.

- 25 Proton nuclear magnetic resonance (^1H NMR) spectra were determined using a Bruker AMX-400 spectrometer at 400 MHz. Chemical shifts are reported in parts per million (ppm) downfield from tetramethylsilane (TMS; $\delta=0$ ppm) using residual CHCl_3 as a lock reference ($\delta=7.25$ ppm).
- 30 Multiplicities are abbreviated in the usual fashion: s=singlet; d=doublet; t=triplet; q=quartet; m=multiplet; br=broad.

- Carbon nuclear magnetic resonance (^{13}C NMR) spectra were performed on a Bruker AMX-400 spectrometer at 100 MHz with composite pulse decoupling. Samples were prepared as with ^1H NMR spectra, and chemical shifts are reported
- 35

relative to TMS (0 ppm); residual CHCl_3 was used as an internal reference ($\delta=77.0$ ppm).

5 All high resolution mass spectral (HRMS) analyses were determined by electron impact ionization (EI) on a JEOL JMS-DX 303HF mass spectrometer with perfluorokerosene (PFK) as an internal standard. Low resolution mass spectra (MS) were determined by either electron impact ionization (EI) or chemical ionization (CI) using the
10 indicated carrier gas (ammonia or methane) on a Delsi-Nermag R-10-10 mass spectrometer. For gas chromatography/mass spectra (GCMS), a DB-5 fused capillary column (30 m, 0.25mm thickness) was used with helium as the carrier gas. Typical conditions used a
15 temperature program from 60-250°C at 40°C/min.

Thin layer chromatography (TLC) was performed using precoated glass plates (silica gel 60, 0.25 mm thickness). Visualization was done by illumination with
20 a 254 nm UV lamp, or by immersion in anisaldehyde stain (9.2 mL p-anisaldehyde in 3.5 mL acetic acid, 12.5 mL conc. sulfuric acid and 338 mL 95% ethanol (EtOH)) and heating to colorization.

25 Flash silica gel chromatography was carried out according to the standard protocol.

Unless otherwise noted, all solvents and reagents were commercial grade and were used as received, except as
30 indicated hereinbelow, where solvents were distilled under argon using the drying methods listed in parentheses: CH_2Cl_2 (CaH_2); benzene (CaH_2); THF (Na/ketyl); Et_2O (Na/ketyl); diisopropylamine (CaH_2).

35 Abbreviations

OTf triflate

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	TLC	thin layer chromatography
	EtOAc	ethyl acetate
	TIPS	triisopropylsilyl
	PMB	p-methoxybenzyl
5	Bn	benzyl
	Ac	acetate
	hex	hexane
	THF	tetrahydrofuran
	coll	collidine
10	LiHMDS	lithium hexamethyldisilazide
	DAST	diethylaminosulfur trifluoride
	DMF	N,N-dimethylformamide
	DMAP	2-dimethylaminopyridine
	DDQ	2,3-dichloro-5,6-dicyano-1,4-benzoquinone
15	TBAF	tetra-n-butylammonium fluoride
	M.S.	molecular sieves
	r.t.	room temperature
	r.b.	round bottom flask

20

EXAMPLE 1

Preparation of Polymer-Bound Glucal 18:

Polymer-bound galactal 7 (500 mg; S.J. Danishefsky, et
25 al., J. Am. Chem. Soc. 1992, 8331) was placed in a 100 mL
polymer flask and dried in vacuo. On cooling to 0°C
under N₂, dry CH₂Cl₂ (20 mL) and freshly prepared Murray
solution (30 mL; R.W. Murray and R. Jeyaraman, J. Org
Chem. 1985, 2847) was added. After stirring at 0°C for
30 -90 min., solubles were filtered using N₂ pressure. The
oxidation procedure was repeated. The resulting epoxide
of 7 kept on a vacuum line for -3 h to dry. A solution
of glucal 19 (1.0 g in 8 mL dry THF) was added, and the
mixture was cooled to -23°C (dry ice-CCl₄). A solution of
35 ZnCl₂ in THF (0.8 mL 1.0 M) was added. The mixture was
slowly allowed to warm to r.t. (over -2 h), and then
stirred at r.t. overnight. The polymer-bound glucal 18

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was rinsed with 3 x 20 mL THF, and dried on a vacuum line.

Preparation of Polymer-Bound Tetrasaccharide 20:

5 Polymer-bound glucal 18 and $\text{Sn}(\text{OTf})_2$ (0.80 g, 1.92 mmol) were combined and dried in vacuo. On cooling to 0°C under N_2 , a solution of fucosyl donor 10 (1.8 g, 4.1 mmol) in 20 mL dry THF with di-*t*-butylpyridine (1.7 mL, 7.57
10 mmol) was added. The mixture was allowed to warm slowly to r.t., and stirred overnight. The polymer was washed with 2 x 20 mL dry THF, 2 x 20 mL dry dioxane, 20 mL DMSO, and 2 x 20 mL THF. The resulting polymer-bound tetrasaccharide 20 was kept on a vacuum line to dry.

15

Preparation of Tetrasaccharide Glycal 21:

The polymer-bound tetrasaccharide 20 (50 mg) was stirred in 2 mL THF, and treated with 0.2 mL each of 1.0 M
20 solutions of TBAF and AcOH in THF. The mixture was stirred at 40°C overnight. The polymer was washed with 3 x 5 mL THF. The combined rinsings were concentrated and column-chromatographed on silica (2:1 EtOAc:hex), providing tetrasaccharide glycal 21 as a colorless gum.
25 Yield: 9.0 mg.

EXAMPLE 2

Preparation of Diol 18':

30

Galactal 7' (0.100 g, 0.304 mmol) in 5 mL dry CH_2Cl_2 at 0°C under a N_2 atmosphere was treated with 10 mL Murray solution (freshly prepared) and stirred at 0°C for 40 min. TLC (1:1 EtOAc:hex) showed no trace of 7'.
35 Solvents were evaporated using a dry N_2 stream. The residual epoxide of 7' was kept on a vac. line ~2h. To the epoxide under a N_2 atmosphere was added a solution of

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glucal derivative 3' (0.150 g, 0.496 mmol) in 3 mL dry THF. On cooling to -78°C , 1.0 M ZnCl_2 in Et_2O (0.50 mL, 0.50 mmol) was added. The mixture was allowed to slowly warm to r.t. (over ~ 2 h) and stirred overnight. TLC (1:1 EtOAc:hex) showed that the reaction was complete. Saturated aq. NaHCO_3 (20 mL) was added, and the mixture was then extracted with EtOAc (3 x 20 mL). The organic layer was dried over MgSO_4 . Column chromatography on silica (1:3 EtOAc:hex) afforded diol 18' as a colorless solid. Yield: 173 mg (89%). $[\alpha]_D^{25} - 9.8^{\circ}$ (c 1.0, CH_2Cl_2).

Preparation of Tetrasaccharide 22:

Diol 18' (86 mg, 0.133 mmol) and fucosyl donor 10 (0.290 g, 0.665 mmol) were azeotropically dried using benzene. The mixture was dissolved in 3 mL dry THF together with 0.65 mL di-*t*-butylpyridine and then added via canula to a flask containing $\text{Sn}(\text{OTf})_2$ (0.30 g, 0.72 mmol) and 4 Å MS (500 mg) at 0°C under N_2 atm. The mixture was stirred at 0°C ~ 7 h. TLC (1:3 EtOAc:hex) shows no trace of diol 18'. The mixture was partitioned between saturated aq. NaHCO_3 (100 mL) and EtOAc (2 x 100 mL). The organic layer was dried over MgSO_4 . The organic layer was filtered through silica using EtOAc to obtain crude material, which was then purified by chromatography on silica (1:9 EtOAc:hex) affording tetrasaccharide 22. Yield: 170 mg (86%).

Preparation of Iodosulfonamide 23:

Procedure 1.

Tetrasaccharide glycal 22 (120 mg, 81.1 μmol) and PhSO_2NH_2 (20 mg, 0.13 mmol) were azeotropically dried using benzene. Added (glove bag) 4 Å MS (0.2 g). After cooling to 0°C under N_2 , dry CH_2Cl_2 (1.0 mL) was added. The mixture was treated with a solution of $\text{I}(\text{coll})_2\text{ClO}_4$.

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(prepared from 100 mg $\text{Ag}(\text{coll})_2\text{ClO}_4$, 5 mL collidine, and 60 mg I_2 in 1 mL dry CH_2Cl_2) via canula through a plug of flame-dried celite and 4 Å MS. The mixture was stirred at 0°C for 40 min. TLC (1:4 EtOAc:hex) showed
5 iodosulfonamide 23 as the major component. The mixture was filtered through celite, which was rinsed with Et_2O . The organic layer was extracted with saturated aq. $\text{Na}_2\text{S}_2\text{O}_3$, saturated aq. CuSO_4 , brine, and then dried over MgSO_4 . Column chromatography on silica (1:4 EtOAc:hex) gave
10 iodosulfonamide 23 as a colorless solid.
Yield: 115 mg (80%).

Procedure 2.

Tetrasaccharide glycal 22 (200 mg, 0.135 mmol), PhSO_2NH_2
15 (42 mg, 0.27 mmol), and 200 mg powdered 4 Å MS in 2.0 mL dry CH_2Cl_2 at 0°C under a N_2 atmosphere was treated with $\text{I}(\text{coll})_2\text{ClO}_4$ (prepared from 120 mg $\text{Ag}(\text{coll})_2\text{ClO}_4$ and 67 mg I_2 in 1 mL dry CH_2Cl_2). The mixture was stirred at 0°C (protected from light using foil) for 30 min. TLC (1:2
20 EtOAc:hex) showed mainly iodosulfonamide with some glycal.

After ~1 h more at 0°C, TLC showed no noticeable improvement. The mixture was filtered through celite, which was washed with Et_2O . After extracting with saturated aq.
25 $\text{Na}_2\text{S}_2\text{O}_3$, saturated aq. CuSO_4 , brine, the organics were dried over MgSO_4 . Column chromatography on silica (1:3 EtOAc:hex) gave 23 as a colorless solid.
Yield: 165 mg (69%). $[\alpha]_D^{23} = -85.7^\circ$ (c 1.0, CH_2Cl_2).

30

Preparation of Hexasaccharide 25:

Iodosulfonamide 23 (60 mg, 34 mmol) in a 35 mL r.b. was treated with 200 mg powdered 4 Å MS (glove bag). To this
35 flask under N_2 was added a solution of protected lactal 24 in THF (1.5 mL). On cooling the mixture to -78°C, a solution of AgBF_4 (40 mg, 0.206 mmol) was added in 0.25 mL

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dry THF. The mixture was stirred and slowly warmed to r.t. overnight. The mixture was warmed to 45°C and stirred ~36 h. TLC showed only a trace of iodosulfonamide. Saturated aq. NH_4Cl (5 mL) was added, and the mixture was extracted with 3 x 10 mL EtOAc. The organic layer was dried over MgSO_4 . Column chromatography on silica (1:3 EtOAc:hex) afforded 25 as a colorless oil. Yield: 42 mg (55%).

$[\alpha]_D^{23} = -33.8^\circ$ (c 2.0, CH_2Cl_2)

Preparation of Hexasaccharide 25a:

Hexasaccharide 25 (55 mg, 24.4 mmol) in ~1.5 mL THF was treated at 0°C with TBAF (0.25 mL, 1.0 M solution in THF, 0.25 mmol), and stirred at r.t. overnight. TLC (1:9 MeOH: CH_2Cl_2) showed a 3:1 mixture of 25a vs. a less polar substance. Additional 1.0 M TBAF (0.10 mL) was added, and the mixture was stirred overnight at r.t. TLC showed that the reaction was complete. Solvents were removed using a N_2 stream. Column chromatography on silica (1:19 MeOH: CH_2Cl_2)

afforded a ~1:2 mixture corresponding to two compounds which differ only in the presence or absence of a 3,4-cyclic carbonate group. Crude yield: 35 mg total weight for two products. The crude mixture was used as such for the next reaction.

Preparation of Peracetylated Hexasaccharide 26:

Hexasaccharide 25a (36 mg) in 0.25 mL dry THF was added via canula to ~8 mL bright blue Na/NH_3 solution at -78°C (dry ice bath) under N_2 atm. After removing the dry ice bath, the mixture was stirred in refluxing NH_3 (dry ice condenser) for 15 min. After adding 2 mL dry MeOH (slowly!), the resulting mixture was stirred while blowing off NH_3 with a N_2 stream. The MeOH solution was treated with Dowex 50 x 8 [H^+] until pH ~8-9, and then

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filtered. The resin was washed with MeOH. The residue was concentrated and kept on a vacuum line to dry. Under a N₂ atmosphere, the residue was treated with 1 mL dry pyridine and 0.5 mL Ac₂O, and stirred at r.t. overnight.

5 TLC (EtOAc) showed that hexasaccharide 26 is major component. Upon concentration, the residue was purified by column chromatography on silica (1:4 hex:EtOAc).

Preparation of Hexasaccharide 17:

10

Hexasaccharide 26 (10.0 mg, 6.3 mmol) under N₂ at 0°C was treated with 0.5 mL dry CH₂Cl₂. Dioxirane solution (0.20 mL) was added, and the mixture was stirred at 0°C -40 min. TLC (EtOAc) showed no trace of 26. Solvents were

15

evaporated with a N₂ stream. The epoxide was dried on a vacuum line for ~2 h. The epoxide was treated under a N₂ atmosphere with 0.5 mL allyl alcohol (passed through basic alumina to dry) and 0.5 mL dry THF. On cooling to -78°C, 1.0 M ZnCl₂ (10 mL) in dry Et₂O was added. After

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warming slowly to r.t., the mixture was stirred overnight. Saturated aq. NaHCO₃ (5 mL) was added, and the mixture was extracted with 3 x 5 mL EtOAc. The combined organic layers were dried over MgSO₄, filtered and concentrated to an oil, which was dried on a vacuum line for -2 h. The residue was treated to pyridine:Ac₂O (2:1, 1.5 mL) while stirring overnight. Solvents were removed, and the residue was purified by column chromatography on silica (1:4 hex:EtOAc), affording hexasaccharide 17 as a colorless solid. Yield: 5.5 mg.

30

Results and Discussion

A Highly Convergent Synthesis of the Lewis Y Blood Group Determinant in Conjugatable Form

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Construction of the Le^y determinant commences with lactal (1a) (W.N. Haworth, E.L. Hirst, M.M.T. Plant, R.J.W.

Reynolds, J. Chem. Soc. 1930, 2644) as shown in Figure 2. Capping both primary hydroxyl groups as their TBDPS ethers under standard conditions was followed by simple engagement of the 3' and 4' hydroxyl functions as a cyclic carbonate 2a. The stereospecific introduction of two α -linked fucose residues gave tetrasaccharide glycal 3a in 51% yield in a single step. The donor used was the known fluorosugar 5a (S.J. Danishefsky, J. Gervay, J.M. Peterson, F.E. McDonald, K. Koseki, T. Oriyama, D.A. Griffith, C-H. Wong, D.P. Dumas, J. Am. Chem. Soc. 1992, 114, 8329) following a modification of the original Mukaiyama conditions. (T. Mukaiyama, Y. Murai, S. Shoda, Chem. Lett. 1981, 431) Glycal 3a corresponds to the Le^y hapten, lacking the N-acetyl function in the glucose residue. The problem was then to introduce this group as well as a galactose spacer module.

Methodology developed previously (D.A. Griffith, S.J. Danishefsky, "On the Sulfonamidoglycosylation of Glycals. A Route to Oligosaccharides With 2-Aminohexose Subunits+", J. Am. Chem. Soc. 1990 112, 5811) proved appropriate to attain these goals. Glycal 3a was treated with iodonium dicollidine perchlorate and benzenesulfonamide to afford iodosulfonamide 4a. Azaglycosylation using the 3-stannyl ether of galactal (9a) (S.J. Danishefsky, K. Koseki, D.A. Griffith, J. Gervay, J. M. Peterson, F.E. McDonald, T. Oriyama, J. Am. Chem. Soc. 1992, 114, 8331) in the presence of silver tetrafluoroborate gave pentasaccharide glycal 6a in 75% yield as shown in Figure 3. Having 6a in hand, one can iterate the azaglycosylation sequence or activate the glycal as its epoxide and continue with further glycosylations. To demonstrate the ability to fashion a conjugatable form of Le^y hapten, formation of the allyl glycoside was important. The feasibility of converting the sulfonamido group into the target acetamide was demonstrated. Glycal 6a was deprotected in two steps as shown. Peracetylation

afforded acetamido glycal 7a. Activation of the glycal as its epoxide with dimethyldioxirane (R.L. Halcomb, S.J. Danishefsky, J. Am. Chem. Soc. 1989, 111, 6661), followed by epoxide opening with allyl alcohol in the presence of zinc chloride gave the desired peracetylated β -allyl pentasaccharide which was deacetylated by action of methoxide to provide the target Le^y hapten as its β -allyl glycoside 8a. (8a $[\alpha]_D$ -72.7° (c. 1 MeOH); IR (thin film) 3350, 2940, 2900, 2830, 1650, 1550, 1365, 1300, 1155, 1070, 1030; ¹H NMR (400 MHz, CD₃OD) δ 5.95 (m, 1H), 5.32 (d, J =17.25 Hz, 1H), 5.14-5.19 (m, 2H), 5.04 (d, J = 3.83 Hz, 1H), 5.02 (d, J =3.50 Hz, 1H). 4.68 (d, J = 8.15 Hz, 2H), 4.51 (d, J = 5.70 Hz, 1H) 3.40-4.38 (m, 27H). 1.96 (s, 3H), 1.23 (m, 6H); HRMS (FAB) calcd for C₃₅H₅₆NO₂₄Na 900.3325 found 900.3310) The aldehyde, derived by ozonolysis of 8a, could be conjugated to a carrier protein by the method of Bernstein and Hall.

This synthesis is the most direct route to the Le^y determinant known. (O. Hindsgaul, T. Norberg, J. Le Pendu, R. U. Lemieux, Carbohydr Res. 1982, 109, 109; U. Spohr, R.U. Lemieux ibid. 1988, 174, 211; for previous syntheses, see: J.C. Jacquinet, P. Sinay, J. Org. Chem. 1977, 42, 720; S. Nilsson, H. Lohn, T. Norberg, Glycoconjugate J. 1989, 6, 21; R.R. Schmidt, A. Topfer, Tetrahedron Lett. 1991, 32, 3353; W. Kinzy, A. Low, Carbohydrate. Res. 1993, 245, 193) The method is stereospecific at each step, and it illustrates the versatility of glycals both as donors and acceptors and takes advantage of 1,2-glycal epoxides and their presumed N-sulfonylaziridine counterparts. The method also makes possible extensive analog preparation and variation of conjugation strategies.

The synthesis of 3a and 6a are shown below:

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3a: To 2.00g (2.47 mmol) of lactal carbonate 2a was added 4.44g (9.86 mmol) of fucosyl fluoride 5a. The mixture was azeotroped 5 times with benzene and placed under high vacuum for two hours. Under an argon atmosphere 2.77 ml (12.33 mmol) of di-tert-butyl pyridine and 16ml of dry ether were added. 2.0 g of freshly activated 4A molecular sieves were added and the mixture stirred one hour at room temperature. In an argon glove bag, 2.34g (12.33 mmol) of stannous chloride (SnCl_2) and 2.56g (12.33 mmol) of silver perchlorate (AgClO_4) were added. The flask was equipped with a reflux condensor and the reaction brought to reflux for 72 hours. The reaction was quenched with 5ml of saturated bicarbonate and filtered through a pad of celite. Diluted with 50ml ethyl acetate and washed 2 times with sat. bicarbonate, 2 times with sat. copper sulfate and 2 times with sat. brine. The organics were dried over MgSO_4 and concentrated. Flash chromatography in 20% ethyl acetate/hexanes afforded 2.10g (51%) of a white foam 3a: $[\alpha]_D -78.9$ (c.555, CHCl_3); IR (thin film) 3040, 3000, 2905, 2860, 2830, 1820, 1800, 1710, 1635, 1585, 1570, 1480, 1460, 1440, 1415, 1370, 1350, 1300, 1260, 1205, 1145, 1100, 950, 735, 695, ^1H NMR (400MHz, CDCl_3) δ 8.09 (d, $J=8.12$ Hz, 2H), 8.00 (d, $J=8.26$ Hz, 2H) 7.66 (m, 4H), 7.59 (d, $J=6.74$ Hz, 4H), 7.56 (t, $J=7.27$ Hz, 1H), 7.30-7.50 (m, 22H) 7.16-7.26 (m, 10H) 7.09 (m, 2H), 6.99 (t, $J=7.59$ Hz, 2H) 6.89 (t, $J=7.97$ Hz, 1H), 6.43 (d, $J=6.08$ Hz, 1H), 5.46 (bs, 1H), 5.38 (bs, 1H), 5.35 (d, $J=3.42$ Hz, 1H), 4.89 (d, $J=11.35$ Hz, 1H), 4.75-4.80 (m, 4H), 4.72 (d, $J=5.88$ Hz, 2H), 4.69 (d, $J=4.27$ Hz, 2H), 4.36-4.55 (m, 5H), 4.28 (q, $J=6.51$ Hz, 1H), 4.17 (bd, $J=5.46$ Hz, 1H), 3.90-4.00 (m, 6H), 3.85 (d, $J=2.99$ Hz, 1H), 3.82 (d, $J=2.89$ Hz, 1H), 3.56-3.78 (m, 4H), 1.07 (m, 24H); HRMS (FAB): calcd for $\text{C}_{99}\text{H}_{106}\text{O}_{20}\text{Si}_2\text{Na}$ 1694.6740 found 1694.6787.

6a: 230 mg (0.12mmol) of iodosulfonamide 4a was azeotroped 5 times with dry benzene and placed under high

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vacuum for two hours. Added 2.4ml of THF solution of 15eq. of tin ether 9a (generated by azeotropic removal of water overnight with a Dean-Stark trap equipped with freshly activated 4A mol. sieves from 561 mg (1.80mmol) of 6a-TIPS-galactal and 673 μ l (1.32mmol) bis(tributyltin) oxide in 80 ml of benzene). To this solution stirring under an argon atmosphere was added 200 mg of freshly activated 4A powdered molecular sieves. Stirred one hour at room temperature. Cooled solution to -78°C and added, via cannula, a solution of 187 mg (.96mmol) of silver tetrafluoroborate in 2.4 ml of THF. Warmed to room temperature over 15 hours and quenched the reaction, which had turned bright yellow, with 2ml. of sat. bicarbonate. The reaction mixture was filtered through a pad of celite into a separatory funnel. The celite pad was washed thoroughly with ethyl acetate. The organics were washed twice with sat. bicarbonate and twice with sat. brine. The organics were dried over MgSO₄. Concentration and chromatography in 25% ethyl acetate/hexanes gave 193 mg (75%) as a white foam 6a: $[\alpha]_D -126.4^\circ$ (c, 505, CHCl₃), IR (thin film) 3500, 3040, 3000, 2905, 2840, 1820, 1800, 1705, 1635, 1590, 1440, 1410, 1255, 1195, 1100, 1080, 1035, 815, 730, 695; ¹H NMR (400 MHz, CDCl₃) δ 8.09 (app t, 4H), 7.08-7.65 (m, 46H), 6.90 (t, $J=7.65$ Hz, 3H), 6.76 (d, $J=6.91$ Hz, 2H), 6.12 (d, $J=6.59$ Hz, 1H), 5.50 (bs 1H), 5.45 (bs 1H), 5.28 (app t, 2H), 3.03-4.91 (m, 36H), 1.09 (m, 45H); LRMS (FAB): calcd for C₁₂₀H₁₄₁NO₂₆SSi₃Na 2153 found 2153.

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A Strategy for the Assembly of Complex, Branched Oligosaccharide Domains on a Solid Support: An Application to a Concise Synthesis of the Lewis^b Domain in Bioconjugatable Form.

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The assembly of the Le^b (type 1) domain is a relatively more difficult undertaking than was the Le^y (type 2)

target, wherein lactal was used as a convenient starting material. In the case of the type 1 determinant, lactal is not a useful starting material. The synthesis of the Le^b system offered an opportunity to apply the polymer-based oligosaccharide construction method. (S.J. Danishefsky, K.F. McClure, J.T. Randolph, R.B. Ruggeri, Science 1993, 260, 1307) The strategy is summarized in Figure 4, wherein polymer-bound glycal 1 is activated for glycosyl donation via direct formation of a 1,2-anhydro derivative 2. Reaction of 2 with acceptor glycal 3 furnishes 4. Reiteration is achieved by means of direct epoxidation and reaction with acceptor 3. The self-policing nature of the method and the simple "one time" purification at the end of the synthesis are useful features.

The present invention discloses an important additional dimension of the polymer-bound method. The logic is captured by inspection of Figure 5. Each glycosylation event generates a unique C₂ hydroxyl. In principle (and in fact, see infra) this hydroxyl can function as a glycosyl acceptor upon reaction with a solution based donor. The glycal linkage of 5, still housed on the support, can be further elongated. In this way, branching at C₂ is accomplished while minimizing the requirement for protecting group machinations. (For an application of this strategy in the synthesis of a complex saponin, see: J.T. Randolph, S.J. Danishefsky, J. Am Chem Soc. 1993, 115, 8473)

In principle, this branching can be implemented at any site in a growing chain. For such an extension, it would be necessary to cap all previously generated hydroxyl groups generated on the "polymer side" (non-reducing end) of the growing domain. Thus, the polymer-bound oligosaccharide can serve as either donor or acceptor, wherever appropriate.

Initial efforts at reduction to practice identified tetrasaccharide glycal 6, bearing H-type 2 blood group specificity, as a goal. Polymer-supported galactal 7 (using as polymer support polystyrene crosslinked with 1% divinylbenzene functionalized using published procedures: T-H. Chan, W.-Q. Huang, J. Chem. Soc., Chem. Commun. 1985, 909; M.J. Farrall, J.M.J. Frechet, J. Org. Chem. 1976, 41, 3877) reacted with a solution of 3,3-dimethyldioxirane (R.W. Murray, R. Jeyaraman, J. Org. Chem. 1985, 50, 2847), to provide the corresponding 1,2-anhydrosugar glycosyl donor, which was treated with a solution of glucal derivative 8 in the presence of $ZnCl_2$ to provide 9 (R.L. Halcomb, S.J. Danishefsky, J. Am. Chem. Soc. 1989, 111, 6661) This polymer-bound disaccharide acted as a glycosyl acceptor upon treatment with a solution of fucosyl fluoride 10 (K.C. Nicolau, C.W. Hummel, Y. Iwabuchi, J. Am. Chem. Soc. 1992, 114, 3126) in the presence of $Sn(OTf)_2$ thereby giving 11. Retrieval of the trisaccharide glycal from the support was accomplished using tetrabutylammonium fluoride (TBAF) to afford 12 in 50% overall yield from 7.

The trisaccharide, retrieved from the polymer, could then be further elaborated. Toward this end, compound 12 was converted to silyl ether 13 by reaction with TIPSCl. The latter was converted to the iodosulfonamide derivative 14 by the action of $I(coll)_2ClO_4$ in the presence of $PhSO_2NH_2$. Reaction of 14 with galactal stannyl ether derivative 15 in the presence of $AgBF_4$ gave 16 77% yield. (D.A. Griffith, S.J. Danishefsky, J. Am. Chem. Soc. 1990, 112, 5811) Tetrasaccharide glycal 16 was deprotected and peracetylated to afford 6. (S.J. Danishefsky, K. Koseki, D.A. Griffith, J. Gervay, J.M. Peterson, F.E. McDonald, T. Oriyama, J. Am. Chem. Soc. 1992, 114, 8331)

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Thus, the synthesis of the full H-type determinant was achieved by sequential polymer- and solution-based

maneuvers. The next target was the more complex Le^b hexasaccharide 17. The campaign proceeded as shown in Figure 6. Polymer-bound galactal 7 was converted to 18 upon epoxidation with 3,3-dimethyldioxirane followed by
5 reaction with glucal derivative 19. This disaccharide diol was then bisfucosylated using fucosyl donor 10 in the presence of Sn(OTf)₂ to afford 20. Retrieval from the support with TBAF provided 21, which was obtained in 40% overall yield from 7. Compound 21 reacted with TIPSCl to
10 give 22.

Iodosulfonamide 23, obtained from 22 using I(coll)₂ClO₄ and PhSO₂NH₂, reacted with lactal derivative 24 in the presence of AgBF₄ to provide hexasaccharide glycal 25 in
15 55% yield. Deprotection of 25 was accomplished in two stages (TBAF to remove the silyl ethers, followed by Na/NH₃ reduction to remove the aromatic protecting groups), and the crude product was peracetylated to give 26 in a 51% overall yield. Compound 26 was converted,
20 via the 1,2-anhydrosugar derivative, to allyl glycoside 17, which can be activated by ozonolysis to the aldehyde (R = CH₂CHO) for subsequent coupling to a protein by the method of Bernstein and Hall.

25 In sum, the present invention extends the solid-support glycal assembly method for complex carbohydrate domain synthesis to include the branching patterns critical for biorecognition. Specifically, the determinant for the binding of H. pylori to human gastric epithelium has been
30 stereospecifically fashioned, with simplicity, in a way which provides significant relief from some of the complexities of protecting group manipulations.

35 Experimental Procedure:

6: ^1H NMR (400 MHz, CDCl_3); δ 6.39 (d, 1H, $J = 6.2$ Hz, H_1 galactal), 5.65 (d, 1H, $J = 8.9$ Hz, NHAc), 5.35 (d, 1H, $J = 3.8$ Hz), 5.33 (m, 1H), 5.29 (d, 1H, $J = 2.6$ Hz), 5.27 (d, 1H, $J = 3.1$ Hz), 5.17-5.09 (m, 2H), 4.97-4.90 (m, 2H), 4.81 (dd, 1H, $J = 3$ Hz, $J = 6.1$ Hz, H_2 galactal), 4.75 (d, 1H, $J = 8.0$ Hz), 4.52 (m, 1H), 4.48 (dd, 1H, $J = 12.0$ Hz), 4.44-4.06 (m, 8H), 3.88-3.77 (m, 4H), 3.61 (m, 1H), 2.18-1.97 (m, 33 H, COCH_3), 1.18 (d, 3H, $J = 6.5$ Hz, CH_3 fucose); ^{13}C NMR (CDCl_3): δ 170.80, 170.77, 170.72, 170.67, 170.62, 170.34, 170.21, 170.09, 170.01, 169.99, 169.65, 144.92 (C_1 galactal), 100.22, 98.83, 98.58, 95.55, 74.48, 73.38, 73.13, 73.06, 71.48, 71.01, 70.68, 67.97, 67.42, 67.18, 67.05, 65.94, 64.83, 62.35, 62.22, 60.88, 60.37, 54.21, 23.23, 22.15, 20.85, 20.82, 20.79, 20.76, 20.65, 20.61, 20.57, 15.51, (C_6 fucose); IR (thin film): 3368.7 (NH), 2965.6, 2934.6, 1746.5 ($\text{C}=\text{O}$), 1537.5, 1435.9, 1371.3, 1228.5, 1065.0, 1046.0; $[\alpha]_D^{25} = -51.1^\circ$ (c 1.8, CH_2Cl_2); HRMS (FAB); calcd. for $\text{C}_{46}\text{H}_{63}\text{NNaO}_{28}$: $m/z = 1100.3434$, found 1100.3436.

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21: Polymer-bound galactal 7 (loading = 0.85 mmol glycal/g), which had been placed in a round-bottom flask equipped with a fritted outlet, was suspended in CH_2Cl_2 under N_2 , cooled to 0°C , and then treated with a solution of 3,3-dimethyldioxirane. The mixture was stirred (teflon-coated magnetic stir bar) for 40 min. at 0°C , after which time solubles were removed by filtration through the fritted outlet (N_2 pressure). The polymer bound 1,2 anhydrosugar was evacuated (ca. 0.1 torr) for several hours in order to dry the material for the next step. This material was once again placed under N_2 before being treated with 19 (~10 molar equivalents as a 0.5 M solution in THF). The suspension was cooled to -40°C , and treated with ZnCl_2 (~2 molar equivalents as a 1.0 M solution in THF). The reaction mixture was allowed to slowly warm to rt (over ca. 2 h), and then stirred an additional 3-4 h. Solubles were removed by filtration,

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and polymer 18 was washed several times with THF and then dried in vacuo. To compound 18 was added, in a glove bag, solid $\text{Sn}(\text{OTf})_2$ (~ molar equivalents), and the mixture was placed under N_2 and cooled to 0°C before being treated with 10 (~5 molar equivalents as a 0.2 M solution in THF and di-tert-butylpyridine (~8 molar equivalents). The suspension was allowed to warm to rt and stirred 8-10 h. The mixture was rinsed with anhydrous THF (2 times), 1,4-dioxane (2 times), again with THF, and then dried in vacuo. Compound 20 (100 mg) was suspended in THF, treated with a 1:3 mixture of AcOH and TBAF (~0.2 M in TBAF, ~10 molar equivalents), and the mixture was stirred for 18 h at 40°C . The polymer was rinsed with THF (3 times), and the combined rinsings were concentrated and purified by column chromatography on silica gel (1:1 EtOAc: hexanes). Compound 21 (18 mg) was obtained as a colorless solid (40% overall yield from 7): ^1H NMR (400 MHz, CDCl_3): δ 7.40-7.25 (m, 30H, Ar H), 6.18 (d, 1H, $J=6.0$ Hz, H_1 glucal), 5.26 (d, 1H, $J=3.5$ Hz, H_1 fucose), 5.09 (d, 1H, $J=3.7$ Hz, H_1 fucose), 4.96 (t, 2H, $J=10.8$ Hz, PhCH_2), (4.90-4.56 (m, 13 H), 4.43 (m, 1H), 4.15-4.06 (m, 4 H), 3.97 (dt, 1H, $J=8.3$ Hz, $J=2.4$ Hz), 3.87-3.65 (m, 10H), 3.64 (d, 1H), 3.57 (d, 1H), 2.69 (br, 1H, OH), 2.52 (br, 1H, OH), 1.11 (d, 3H, $J=7.0$ Hz, CH_3 fucose), 1.09 (d, 3H, $J=7.0$ Hz, CH_3 fucose); ^{13}C NMR (CDCl_3): δ 153.37 (C=O), 145.75 (C_1 glucal), 138.60, 138.52, 138.19, 137.61, 128.55, 128.52, 128.44, 128.24, 128.16, 128.07, 127.62, 127.56, 127.45, 98.71, 98.38, 97.65, 97.34, 79.26, 78.87, 78.67, 78.01, 77.79, 77.65, 76.37, 76.10, 74.92, 74.40, 74.16, 73.95, 72.86, 72.64, 72.53, 67.43, 67.29, 61.31, 60.90, 16.65 (C_6 fucose), 16.53 (C_6 fucose); IR (thin film): 3467.0 (OH), 3029.6, 2923.6, 1807.2 (C=O), 1647.3, 1496.0, 1453.5, 1358.1, 1240.2, 1095.6, 1049.2, 738.5, 697.2; $[\alpha]_{\text{D}23} = -82.5^\circ$ (c 0.4, CH_2Cl_2); HRMS (FAB); calcd. for $\text{C}_{67}\text{H}_{74}\text{NaO}_{18}$: $m/z = 1189.4772$, found 1189.4757.

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25: To a mixture of 23 (60 mg, 34 μ mol) and powdered 4A molecular sieves (200 mg) under N_2 was added, via canula, a solution of 24 (0.21 mmol) in anhydrous THF (1.5 mL). The stirred suspension was cooled to $-78^\circ C$ before being
5 treated with a solution of $AgBF_4$ (0.21 mmol) in 0.25 mL of anhydrous THF. The mixture was stirred and allowed to slowly warm to rt overnight. The suspension, which had developed a bright-yellow color, was heated, with stirring, at $45^\circ C$ for an additional 36 h, until the TLC
10 (2.5 EtOAc:hexanes) showed no trace of 23. The mixture was treated with saturated aqueous NH_4Cl (5 mL) and then extracted with EtOAc (3 x 10 mL), and the organics were dried over $MgSO_4$. The crude product was purified by silica gel chromatography (1:3 EtOAc:hexanes) to give 25
15 as a colorless oil (42 mg, 55%): 1H NMR (400 MHz, acetone- d_6): δ 8.17(d, 2 H, $J=7.3$ Hz, $PhSO_2$), 7.50-7.20 (m, 33H, ArH), 6.52 (d, 1 H, $J=10.5$ Hz, NH), 6.30 (dd, 1 H, $J=6.0$ Hz, H_1 glucal), 5.35-5.32 (m, 2H), 5.25 (d, 1H, $J=7.9$ Hz), 5.15 (m, 2 H), 4.99-4.92 (m, 3H), 4.86-4.52
20 (m, 14 H), 4.45 (dd, 1H, $J=7.91$ Hz, $J=2.4$ Hz), 4.32-4.23 (m, 3H), 4.22 (dd, 1 H), 4.17 (d, 1 H, $J=10.1$ Hz), 4.08-3.84 (m, 18 H), 3.79-3.73 (m, 2H), 3.66 (m, 1H), 3.55 (t, 1 H, $J=6$ Hz), 3.50 (dd, 1 H, $J=9.7$ Hz), 1.33 (d, 3 H, $J=6.5$ Hz, CH_3 fucose), 1.31 (d, 3H, $J=6.4$ Hz, CH_3
25 fucose), 1.20-0.98 (m, 84 H, 3 x $Si(i-Pr)_3$); ^{13}C NMR (acetone- d_6): 145.66 (C=O), 132.72, 131.48, 131.45, 131.28, 131.16, 130.77, 130.48, 121.31, 120.11, 119.86, 119.78, 119.25, 95.63, 94.70, 91.37, 89.64, 89.31, 86.52, 73.38, 72.24, 71.00, 70.71, 70.37, 69.80, 69.59, 69.06,
30 68.23, 67.92, 67.38, 67.10, 66.49, 65.67, 65.33, 64.60, 64.34, 64.03, 63.45, 63.30, 59.46, 58.83, 58.37, 54.45, 53.32, 49.86, 19.67 (C_6 fucose), 18.42 (C_6 fucose), 9.55, 9.48, 9.45, 9.31, 9.23, 3.82, 3.70, 3.64; IR (thin film): 3491.9 (OH), 3030.1, 2941.2, 2865.5, 1835.8, 1819.5,
35 1649.8, 1496.2, 1462.3, 1349.9, 1245.5, 1155.2, 1095.1, 1049.4, 882.2, 734.8, 692.0; $[\alpha]_{D23} = -33.8^\circ$ (c 2.0,

CH₂Cl₂); HRMS (FAB): calcd for ¹²C₁₂₀¹³CH₁₇₉NNaO₂₉SSi₄: m/z = 2278.1292, found 2278.1296.

17: ¹H NMR (400 MHz, CD₃OD): δ 6.00 (m, 1H, J = 5.6 Hz, CH₂CH=CH₂), 5.37 (dd, 1 H, J = 1.6 Hz, J = 7.3 Hz, CH₂CH=CH₂), 5.20 (dd, 1 H, J = 1.6 Hz, J = 9.5 Hz, CH₂CH=CH₂), 5.18 (d, 1 H, J = 3.9 Hz, H₁ fucose), 5.10 (d, 1H, J = 3.8 Hz, H₁ fucose), 4.64 (d, 1 H, J = 6.9 Hz), 4.45 (d, 1H, J = 7.4 Hz), 4.43-4.23 (m, 2H), 4.27 (dd, 1H, J = 9.3 Hz, J = 10.6 Hz), 4.23-4.11 (m, 2H), 4.02-3.29 (m, 31 H), 2.06 (s, 3H, NAc), 1.31 (d, 3H, J = 6.6 Hz, CH₃ fucose), 1.29 (d, 3 H, J = 6.6 Hz, CH₃ fucose); ¹³C NMR (CD₃OD): δ 173.20 (C=O), 135.73 (CH₂CH=CH₂), 105.13, 103.30, 102.49, 101.62, 99.63, 96.86, 80.79, 80.67, 78.44, 76.67, 76.49, 75.89, 74.80, 74.59, 73.94, 73.61, 73.40, 71.55, 71.38, 71.16, 70.42, 70.26, 70.14, 67.77, 67.30, 67.21, 62.79, 62.34, 61.99, 55.54, 22.97 (NAc), 16.65 (2 C's, C₆ fucose); IR (thin film): 3376.6 (OH), 2924.2, 1652.5 (C=O), 1383.1, 1032.4; [α]_{D25} = -12.8° (c 0.25, MeOH); HRMS (FAB): calcd. for C₄₁H₆₉NNaO₂₉: m/z = 1062.3853, found 1062.3837

Glycal Assembly Method Applied to the Synthesis of Human Breast Tumor-Associated Antigen

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The present invention provides a convergent synthesis of the hexasaccharide wherein the two trisaccharide domains have been efficiently assembled in forms readily amenable for coupling. The synthesis of the ABC trisaccharide is presented in Figure 8. The α-linkage of this trisaccharide might be formed by employing a fluorosugar donor 4b, using established conditions. (Gordon, D. M.; Danishefsky, S. J., Carbohydr. Res., 1990, 206, 361-366.) Preparation of the appropriate disaccharide acceptor commenced with 5b (Danishefsky, S. J.; Behar, V.; Randolph, J. T.; Lloyd, K. O., J. Am. Chem. Soc., 1995, 0000), itself obtained from a glycal coupling.

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Benzylation followed by desilylation, carbonate removal and selective dibenylation afforded the disaccharide 6b. The acceptor thus obtained was reacted with the fluorosugar 4b using modified Mukaiyama conditions (Mukaiyama, T.; Murai, Y.; Shoda, S., Chem. Lett., 1981, 431-433) to provide the trisaccharide glycal 7b. Deprotection of the PMB ether provided the ABC trisaccharide 8b, which was poised for coupling with a suitable DEF trisaccharide donor.

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The synthesis of the DEF trisaccharide is described in Figure 9. Epoxidation of the galactal 9b and standard coupling (Halcomb, R.L.; Danishefsky, S.J., J. Am. Chem. Soc., 1989, 111, 6661-6666.) with acceptor 10b afforded, regioselectively, the disaccharide 11b. Fucosylation employing the fluoro-fucose 12b (Dejter-Juszynski, M.; Flowers, H.M., Carbohydr. Res., 1973, 28, 61) provided a 5:1 ratio of monoglycosylation regioisomers, the major isomer being the desired trisaccharide 13b. This material was treated under standard conditions to afford the trans-diaxial iodosulfonamide 14b.

Direct coupling reactions (Griffith, D.A.; Danishefsky, S.J., J. Am. Chem. Soc., 1990, 112, 5811-5819; Danishefsky, S.J.; Koseki, K.; Griffith, D.A.; Gervay, J.; Peterson, J.M.; McDonald, F.E.; Oriyama, T., J. Am. Chem. Soc., 1992, 114, 8331-8333) employing iodosulfonamides such as 14b with ABC trisaccharide acceptors failed, leading to a different donor functionality in the trisaccharide. In practice, the iodosulfonamide 14b was treated with excess lithium ethanethiolate to afford the ethyl thioglycoside 15b (Figure 10). Precedent established by the present inventors lead to the prediction of sulfonamide participation to provide the desired β -linked product from 15b. (Griffith, D.A., Ph.D. Thesis, Yale University, 1992) When donor 15b was treated with MeOTf in the

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presence of acceptor 8b, a 10:1 mixture of hexasaccharide isomers was obtained. The major product 16b was obtained in 70-85% yield.

5 Ceramide attachment and elaboration commenced with
epoxidation of 16b, followed by reaction with the stannyl
ether 17b promoted by $\text{Zn}(\text{OTf})_2$. (Liu, K.K.-C.;
Danishefsky, S.J., J. Am. Chem. Soc., 1993, 115, 4933-
4934) Although the yield of this ceramide coupling is
10 low, when this reaction was performed on trisaccharide
7b, the corresponding product was obtained in 66% yield.
This material can then be used to obtain 18b. Following
acetylation, the ceramide side-chain was elaborated by
reduction of the azide functionality using Lindlar's
15 catalyst under an atmosphere of H_2 in the presence of
palmitic anhydride to provide 18b. Desilylation and
saponification was followed by dissolving metal
deprotection and MeOH quench. Peracetylation of the
crude mixture, followed by saponification provided the
20 glycosphingolipid 1b. Only the chemical shifts and
coupling constants of the anomeric protons have been
reported for the natural material. The spectrum of
synthetic 1b is in complete agreement with this data.
Furthermore, the product was characterized by exact mass,
25 and ^1H and ^{13}C NMR. The synthetic material has also been
shown to bind to the monoclonal antibody MBrl.

In addition, the present invention provides the
corresponding allyl glycoside (Figure 11). Deprotection
30 of 16b, as above, and acetylation afforded the peracetate
of the hexasaccharide glycal. Epoxidation, reaction with
allyl alcohol, and saponification provided the allyl
glycoside 19b.

35 As in the case of the Le determinant, ozonolysis of the
allyl group of 19b will set the stage for reductive
coupling to lysine residues of proteins.

Synthesis of 3b:

3-O-(4-Methoxybenzyl)-D-galactal

A suspension of D-galactal (2b) (3.70g, 25.3 mmol) and dibutyltin oxide (6.30g, 1.0 equiv) in dry benzene (150 mL) was heated to reflux for 2h with azeotropic removal of water. The reaction was cooled and treated with PMBCl (3.80 mL, 1.1 equiv) and tetrabutylammonium bromide (9.10g, 1.1 equiv) and refluxed for 4h. The reaction was filtered through silica column and eluted with EtOAc/hexanes (4:1). Fractions containing product were concentrated and the residue triturated in hexanes to give 4.50g (67%) of product as white crystalline solid.

mp (hexanes) 117-118 °C; $(\alpha)^{25}_D = -23.0^\circ$ (CHCl_3 , $c = 1.1$); IR (KBr) 3313 (br), 1645, 1513, 1228, 1082, 821 cm^{-1} ; ^1H -NMR (400 MHz, CDCl_3) δ 7.28 (2H, d, $J = 8.4$ Hz), 6.89 (2H, d, $J = 8.4$ Hz), 6.44 (1H, dd, $J = 6.4$ Hz), 4.70 (1H, dt, $J = 6.3, 1.9$ Hz), 4.59-4.52 (2H, ABq, $J = 11.4$ Hz), 4.20-4.18 (1H, m), 4.04-3.97 (1H, m), 3.90-3.82 (2H, m), 3.81 (3H, s), 2.73 (1H, d, $J = 3.1$ Hz, C4-OH), 2.54 (1H, dd, $J = 8.2, 4.2$ Hz, C6-OH); ^{13}C -NMR (100 MHz, CDCl_3) δ 159.46, 145.02, 142.05, 129.46, 113.95, 99.36, 76.12, 70.17, 70.14, 63.65, 62.74, 55.26; LRMS(NH_3) 284 ($\text{M} + \text{NH}_4$) $^+$, 266 (M) $^+$, 249.

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4,6-di-O-benzyl-3-O-(4-methoxybenzyl)-D-galactal (3b).

A solution of 3-O-(4-methoxybenzyl)-D-galactal (2.28 g, 8.56 mmol) and benzyl bromide (3.75 mL, 3.68 mol equiv; freshly passed through basic alumina) in DMF (30 mL) under N_2 at 0 °C was treated with NaH (1.37 g, 4.0 mol equiv) in two portions. The reaction was stirred 0.5h at 0 °C and 1h at rt. The reaction was carefully poured into 50 g of crushed ice, diluted to 100 mL with water, then extracted with EtOAc-hexanes (1:1, 100 mL x 3). Organic extracts were washed with water (100mL x 2), dried (Na_2SO_4) and concentrated. Flash chromatography with 15%

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EtOAc-hexanes gave 3.58 g (94%) of the title compound as a clear liquid.

$[\alpha]_D^{25} = -48.2^0$ (CHCl_3 , $c = 0.85$); IR (neat) 3030, 2867, 1645, 1613, 1513 1247, 1092, 821, 736 cm^{-1} ; $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 7.34-7.23(12H, m), 4.62 (1H, d, $J = 12.0$ Hz), 4.59-4.51 (2H, ABq, $J = 11.7$ Hz), 4.50-4.39 (2H, ABq, $J = 11.9$ Hz) $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 159.04, 143.99, 138.30, 137.90, 130.43, 128.26, 128.20, 128.03, 127.77, 127.57, 127.56, 113.67, 100.00, 75.58, 73.28, 73.17, 71.13, 70.42, 70.28, 68.35, 55.15; LRMS (NH_3) 464 ($\text{M} + \text{NH}_4^+$, 100), 326 (18), 309 (48), 253 (17).

Synthesis of 4b:

A solution of galactal 3b (3.20g, 7.17 mmol) in dry CH_2Cl_2 under N_2 at 0°C was treated with dimethyldioxirane (0.09M, 80 mL) and stirred until all of the glycal was consumed (0.5-1h; TLC 30% EtOAc in hexanes). Volatiles were removed at 0°C with stream of dry N_2 . The residue was dissolved in 30 mL of dry THF under N_2 at 0°C and treated TBAF (36 mL, stored over molecular sieves) then stirred at ambient temperature for 20h. The dark brown solution was filtered through a pad of silica (~4cm depth) and washed with EtOAc (200 mL). The filtrate was washed with water (200 mL x 3) and dried (MgSO_4) and concentrated. The residue was redissolved in 30% EtOAc-hexanes (50 mL) and filtered through short silica column (10 cm d x 4 cm h) and washed with the same solvent system (1L). The filtrate was concentrated to give 2.59g of fluorohydrin with >90% purity. The residue was dissolved in dry DMF (30 mL) under N_2 at 0°C and treated with benzyl bromide (958 μL , 1.5 equiv, freshly filtered through basic alumina), finally with NaH (322 mg, 60% dispersion, 1.5 equiv) and stirred for 30 min at 0°C and 30 min at rt. The reaction was quenched by pouring into 100g of ice, and extracted with 1:1 EtOAc-hexanes (150 mL x 2). The organic extracts were washed with water (150 mL x 2),

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dried (MgSO_4) and concentrated in vacuo. Flash chromatography with 10% EtOAc-hexanes gave 2.00g (49%) of the title compound as a yellowish liquid.

- 5 $[\alpha]_D^{25} = +15.3^0$ (CHCl_3 , $c = 0.85$); IR (CHCl_3 film) 2916, 1612, 1513, 1248, 1103, 1056, 734 cm^{-1} ; $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 7.35-7.24 (17H, m), 6.84 (2H, d, $J = 8.4$ Hz), 5.15 (1H, dd, $J = 53.2, 7.0$ Hz), 4.92 (1H, d, $J = 11.6$ Hz), 4.48-4.74 (2H, ABq, $J = 11.8$ Hz), 3.96-3.89 (1H, m),
10 3.86 (1H, br s), (3H, s), 3.65-3.56 (3H, m), 3.51 (1H, dd, $J = 9.8, 2.8$ Hz); $^{13}\text{C-NMR}$ (100 MHz, CDCl_3) δ 159.22, 138.33, 138.11, 137.62, 130.16, 129.19, 128.40, 128.29, 128.21, 128.04 (2C), 127.90, 127.81, 127.69, 127.59, 113.77, 110.20 (d, $J = 214$ Hz), 80.60 (d, $J = 11.3$ Hz),
15 79.00 (d, $J = 20.5$ Hz), 74.92, 74.52, 73.59 (d, $J = 5.0$ Hz), 73.54, 72.99, 72.70, 68.34, 55.20; LRMS (NH_3) 454 ($M + \text{NH}_4^+$, 100).

Synthesis of 6b:

- 20 A solution of TIPS-carbonate galactal 5b (Danishefsky, S.J.; Behar, V.; Randolph, J.T.; Lloyd, K., J. Am. Chem. Soc., 1995, 0000) (4.28g, 5.62 mmol) in THF (25mL)-MeOH (5 mL) was treated with TBAF solution (1.0M, 6.75 mL, 1.2 equiv). After 6h, additional TBAF (4mL) was added and
25 stirred additional 3h. The reaction was concentrated and directly chromatographed with 4:1 EtOAc-hexanes to obtain 2.20g of the triol. Remaining mixtures of cyclic carbonate and mixed carbonate was hydrolysed in MeOH with MeONa (1.0 mL, 25 wt%) and purified chromatographically.
30 Total yield was 3.02g (93%). This material was directly used for the dibenzylation step.

- $^1\text{H-NMR}$ (400 MHz, CDCl_3) δ 7.35-7.24 (15H, m), 6.43 (1H, d, $J = 6.3$ Hz), 4.87 (1H, dd, $J = 6.3, 3.4$ Hz), 4.84 (1H, d, $J = 11.4$ Hz), 4.63 (2H, apparent s), 4.61 (1H, d, $J = 11.4$ Hz), 4.53-4.47 (3H, m), 4.19-4.16 (3H, m), 3.87-3.84 (2H, m), 3.78-3.66 (3H, m), 3.46 (2H, apparent d, $J = 4.6$

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Hz), 3.29 (1H, t, J = 5.5 Hz), 3.08 (1H, br), 2.73 (2H, br); ¹³C-NMR (100 MHz, CDCl₃) δ 144.70, 138.41, 138.22, 137.83, 128.45, 128.33 (2C), 128.12, 127.84, 127.73, 127.64, 127.57, 102.28, 99.74, 78.99, 76.03, 74.64, 74.07, 73.24 (2C), 73.17, 72.64, 70.20, 69.10, 67.79, 62.15.

A mixture of triol glycal from above (2.95g, 5.1 mmol), dibutyltin oxide (1.33g, 1.05 equiv) and bistrabutyltin oxide (1.69 mL, 0.65 equiv) in dry benzene (50 mL) under N₂ was refluxed for 5h with azeotropic removal of water. The reaction was cooled below boiling and treated with benzyl bromide (2.43 mL, 4.0 mol equiv) and tetrabutylammonium bromide (3.29g, 2.0 equiv). 10 mL of benzene was distilled off and the reaction refluxed for 16h. The reaction was directly loaded on silica column and eluted with 15-20% EtOAc-hexanes to give 3.48g (90%) of product **6b** as a clear oil.

[α]_D²³ = -3.3° (CHCl₃, c = 0.87); IR (CHCl₃ film) 2867, 1652, 1454, 1364, 1097, 736 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ 7.35-7.21 (25H, m), 6.45 (1H, d, J = 6.2 Hz), 4.88 (1H, dd, J = 6.2, 3.9Hz), 4.83 (1H, d, J = 10.9 Hz), 4.69 (2H, apparent s), 4.68 (1H, d, J=10.9Hz), 4.59 (2H, apparent s), 4.55 (1H, d, J = 7.8 Hz), 4.49 (2H, apparent s), 4.47 (2H, apparent s), 4.29 (1H, dd, J = 9.6, 5.8 Hz), 4.18 (1H, t, J = 4.4 Hz), 4.13 (1H, m), 3.99 (1H, br s), 3.85 (1H, dd, J = 10.6, 6.4 Hz), 3.75-3.60 (4H, m), 3.47-3.41 (2H, m); ¹³C-NMR (100 MHz, CDCl₃) δ 144.43, 138.64, 138.42, 137.99, 137.84, 137.80, 128.40, 128.34, 128.26, 128.23, 128.18, 128.15, 127.82, 127.75, 127.69, 127.67, 127.65, 127.55, 127.51, 127.46, 127.31, 102.56, 99.56, 80.57, 78.69, 75.72, 75.10, 73.57, 73.32, 72.94, 72.28, 71.94, 70.12, 68.90, 67.85, 66.62; LRMS (NH₃) 776 (M + NH₄⁺, 100).

Synthesis of **7b**:

Lactal 6b (1.32g, 1.74 mmol, 1.0 equiv) and fluoro sugar 4b (1.49g, 2.60 mmol, 1.5 equiv) were combined in ether and concentrated. The mixture were dried by evaporation in dry benzene (25 mL x 2), in vacuum for 2h then treated with di-t-butylpyridine (389 uL, 1.0 equiv) in glove bag and dissolved in dry ether (18 mL) under nitrogen atmosphere. In a separate 50 mL flask was placed 4A M.S. (4.0g) then flame-dried under vacuum, cooled to room temperature. Anhydrous silver perchlorate (360 mg, 1.0 equiv) and SnCl₂ (329 mg, 1.0 equiv) were added in glove bag and flushed with nitrogen. The salt mixture was placed in water bath and sugar solution was introduced via double tipped needle and the mixture sonicated for 2 min. The reaction was wrapped with aluminum foil and stirred for 45h at rt. The filtrate (200 mL) was washed with dil NaHCO₃ (100 mL x 2), dried (MgSO₄) and concentrated. Flash chromatography with 15-20 % EtOAc/hexanes yielded trisaccharides (1.107g, 49%) and impure lactal. The trisaccharide portion was rechromatographed with 2% ether-methylene chloride to give 879 mg (39%) of the desired α -product and 195 mg (8.6%) of β -product. The impure lactal fraction was rechromatographed with 3-4% ether-methylene chloride to give 479 mg (36%) of clean lactal. 77% of coupling (61% α -product) yield based on recovered starting material.

$[\alpha]_D^{25} = +41.8^{\circ}$ (CHCl₃, c = 1.8); IR (CHCl₃ film) 2867, 1648, 1513, 1496, 1453, 1364, 1248, 1097, 735 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ 7.33-7.12 (42H, m) 6.83 (2H, d, J = 8.4 Hz), 6.45 (1H, d, J = 6.0 Hz), 5.03 (1H, d, J = 2.3 Hz), 4.91-4.76 (6H, m), 4.68-4.40 (12H, m), 4.23-3.97 (11H, m), 3.86-3.82 (1H, dd, J = 2.3 Hz), 3.76 (3H, s), 3.69-3.64 (2H, m), 3.53 (1H, t, J = 8.7 Hz), 3.47-3.43 (1H, m), 3.40-3.36 (1H, m), 3.34-3.31 (1H, dd, J = 9.9, 2.8 Hz), 3.22 (1H, dd, J = 8.3, 4.8 Hz); ¹³C-NMR (100 MHz, CDCl₃) δ 158.93, 144.59, 138.98, 138.84, 138.78, 138.64, 138.58, 138.06, 138.02 (2C), 130.82, 129.04, 128.33,

128.24, 128.21, 128.15, 128.08, 128.05, 127.83, 127.81, 127.72, 127.64, 127.58, 127.55, 127.50, 127.44, 127.41, 127.36, 127.33, 127.31, 113.65, 103.02, 100.39, 100.01, 80.93, 78.93, 78.70, 76.53, 76.11, 75.14, 74.84, 74.79, 5 74.35, 73.91, 73.59, 73.36, 73.15, 73.10, 72.98, 72.15, 72.10, 71.99, 70.55, 69.25, 67.92 (2C), 67.69, 55.19.

Synthesis of 8b:

A solution of PMB-trisaccharide (37 mg, 0.028 mmol) in 10 CH_2Cl_2 (1 mL) at 0°C . The reaction was directly loaded on silica column and eluted with 20% EtOAc-hexanes to give 28 mg (84%) of desired product.

[α] $^{25}_D$ = +45.6 $^\circ$ (CHCl_3 , c = 1.78); IR (CHCl_3 film) 2866, 15 1648, 1496, 1453, 1248, 1097, 735 cm^{-1} ; ^1H -NMR (400 MHz, CDCl_3) δ 7.36-7.15 (40H, M), 6.43 (1H, d, J = 6.2 Hz), 5.09 (1H, d, J = 3.3 Hz), 4.85 (1H, dd, J = 6.2, 3.6 Hz), 4.83-4.65 (5H, m), 4.61-4.41 (9H, m), 4.29-4.08 (8H, m), 4.02 (1H, d, J = 2.6 Hz), 3.97 (1H, d, J = 2.2 Hz), 3.93 20 (1H, t, J = 8.4 Hz), 3.86-3.78 (2H, m), 3.67-3.61 (2H, m), 3.53 (1H, dd, J = 8.5, 4.8 Hz); ^{13}C -NMR (100 MHz, CDCl_3) δ 144.38, 138.78, 138.62, 138.47, (2C), 138.20, 138.00, 137.88, (2C, 128.31, 128.29, 128.23, 128.19, 128.16, 128.05, 127.88, 127.83, 127.62, 127.57, 127.49, 25 127.45, 127.43, 127.41, 127.37, 127.32, 127.23, 102.68, 99.89, 99.34, 80.82, 78.72, 77.49, 77.10, 75.88, 75.13, 75.03, 74.23, 73.62, 73.05, 73.01, (3C), 72.62, 72.19 (2C), 70.46, 69.66, 68.92, 67.85, 67.74, 67.54.

30 Synthesis of 11b:

Glycal 9b (4.32 g, 3.14 mmol) was dissolved in CH_2Cl_2 (20 ml) and cooled to 0°C . It was then treated with dimethyldioxarane (219 ml, 3.14 mmol) at 0°C . The epoxidation finished within 1 h and then the reaction 35 mixture was concentrated to dryness using dry N_2 stream. The residue was further azeotroped once with benzene (20 ml) and put on a vacuum line for 30 min at 0°C before

being dissolved in THF (60 ml) and cooled to -78°C . Into the above solution was added, via canula, azeotropically dried galactal 10b (3.32 g, 10.95 mmol, 20 ml THF) and followed by ZnCl_2 (26.3 ml, 1.0 M in ether). The reaction mixture was warmed up to room temperature and stirred overnight. After treatment with sat'd aq. Na_2CO_3 (40 ml), the reaction mixture was concentrated and extracted with ether (500 ml). The organic phase was washed with sat'd aq. NaCl , dried (MgSO_4) and concentrated. The crude product was purified by silica gel chromatography (1:4 EtOAc-hexanes) to give 6.20 g of 11b as a white foam (87.4%).

IR (CHCl_3 film) ν_{max} cm^{-1} ; ^1H -NMR (400 MHz, CDCl_3) δ 6.45 (1H, dd, $J = 6.4, 1.6$ Hz), 4.85 (1H, dd, $J = 6.4, 2.0$ Hz), 4.72-4.68 (2H, m), 4.65 (1H, d, $J = 7.2$ Hz), 4.55 (1H, m), 4.21 (1H, m), 4.08 (1H, dd, $J = 9.6, 5.6$ Hz), 3.96-3.82 (6H, m), 3.33 (1H, d, $J = 3.2$ Hz, OH), 3.27 (1H, d, $J = 2.8$ Hz, OH), 1.16-1.04 (42H, m); ^{13}C -NMR (100 MHz, CDCl_3) δ 154.45, 145.75, 99.14, 98.27, 77.83, 76.59, 74.27, 72.04, -71.62, 70.86, 64.52, 62.57, 61.60, 17.84, 11.78, 11.77. ; LRMS (NH_3) 664 ($M + \text{NH}_4^+ \cdot 100$), 647 ($M + 1^+, 5$), 422 (21), 380 (25).

25 Synthesis of 13b:

Disaccharide 11b (2.64 g, 4.08 mmol) was azeotropically dried three times (3 x 10 ml) together with fluoro-fucose 12b (1.64 g, 3.77 mmol) and molecular sieves (4 A, 4.0 g) in THF (20 ml) with 2,6-di-tert-butylpyridine. The solution was added via canula to a flask containing AgClO_4 (1.56 g, 7.54 mmol), SnCl_2 (1.43 g, 7.54 mmol) and molecular sieves (4 A, 4.0 g) in THF (15 ml) at -40°C . The reaction mixture was stirred 30 min at -40°C and then 34 h at 5°C until the disappearance of fluoro-fucose. After treatment with sat'd aq. NaHCO_3 (40 ml) at 5°C , the mixture was extracted with EtOAc (700 ml). The organic phase was washed with sat'd NaCl , dried (MgSO_4) and

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concentrated. The crude product was purified by silica gel chromatography to give 1.93 g of the desired trisaccharide glycal 13b (48%, based on fluoro-fucose used) and 500 mg of the recovered disaccharide with only a trace of the other monofucosyl product.

Synthesis of 15b:

An azeotropically dried mixture of the trisaccharide glycal 13b (1.11 g, 1.05 mmol) and benzensulphonamide (0.82 g, 5.24 mmol) was dissolved in the THF (20 ml) together with molecular sieves (4 A, 2.6 g). The mixture was cooled to -40°C and then was added, via canula, a solution of $\text{I}(\text{sym-coll})_2\text{COI}_4$ prepared in situ by stirring I_2 (0.54 g, 2.09 mmol) with $\text{Ag}(\text{sym-coll})_2\text{COI}_4$ (0.986 g, 2.29 mmol) in THF (20 ml) at room temperature for about 30 min until the disappearance of the brown color of I_2 . The mixture was warmed up to 0°C within 1 h and stirred for another 1 h. After quenching with sat'd aq. $\text{Na}_2\text{S}_2\text{O}_3$, the mixture was filtrate and extracted with EtOAc (3 x 100 ml). The combined organic phase was washed with sat'd aq. CuSO_4 (100 ml), sat'd NaCl (100 ml x 2) and dried (Na_2SO_4). After concentration, the crude product was purified by silica gel chromatography (1:4 EtOAc-hexanes) to give 981 mg of a colorless oil as a 2:1 mixture of the desired α -trans-diaxial iodosulphonamide and its cis isomer. The iodosulphonamide mixture was then added with stirring into a flask containing ethanthiol (226.3 mg, 3.64 mmol) and lithium hexamethydisilylazide (1.46 ml, 1.46 mmol) in DMF (10 ml) at -40°C . The reaction mixture was stirred at -40°C overnight, and then quenched with sat'd aq. NaHCO_3 and extracted with ether (3 x 100 ml). The combined organic phase was washed with sat'd aq. NaCl and dried (Na_2SO_4). After concentration, the crude product was purified by silica gel chromatography (3:97 EtOAc- CHCl_3) to yield 438 mg of 15b (33%) and 333 mg of the intact cis iodosulphonamide.

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Synthesis of 16b:

A mixture of acceptor trisaccharide 8b (92 mg, 0.077 mmol, 1.0 equiv), thioglycoside 15b (198 mg, 2.0 equiv) and freshly activated 4A-MS (560 mg) under N₂ at rt was
5 suspended in CH₂Cl₂-Et₂O (1:2, 3.9 mL) and stirred for 10 min. The reaction was cooled to 0°C, then treated with methyl triflate (52.4 uL, 6.0 equiv). The reaction was stirred for 4.5h at 0°C and 1.5h while warming to 15°C. The reaction was quenched with TEA(1.0 mL), filtered
10 through a pad of silica and rinsed with Et₂O. The filtrate (70 mL) was washed with sat'd NaHCO₃ (50 mL x 2), dried (Na₂SO₄) and concentrated. The crude product was purified by HPLC (17% EtOAc in hexanes, 15 mL/min, 260 nm UV dection) to give 158 mg (85%) of the desired product
15 and 27.7 mg of α -linked byproduct (ca 55% purity).

Retention time=22 min; $[\alpha]_D^{23} = -13.3^0$ (CHCl₃, c = 1.4); IR (CHCl₃ film) 2940, 2865, 1792, 1652, 1454, 1161, 1101, 734 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ 7.8 (2H, m), 7.38-7.06 (58H, m), 6.43 (1H, d, J = 6.1 Hz), 5.15 (1H, br s), 5.07 (1H, d, J = 3.6 Hz), 5.03 (1H, d, J = 3.6 Hz), 4.99 (1H, d, J = 11.6 Hz), 4.89-4.61 (12H, m), 4.54-4.46 (4H, m), 4.42 (2H, app s), 4.38 (1H, d, J = 11.9 Hz), 4.34-4.26 (3H, m), 4.21-4.18 (4H, m), 4.13-4.03 (7H, m), 3.98-3.76 (14H, m), 3.70-3.61 (4H, m), 3.46-3.27 (7H, m), 2.84 (1H, OH), 1.16 (3H, d, J = 6.4 Hz), 1.13-1.02 (42H, m); ¹³C-NMR (100 MHz, CDCl₃) δ 155.35, 144.55, 140.78, 138.99, 138.75, 138.68, xxx, 138.54, 138.43, 138.13, 138.03, 137.94, 137.82, 132.31, 128.81, 128.52, xxx, 128.38, 128.36, 128.27, 128.24, 128.20, 128.16, 128.02, 127.93, 127.72, 127.66, 127.58, 127.48, 127.43, 127.37, 127.20, 103.41, 102.75, 99.79, 99.55, 98.29, 97.76, 80.49, 80.39, 79.09, 78.91, 78.25, 77.68, xxx, 76.51, 75.88, 75.09, 74.99, 74.91, 74.73, 74.15, 74.02, 73.92, 73.52, 73.19, 73.10, 72.94, 72.67, 72.25, 72.07, 71.76, 71.56, 71.33, 70.33, 69.45, 69.32, 68.48, 68.08, 67.86, 67.75, 61.97, 61.60, 56.14, 17.99, 17.96, 17.95, 17.92, 16.75, 11.86; HRMS

(FAB) calcd for $C_{138}H_{169}NO_{30}SSi_2Na$ (M + Na) 2432.0920, found 2432.0970.

Synthesis of 19b:

5 A solution of hexasaccharide glycal 16b (85 mg, 0.035 mmol) in THF (6mL) under N_2 at rt was treated with TBAF (1.0 M, 353 uL, 10 equiv). After 38h at rt, the reaction was concentrated to ca 1 mL, then dissolved in EtOAc (60 mL), washed with water (30 mL x 2), dried (Na_2SO_4) and
10 concentrated. Flash chromatography with 4% MeOH in CH_2Cl_2 gave 70.0 mg (98%) of the desilyl-decarbonated product.

$[\alpha]_D^{25} = 1.8^0$ ($CHCl_3$ film) 2868, 1652, 1455, 1157, 1094, 735 cm^{-1} ; 1H -NMR (400 MHz, $CDCl_3$) δ 7.80 (2H, d, J = 7.4 Hz), 7.47 (2H, d, J = 7.2 Hz), 7.37-6.95 (56H, m), 6.45 (1H, d, J = 6.3 Hz), 5.86 (1H, br s), 5.35 (1H, d, J = 11.6Hz), 5.30 (1H, D, J = 2.8 Hz), 4.95 (1H, d, J = 11.3 Hz), 4.89 (1H, d, J = 3.5 Hz), 4.8644.67 (9H, m), 4.54-4.39 (9H, m), 4.34 (1H, dd, J = 10.4, 2.8 Hz), 4.26-4.06 (9H, m), 3.98-3.45 (23H, m), 3.41 (1H, d, J = 10.0 Hz), 3.29-3.20 (5H, m), 0.73 (3H, d, J = 6.3 Hz); ^{13}C -NMR (100 MHz, $CDCl_3$) δ 144.87, 142.49, 139.49, 139.11, 138.87, 138.63, 138.54, 138.37, 138.00, 137.98, 137.97, 137.18, 131.64, 128.74, 128.52, 128.43, 128.33, 128.28, 128.25, 128.21, 128.02, 127.99, 127.97, 127.80, 127.074, 127.67, 127.63, 127.61, 127.54, 127.53, 127.50, 127.44, 127.33, 127.31, 127.02, 126.86, 103.39, 102.78, 100.75, 100.09, 99.80, 99.75, 81.42, 80.64, 78.98, 78.86, 77.82, 77.40, 77.26, 76.26, 75.16, 75.09, 75.07, 74.95, 74.69, 74.30, 73.58, 73.17, 73.11, 72.71, 72.67, 72.65, 72.55, 72.36, 72.18, 69.65, 69.53, 68.54, 68.18, 68.08, 67.85, 67.79, 67.21, 54.95, 16.60.

To liquid ammonia (ca 8 mL) under N_2 at -78^0C was added
35 metallic sodium (95 mg) and stirred for 2 min. To the blue solution was added a solution of the hexasaccharide glycal above (70 gm, 33.8 umol) in dry THF(2mL). After

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45 min at 78°C, the reaction was quenched with absolute methanol (4 mL). Most of ammonia was removed with stream of nitrogen (final volume was ca 4 mL) and the reaction diluted with methanol to ca 10 mL. To the solution was
5 added Dowex 50-X8 (890 mg, washed and dried) and stirred for 5 min. The solution was filterate and rinsed with methanol, finally with ammoniacal methanol (5 mL), and the filterate was concentrated in vacuo. The residue and DMAP (2.4 mg) were placed under N₂ and suspended in DMF
10 (1.0 mL), THF (1.0 mL) and TEA (1.0 mL), then treated with Ac₂O (0.3 mL). After 20h (TLC analysis with EtOAc), the reaction was poured into water (40 mL), and extracted with EtOAc (40 mL x 2), washed with dil NaHCO₃ (30 mL), with water (30 mL), dried (Na₂SO₄) and concentrated.
15 Flash chromatography with 80% EtOAc in CH₂Cl₂ gave 52.0 mg (93%) of product as white foam.

mp 132-134°C; $[\alpha]_D^{23} = +4.7^\circ$ (CHCl₃, c = 1.4); IR (CHCl₃ film) 1742, 1652, 1371, 1227, 1069 cm⁻¹; ¹H-NMR (400 MHz, CDCl₃) δ 6.68 (1H, d, J = 6.8 Hz), 6.42 (1H, d, J = 6.0 Hz), 5.58 (1H, d, J = 3.2 Hz), 5.47 (1H, d, J = 3.4 Hz), 5.40-5.37 (2H, m), 5.29 (1H, dd, J = 10.9, 3.1 Hz), 5.25-5.15 (5H, m) 5.06 (1H, dd, J = 11.2, 3.3 Hz), 5.02 (1H, d, J = 3.6 Hz), 4.99-4.92 (2H, m), 4.84-4.81 (2H, m),
20 4.67 (1H, d, J = 7.8 Hz), 4.56-4.51 (2H, m), 4.45-4.38 (3H, m), 4.29 (1H, dd, J = 10.6, 3.4 Hz), 4.22-3.95 (13H, m), 3.90-3.77 (3H, m), 2.19-1.92 (51H, m), 1.15 (3H, d, J = 6.4 Hz).

30 Peracetyl hexasaccharide glycal above (52 mg) was divided into two portions (22 mg and 30 mg). A solution of hexasaccharide glycal (22.0 mg, 13.4 μ mol) in dry CH₂Cl₂ (2 mL) under N₂ at 0°C, then treated with allyl alcohol (5 mL). The mixture was stirred for 15h at room temperature.
35 Excess allyl alcohol was removed in vacuo. The other batch (30 mg) was treated similarly. The crude products were combined and chromatographed with 85%

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EtOAc-CH₂Cl₂ to give 35.8 mg (66%) of less polar product and 15.7 mg (29%) of more polar product. A 33.2 mg (19 μ mol) of the less polar material under N₂ was dissolved in absolute MeOH (14 mL) and treated with MeONa solution in methanol (165 μ L, 25% by weight). After 6h, the reaction was neutralized with Dowex 50-X8 (200 mg, washed and dried), filtered and concentrated to give quantitative yield of the title compound 19b.

mp 204-206⁰(dec); [α]_D²⁵ = +5.5⁰ (MeOH, c = 0.67); IR (MeOH film) 3356 (br), 2923, 1658, 1374, 1071 cm⁻¹; ¹H-NMR (400 MHz, CD₃OD) δ 5.99-5.93 (1H, m), 5.24 (1H, d, J = 3.8 Hz), 5.18-5.14 (1H, m), 4.93 (1H, d, J = 3.9 Hz), 4.56-4.54 (2H, m), 4.42-4.06 (10H, m), 3.99 (1H, s), 3.91-3.47 (26H, m), 3.41-3.37 (1H, m), 3.27 (1H, t, J = 8.8 Hz), 2.01 (3H, s), 1.24 (3H, d, J = 6.5 Hz); ¹³C-NMR (100 MHz, CD₃OD, ref = δ 49.05) δ 174.55, 135.73, 117.57, 105.48, 105.42, 103.94, 103.26, 102.79, 101.08, 81.21, 80.67, 80.05, 79.20, 78.09, 76.79, 76.56, 76.48, 76.44, 76.41, 75.54, 74.86, 74.68, 73.57, 72.63, 72.50, 71.57, 71.16, 70.64, 70.41, 69.68, 68.16, 62.67, 62.64, 62.57, 61.96, 61.63, 53.11, 23.58, 16.78.

For the purposes of the preparative synthesis of structure 1b a ceramide precursor was attached to the ABC trisaccharide (Scheme 5). Epoxidation of 7b, followed by reaction with the ceramide precursor 17b (as its tributylstannyl ether) promoted by Zn(OTf)₂ provided 20b. Acetylation and PMB removal proceeded smoothly to furnish 21b which is poised for coupling with a suitable DEF trisaccharide donor.

When trisaccharide 15b was treated with MeOTf in the presence of acceptor 21b, a 4:1 mixture of hexasaccharide isomers was obtained. The major product 22b was obtained in 50% yield.

The ceramide side-chain was elaborated by reduction of the azide functionality using Lindlar's catalyst under an atmosphere H_2 in the presence of palmitic anhydride to provide 18b directly. Desilylation was followed by dissolving metal deprotection of the sulfonamide and benzyl groups and MeOH quench to remove the carbonate and acetate groups. Peracetylation of the crude mixture afforded a 78% yield of peracetylated bexasaccharide. Saponification of this material using NaOMe provided the natural product 1b in 96% yield. The coupling constants and chemical shifts of the anomeric protons of 1b matched reported data. In addition, the product was characterized by exact mass, and 1H and ^{13}C NMR.

Synthesis of 20b:

The benzylated ceramide precursor (475 mg, 1.14 mmol) was dissolved in 4 mL PhH. Bis(tributyltin) ether (0.29 mL, 0.34 g, 0.57 mmol) was added and the reaction vessel (equipped with a Dean-Stark trap) was heated to reflux. After 3 h the reaction was allowed to cool and was concentrated under a flow of N_2 . In a separate flask, the glycal 7b was dissolved in 1 mL anhydrous CH_2Cl_2 and the resulting solution was cooled to $0^\circ C$ and a solution of 3,3-dimethyldioxirane (2.8 mL, 0.25 mmol, 0.09 M in acetone) was added. After 45 min the solution was concentrated under a flow of N_2 , then under vacuum. The tin ether was dissolved in 1 mL anhydrous THF and added via cannula to a mixture of $Zn(OTf)_2$ (170 mg, 0.468 mmol) in 1 mL THF at $-78^\circ C$ (wash 1 x 0.5 mL THF). The reaction was allowed to warm to room temperature over 12 h and then was quenched with distilled water. The aqueous phase was extracted 3 x with EtOAc. The combined organic phases were dried over anhydrous $MgSO_4$. Flash column chromatography (3:1 hexane/EtOAc, 3 x 16 cm silica gel) afforded 265 mg (66%) of the target compound 20b.

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¹H NMR (CDCl₃) δ 7.43-7.15 (m, 45H), 7.03 (d, J= 8.6 Hz, 2H), 6.79 (d, J= 8.6 Hz, 2H), 5.76 (dt, J= 6.7, 15.4 Hz, 1H), 5.43 (dd, J= 8.5, 15.4 Hz, 1H), 5.07 (d, J= 3.5 Hz, 1H), 5.05 (d, J= 12.0 Hz, 1H), 4.90 (d, J= 12.9 Hz, 2H), 4.83-4.77 (m, 3H), 4.69 (d, J= 12.0 Hz, 1H), 4.61 (d, J= 11.9 Hz, 1H), 4.54-4.45 (m, 3H), 4.42-4.25 (m, 7H), 4.18-4.05 (m, 6H), 4.01-3.91 (m, 4H), 3.83 (dd, J= 4.4, 10.6 Hz, 1H), 3.79 (s, 3H), 3.71-3.65 (m, 4H), 3.57-3.32 (m, 7H), 3.20 (m, 1H), 2.29 (bs, 1H), 2.11 (bq, J= 6.7 Hz, 2H), 1.42-1.29 (m, 22H), 0.91 (t, J= 6.6 Hz, 3H); ¹³C NMR (CDCl₃) δ 158.8, 139.1, 139.0, 138.7, 138.6, 138.34, 138.29, 138.2, 138.1, 130.8, 128.7, 128.55, 128.50, 128.4, 128.33, 128.28, 128.26, 128.12, 128.06, 127.84, 127.76, 127.7, 127.64, 127.60, 127.5, 127.45, 127.36, 125.8, 113.5, 102.7, 100.6, 81.9, 81.5, 79.4, 77.4, 77.0, 76.7, 76.6, 76.4, 75.5, 74.9, 74.7, 74.4, 73.9, 73.3, 73.2, 73.11, 73.06, 72.3, 72.1, 70.0, 69.4, 68.7, 68.1, 67.9, 67.7, 64.2, 55.2, 32.4, 31.9, 29.70, 29.65, 29.5, 29.4, 29.2, 29.0, 22.7, 14.2; IR (thin film) 3447, 3062, 3029, 2923, 2853, 2099, 1612, 1586, 1514, 1496, 1454, 1364 cm⁻¹; [α]_D²⁵ +25.0 (c 0.70).

Synthesis of 21b:

The above trisaccharide (256 mg, 0.147 mmol) was dissolved in 2 mL anhydrous CH₂Cl₂. Triethylamine (0.105 mL, 76 mg, 0.753 mmol), DMAP (2 mg, 0.02 mmol) and acetic anhydride (0.042 mL, 45 mg, 0.445 mmol) were added sequentially. The reaction was stirred for 1 h then quenched with saturated aqueous NaHCO₃. The extracts were dried with anhydrous MgSO₄, filtered and concentrated. Purification by flash column chromatography (4:1 hexane/EtOAc, 2 x 16 cm silica gel) afforded 235 mg (90%) of the desired compound.

¹H NMR (CDCl₃) δ 7.42-7.17 (m, 45H), 7.03 (d, J= 8.6 Hz, 2H), 6.81 (d, J= 8.6 Hz, 2H), 5.75 (dt, J= 6.7, 15.4 Hz,

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1H), 5.43 (dd, J= 8.6, 15.4 Hz, 1H), 5.07 (d, J= 3.4, 1H), 4.99-4.90 (m, 4H), 4.85 (d, J= 11.3 Hz, 2H), 4.77 (d, J= 11.9 Hz, 1H), 4.76 (d, J= 12.4 Hz, 1H), 4.70 (d, J= 12.0 Hz, 1H), 4.62 (d, J= 11.7 Hz, 1H), 4.57-4.52 (m, 3H), 4.49-4.34 (m, 7H), 4.30 (d, J= 11.8 Hz, 1H), 4.25 (d, J= 11.8 Hz, 1H), 4.14-4.06 (m, 7H), 4.01-3.95 (m, 2H), 3.91 (dd, J= 5.6, 8.6 Hz, 1H), 3.85 (dd, J= 4.3, 11.1, Hz, 1H), 3.80 (s, 3H), 3.74 (d, J= 9.8 Hz, 1H), 3.69 (dd, 7.7, 9.9 Hz, 1H), 3.63-3.51 (m, 5H), 3.43-3.34 (m, 3H), 3.22 (dd, J= 4.6, 8.2 Hz, 1H), 2.12 (dd, J= 6.8, 13.6, 2H), 1.87 (s, 3H), 1.43-1.30 (m, 22H), 0.93, (t, J= 6.6 Hz, 3H); ¹³C NMR (CDCl₃) δ 169.3, 158.8, 139.1, 139.0, 138.69, 138.65, 138.6, 138.31, 138.26, 138.2, 138.1, 138.0, 130.8, 128.8, 128.6, 128.41, 128.35, 128.30, 128.28, 128.14, 128.0, 127.9, 127.8, 127.64, 127.60, 127.58, 127.51, 127.47, 127.38, 126.0, 113.5, 102.7, 100.8, 100.6, 81.5, 79.9, 79.5, 79.4, 79.3, 77.4, 77.1, 76.8, 75.5, 75.3, 74.9, 74.5, 74.2, 73.9, 73.2, 73.1, 73.0, 72.4, 72.2, 72.1, 70.2, 69.4, 68.1, 68.0, 67.9, 67.5, 63.8, 55.2, 32.4, 32.0, 29.72, 29.67, 29.5, 29.4, 29.2, 29.1, 22.7, 20.9, 14.2; IR (thin film) 3028, 2923, 2852, 2098, 1751, 1611, 1513, 1496, 1453, 1365, 1232 cm⁻¹; [α]_D²³ +20.3 (c 0.45).

The trisaccharide from above (230 mg, 0.129 mmol) was dissolved in 4 mL CH₂Cl₂. Distilled water (1 mL) was added and the mixture was cooled to 0°C. DDQ (35 mg, 0.15 mmol) was added and the reaction was stirred for 1 h. The reaction was quenched with saturated aqueous NaHCO₃. The aqueous phase was extracted 3 x with CH₂Cl₂. The combined organic phases were washed dried over anhydrous MgSO₄. Flash column chromatography (4:1 hexane/EtOAc, 2 x 16 cm silica) afforded 182 mg (85%) of the target compound 21b.

¹H NMR (CDCl₃) δ 7.38-7.13 (m, 45H), 5.73 (dt, J= 6.7, 15.4 Hz, 1H), 5.41 (dd, J= 8.6, 15.4 Hz, 1H), 5.09 (d, J=

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3.2 Hz, 1H), 4.98 (d, J= 12.5 Hz, 1H), 4.95 (dd, J= 8.0, 9.2 Hz, 1H), 4.87 (d, J= 11.2 Hz, 1H), 4.80 (d, J= 11.3 Hz, 1H), 4.77 (d, J= 10.9 Hz, 1H), 4.70 (d, J= 11.4 Hz, 1H), 4.65-4.50 (m, 6H), 4.45-4.42 (m, 3H), 4.38-4.34 (m, 3H), 4.28 (bs, 2H), 4.15 (d, J= 11.7 Hz, 1H), 4.11 (d, J=11.8 Hz, 1H), 4.08-4.01 (m, 3H), 3.98-3.94 (m, 3H), 3.88 (dd, J= 5.5, 8.5 Hz, 1H), 3.82 (dd, J= 4.3, 7.0 Hz, 1H), 3.77 (dd, J= 3.1, 10.1 Hz, 1H), 3.70 (d, J= 9.8 Hz, 1H), 3.64-3.51 (m, 5H), 3.46 (dd, J= 5.4, 9.4, 1H), 3.39 (m, 1H), 3.34-3.30 (m, 2H), 3.21 (dd, J= 4.7, 8.4 Hz, 1H), 2.09 (m, 2H), 1.90 (s, 3H), 1.84 (d, J= 5.1 Hz, 1H), 1.41-1.27 (m, 22H), 0.90 (t, J= 6.5 Hz, 3H); ¹³C NMR (CDCl₃) δ 169.3, 165.9, 139.3, 138.7, 138.6, 138.5, 138.3, 138.2, 138.1, 138.0, 128.5, 128.4, 128.32, 128.27, 128.25, 128.17, 128.00, 127.94, 127.91, 127.8, 127.75, 127.70, 127.67, 127.61, 127.55, 127.49, 127.45, 127.21, 125.9, 107.8, 102.6, 100.8, 99.4, 81.4, 80.6, 79.3, 77.5, 77.3, 77.0, 76.9, 76.7, 75.5, 75.3, 75.2, 74.3, 73.2, 73.1, 73.0, 72.9, 72.3, 72.1, 70.1, 70.0, 69.1, 68.1, 68.0, 67.8, 67.4, 63.8, 32.4, 31.9, 29.7, 29.6, 29.5, 29.4, 29.2, 29.1, 22.7, 20.9, 14.1; IR (thin film) 3570, 3087, 3062, 3029, 2924, 2853 2099, 1950, 1873, 1752, 1496, 1453, 1366, 1231 cm⁻¹; [α]_D²³ +17.6 (c 1.40).

25 Synthesis of 22b:

Thioglycoside 15b (188 mg, 0.151 mmol) and the acceptor 21b (125 mg, 0.0751 mmol) were azeotropically dried with benzene twice. The mixture was then dissolved in 2.6 mL anhydrous Et₂O and 1.3 mL CH₂Cl₂ and to this solution was added 500 mg of 4 Å mol. sieves. This mixture was stirred for 1 h and then was cooled to 0 °C and MeOTf (0.051 mL, 74 mg, 0.45 mmol) was added. The reaction was stirred at 0°C for 9 h. Triethylamine (1 mL) was then added and reaction was filtered through a plug of silica and washed with Et₂O. The filtrate was washed with saturated aqueous NaHCO₃ and dried over anhydrous MgSO₄. Purification by preparative HPLC (85:15 hexane/EtOAc)

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afforded 108 mg (50%) of the target compound 22b. The b/a ratio of the reaction was 4:1.

¹H NMR (CDCl₃) δ 7.75 (d, J= 7.2 Hz, 2H), 7.46-7.05 (m, 63H), 5.75 (dt, J= 6.8, 15.2 Hz, 1H), 5.43 (dd, J= 8.6, 15.5 Hz, 1H), 5.13 (m, 2H), 5.09 (d, 3.6 Hz, 1H), 5.05 (d, J= 11.6 Hz, 1H), 5.00 (d, J= 11.5 Hz, 1H), 4.94-4.86 (m, 5H), 4.83-4.65 (m, 14H), 4.59 (d, 11.7 Hz, 2H), 4.53-4.43 (m, 6H), 4.39-4.31 (m, 4H), 4.23 (d, J= 11.9 Hz, 1H), 4.18 (d, J= 11.9 Hz, 1H), 4.15-4.08 (m, 2H), 4.05-3.57 (m, 31H), 3.54 (d, J= 9.1 Hz, 1H), 3.49-3.45 (m, 2H), 3.38 (m, 1H), 3.31-3.23 (m, 3H), 2.92-2.89 (m, 2H), 2.75 (bt, 6.0 H, H), 2.12 (bq, J= 6.9 Hz, 2H), 1.85 (s, 3H), 1.20-1.09 (m, 42H), 0.92 (t, J= 6.6 Hz, 3H); ¹³C NMR (CDCl₃) δ 169.1, 165.9, 155.5, 140.9, 139.2, 139.0, 138.8, 138.64, 138.59, 138.47, 138.43, 138.3, 138.2, 138.10, 138.07, 138.0, 132.1, 129.1, 128.69, 128.65, 128.56, 128.43, 128.40, 128.36, 128.35, 128.26, 128.17, 128.12, 128.08, 127.97, 127.77, 127.66, 127.64, 127.60, 127.54, 127.49, 127.45, 127.41, 127.3, 126.0, 103.0, 102.7, 100.8, 99.7, 99.2, 98.0, 81.2, 80.6, 79.5, 79.2, 79.0, 78.3, 77.7, 76.8, 76.5, 75.5, 75.3, 75.1, 75.03, 74.97, 74.91, 74.87, 74.0, 73.2, 73.10, 73.07, 72.98, 72.93, 72.6, 72.3, 72.1, 72.0, 71.32, 71.25, 70.2, 69.4, 69.32, 69.25, 68.1, 67.9, 67.5, 68.3, 62.1, 62.0, 56.1, 32.4, 31.9, 29.71, 29.68, 29.66, 29.48, 29.38, 29.2, 29.1, 22.7, 20.7, 18.13, 18.11, 18.01, 17.98, 16.9, 14.2, 11.9; IR (thin film) 3344, 3030, 2924, 2864, 2101, 1789, 1754, 1496, 1453, 1366, 1232 cm⁻¹.

Synthesis of 18b:

The hexasaccharide 22b (66 mg, 0.023 mmol) was dissolved in 1 mL EtOAc. Lindlar's catalyst (66 mg) was added followed by the addition of palmitic anhydride (23 mg, 0.046 mmol). The system was purged under vacuum and then put under 1 atm of H₂. After 24 h the reaction was filtered through a plug of silica gel, washed with EtOAc,

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and concentrated. Purification by preparative HPLC (4:1 hexane/EtOAc) afforded 64 mg (90%) of the desired product 18b.

5 ¹H NMR (CDCl₃) δ 7.72 (d, J= 7.2 Hz, 2H), 7.42-7.02 (m, 63H), 5.65 (d, J= 9.1 Hz, 1H), 5.62 (dt, J= 6.6, 15.3 Hz, 1H), 5.31 (dd, J= 8.6, 15.3 Hz, 1H), 5.10 (m, 2H), 5.05 (d, J= 3.6 Hz, 1H), 5.02 (d, J=11.5 Hz, 1H), 4.96 (d, J= 11.4 Hz, 1H), 4.90-4.62 (m, 13H), 4.57-4.38 (m, 8H),
10 4.32-4.26 (m, 3H), 4.21-4.07 (m, 9H), 4.01-3.41 (m, 31H), 3.30 (m, 1H), 3.23 (m, 3H), 2.20 (m, 4H), 1.82 (s, 3H), 1.52 (bm, 2H), 1.32-1.19 (m, 53H), 1.15-1.08 (m, 42H), 0.88 (t, J= 6.8 Hz, 6H); IR (thin film) 3531, 3346, 3063, 3030, 2924, 2854, 1790, 1748, 1674, 1496, 1454, 1365,
15 1236 cm⁻¹; [α]_D²³ -17.9 (c 0.65).

Synthesis of 1b:

The hexasaccharide from above (20 mg, 0.0065 mmol) was dissolved in 0.5 mL THF. A solution of tetrabutylammonium fluoride (1.0 M in THF, 0.050 mL, 0.050 mmol)
20 was added and the reaction was stirred for 2 h. The solution was filtered through a plug of silica, washed with EtOAc and concentrated. The residue was dissolved in 1 mL of anhydrous MeOH and NaOMe (10 mg, 0.19 mmol)
25 was added. The reaction was stirred for 3 h, neutralized with 40 mg of Dowex-50 resin, filtered and concentrated. Purification by flash column chromatography (1.5 x 4 cm 10-40 u silica gel, 95:5 CH₂Cl₂/MeOH) afforded 16.5 mg (94%) of the desired compound.

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¹H NMR (CDCl₃) δ 7.78 (d, J= 7.6 Hz, 2H), 7.46 (d, J= 7.4 Hz, 2H), 7.41-6.97 (m, 61H), 6.02 (d, J= 9.1 Hz, 1H), 5.76 (bs, 1H), 5.67 (dt, J= 6.6, 15.3 Hz, 1H), 5.37-5.30 (m, 2H), 5.19 (d, J= 2.6 Hz, 1H), 4.96 (d, J= 11.3 Hz, 1H),
35 4.93 (d, J= 3.4 Hz, 1H), 4.90-4.83 (m, 3H), 4.78-4.66 (m, 7H), 4.56 (d, J= 11.1 Hz, 1H), 4.53 (d, J= 10.2 Hz, 1H), 4.47-4.32 (m, 5H), 4.28-4.06 (m, 14H), 4.01-3.13

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(m, 36H), 2.73 (bt, 1H), 2.61 (bs, 1H), 2.54 (bs, 1H),
(2.05 (m, 4H), 1.50 (m, 2H), 1.38-1.23 (m, 46H), 0.88 (t,
J= 6.6 Hz, 6H), 0.78 (d, 6.3 Hz, 3H); ^{13}C NMR (CDCl₃) δ
173.4, 142.4, 139.5, 139.0, 138.7, 138.5, 138.33, 138.26,
5 138.14, 138.09, 137.9, 137.2, 137.1, 131.6, 129.0, 128.8,
128.54, 128.47, 128.37, 128.32, 128.27, 128.22, 128.17,
128.14, 128.05, 127.99, 127.79, 127.73, 127.68, 127.63,
127.59, 127.49, 127.46, 127.37, 127.32, 126.98, 126.58,
104.1, 102.83, 102.76, 100.3, 100.2, 82.1, 81.5, 81.2,
10 79.6, 79.2, 79.0, 78.0, 77.3, 77.0, 76.7, 75.6, 75.3,
75.1, 75.0, 74.8, 74.6, 73.5, 73.4, 73.2, 73.0, 72.7,
72.6, 71.9, 70.1, 69.6, 68.5, 68.2, 68.0, 67.5, 62.4,
61.9, 54.8, 52.3, 36.9, 32.3, 31.9, 29.71, 29.67, 29.54,
29.50, 29.43, 29.37, 29.28, 29.20, 25.7, 22.7, 16.7,
15 14.1; IR (thin film) 3424, 3062, 3023, 2923, 2852, 1641,
1530, 1496, 1453, 1362, 1325 cm^{-1} ; $[\alpha]_D^{25}$ -3.2 (c 0.83).

A flask was equipped with a dry ice condenser and was
charged with 4 mL NH₃. Sodium (18 mg, 0.78 mmol) was added
20 and to the resulting blue solution was added 29 mg of the
above hexasaccharide (0.010 mmol). The reaction was
stirred at -78°C for 45 min. Quench by the addition of
MeOH (3 mL). Nitrogen was blown over the solution to
evaporate the NH₃. The reaction was neutralized with 170
25 mg of Dowex-50 resin, filtered and concentrated. The
resulting residue was dissolved in 1 mL of 4:1 THF/DMF.
Triethylamine (0.5 mL) was added followed by the addition
of DMAP (3 mg) and acetic anhydride (0.200 mL). After 2
h the reaction was concentrated in vacuo. Purification
30 by flash column (1.5 x 5 cm 10-40 μ silica, 9:1
EtOAc/hexane) afforded 18 mg (78%) of the peracetate. A
sample of this hexasaccharide (15 mg, 0.0065 mmol) was
dissolved in 0.5 mL of anhydrous MeOH and a NaOMe
solution (30% in MeOH, 0.010 mL, 0.05 mmol) was added.
35 The solution was stirred for 3 h, neutralized with 9 mg
Dowex-50 resin, filtered and concentrated. The residue
was purified by flash column chromatography (1.5 x 4 cm

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C-18 reverse phase silica, MeOH) to afford 9.6 mg of the natural product 1. Spectral data agree with those reported by Hakomori, et al.

5 Biological Results

The MBR1 hexasaccharide has been prepared in two forms, the natural "B" form and the unnatural "A" form as shown below.

10

The natural structure ("β") is:

Fucal→GalB1→3GalNAcB1→3Galα1→4G1B1→4GcB1→1Cer

The unnatural structure "α" is:

15 Fucα1→2GalB1→3GalNAcα1→3Galα1→4GalB1→1Cer

Both have been linked to ceramide to facilitate testing for immunological reactivity with monoclonal antibody (mAb) MBR1.

20

By Thin Layer Chromatography (TLC) the 2 preparations migrate as similar single bands. Immune TLC (see Ritter, G., et al., Cancer Res. 50, 1403-10 (1990)) demonstrates that both forms react with the MBR1 monoclonal antibody specifically but that the β-form reacts 10 times more strongly (comparable staining is seen with 1/10 the amount of antigen). The high level of reactivity of the β structure with mAb MBR1 was confirmed using flow cytometry inhibition assays. Reactivity of MAb MBR1 with breast cancer cell lines such as MCF-7 was 98% inhibited by 8 μg/ml of the β linkage preparation but was only 6% inhibited by 8 μg of the α-linkage preparation. GD3 ganglioside (negative control) showed no inhibition at all.

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**Application of the Glycal Assembly Method to the Concise
Synthesis of Neoglycoconjugates of the Le^y Carbohydrate
Epitope**

5 Demonstrated herein is the use of the glycal assembly
method to obtain glycals of the Le^y specificity,
conjugation to a carrier protein and to a ceramide, and
results of immunizations with these conjugates. The
conjugation strategy used relies on the protocol of
10 Bernstein and Hall (Bernstein, M.A., and Hall, L.D.,
Carbohydr. Res., 1980, 78, C1) which calls for reductive
coupling of a glycolaldehyde glycoside with the intended
carrier, presumably at the ϵ -amino residues of exposed
lysines. The interfacing of the glycal assembly logic
15 with this conjugation strategy led to the paradigm shown
in Figure 1.

**Synthesis of a Le^y epitope and conjugation to protein
carriers**

20
Synthesis of a Le^y pentasaccharide. (For previous
syntheses of Lewis Y see: Jacquinet, J.C., and Sinay, P.,
J. Org. Chem., 1977, 42, 720; Nilsson, S., et al.,
Glycoconj. J., 1989, 6, 21; Schmidt, R.R., and Topfer,
25 A., Tetrahedron Lett., 1991, 32, 3353; Kinzy, W., and
Low, A., Carbohydr. Res., 1993, 245, 193; Hindsgaul, O.,
et al., Carbohydr. Res., 1982, 109, 109; Windmuller, R.,
and Schmidt, R.R., Tetrahedron Lett., 1994, 35, 7927.)
A pentasaccharide containing the Le^y specificity was
30 prepared as shown in Figures 22(a) and (b). In the
synthesis of this determinant, the N-acetyllactosamine
backbone of the target was exploited. Lactal 7c
(Haworth, W.N., et al., J. Chem. Soc., 1930, 2644) was an
attractive starting material if a concise way of
35 identifying the C₃ and C₂-hydroxyls could be effected.

Readily available lactal was silylated at the two primary sites. Following these silylations, the 3' and 4' hydroxyls were engaged as cyclic carbonate, 8c. Accordingly, the pertinent hydroxyl groups were exposed for further transformation. Difucosylation of 8c utilizing fluorosugar 9c (Danishefsky, S.J., et al., J. Am. Chem. Soc., 1992, 114, 8329) as the donor (Mukaiyama, T., et al., Chem. Lett., 1981, 431) provided access to the Le^y series as glycal 10c. The use of a fucosylating agent with a protecting group (4-benzoate) that had the potential to donate into an oxonium intermediate was chosen to ensure α -selectivity. The glycal double bond was activated for azaglycosylation by a previously developed iodosulfonamidation protocol to afford 11c. (Danishefsky, S.J., et al., J. Am. Chem. Soc., 1992, 114, 8331; Griffith, D.A., and Danishefsky, S.J., J. Am. Chem. Soc., 1990, 112, 5811.)

Use of the iodosulfonamide to glycosylate the tin ether of galactal 12c (Danishefsky, S.J., et al., J. Am. Chem. Soc., 1992, 114, 8331) in the presence of silver tetrafluoroborate led to glycal 13c as shown in Figure 22(b). Deprotection followed by peracetylation afforded peracetyl glycal 14c. Reaction of 14c with dimethyldioxirane (Halcomb, R.L., and Danishefsky, S.J., J. Am. Chem. Soc., 1989, 111, 6661) followed by opening of the epoxide with allyl alcohol, followed by deacetylation with catalytic methoxide led to pentasaccharide 15c.

To effect conjugation of the Le^y determinant to a protein carrier, 15c was ozonolyzed in MeOH at -78°C. Work-up with dimethylsulfide afforded aldehyde 16c which was reductively attached to its BSA-protein carrier. The reductive amination protocol described by Bernstein and Hall was adapted to the present case. Thus, treatment of 16c with BSA in pH 8 phosphate buffer and excess sodium cyanoborohydride led to conjugate 17c which was purified

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by exhaustive dialysis. TFA analysis showed the expected sugar composition: 2 parts galactose, 2 parts fucose and 1 part N-acetylglucosamine. (For sugar analysis protocols see: Lloyd, K.O., and Savage, A., Glycoconj. J., 1991, 8, 493; Hardy, M.R., and Townsend, R.R., Proc. Natl. Acad. Sci., 1988, 85, 3289; for protein analysis see: Bradford, M.M., Anal. Biochem., 1976, 72, 248.) Carbohydrate:protein analyses showed the uptake of an average of 15 Le^y moieties per carrier molecule. This conjugate was recognized by an antibody to the Le^y blood group.

For an extended Lewis^y epitope the rollover of iodosulfonamide 11c was attempted with the regiospecifically generated tin ether lactal 18c. Under the usual reaction conditions the best yield of the desired hexasaccharide 19c was 15% as indicated in Figure 23(a).

Similar results were obtained with a variety of other Le^y donors generated from iodosulfonamide 11c (Figure 23(b)). This led to the hypothesis that the Le^y tetrasaccharide donor may be poor due to the steric demands around the newly forming glycosidic linkage. Only the relatively flattened structure of galactal appears to add to the donor adequately.

Synthesis of a ceramide-linked glycoconjugate

In the synthesis of the closely related Le^b carbohydrate antigen (Danishefsky, S.J., et al., J. Am. Chem. Soc., 1995, 117, 5701), it was found that simplification of the protecting group scheme was possible by use of the perbenzylated fucosyl fluoride 20c. No erosion of a selectivity in the difucosylation of 21c was evident resulting in a 70% yield of Le^b glycal 22c. When

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identical conditions were used to generate the Le^y glycal quite the opposite was true as shown in Figure 18(a).

Two major difucosylated products were obtained in a ratio of about 1.8 to 1. Identification of these products required more than direct inspection of the 1-D ¹H NMR spectra. The tetrasaccharide with the two alpha-linked fucose residues 23c showed one obvious alpha linkage (5.30 ppm, d, J = 3.2 Hz). The other alpha link was anomalously upfield (4.62 ppm, d, J = 3.7 Hz). The coupling constant was measured by a DQF-COSY (phase-sensitive) experiment. The minor product 24c contained a b-linked fucose at the glucal 3 position. A simple COSY experiment established the existence of an a-linked fucose (5.09 ppm, d, J = 3.6 Hz) and a b-linked fucose (4.35 ppm, d, J = 7.5 Hz). From the proton assignments in the COSY experiment and with the aid of HMQC and HMBC experiments the b-linked fucose moiety was shown to reside at the glucal 3 position (very strong coupling between C-3 of glucal and H-1 of b-fucose in the HMBC experiment). The chemical yield (80%) for this experiment was exceptionally good despite the lack of stereoselectivity. A similar erosion of selectivity was observed during the study of the monofucosylation of a 6-mono protected glucal in the synthesis of the sialyl Le^x carbohydrate antigen. (Gervay, J., et al., J. Org. Chem. 1993, 58, 5465.)

A large body of empirical evidence suggests that α -glycosylations (Löhn, H. Carbohydr. Res. 1985, 139, 105; Kahne, D., et al., J. Am. Chem. Soc. 1989, 111, 6881; Rainer, H., et al., Liebigs Ann. Chem. 1992, 103; Wessel, H.P., Tetrahedron Lett. 1990, 31, 6863) are generally favored in nonpolar solvents. (In fucosyl fluoride 20c, the more hindered hydroxyls tend to favor the alpha linkage.) This suggests it is possible to enhance the

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selectivity of the reaction by decreasing solvent polarity.

The logical choice was to switch from THF to toluene. Conducting the reaction in toluene as solvent led to no reaction. This was attributed, however, to the insolubility of the tin triflate activator in toluene. The first success in modifying the reaction conditions was the use of a mixed solvent system of 10:1 toluene:THF. The selectivity for the doubly alpha-fucosylated product increased about two-fold with a comparable chemical yield of 83%. The best ratio of 23c to 24c (>40:1 as determined by HPLC) was obtained with the use of a mixed solvent system of 10:1 toluene: dichloromethane (DCM) with a modest yield of 50%. Reaction under these conditions was very slow, and presumably suffers from the lack of solubility of the activating tin triflate reagent. Because the isomers were easily separated it was practical to run the glycosylation for scale up with the 10:1 toluene:THF solvent system.

In a survey to find more active Le^y donors in simple model systems, the fluorosugar 25c was found to be a reasonably good donor when activated with zirconocene dichloride (Matsumoto, T., et al., Tetrahedron Lett., 1988, 29, 3567) and silver triflate. 25c was formed by reaction of iodosulfonamide 26c with water/THF/triethylamine in the presence of silver carbonate followed by reaction of the resulting free reducing sugar with DAST as shown below in Figure 18(b).

With the aim of synthesizing a Le^y ceramide conjugate reaction of 25c with azidosphingosine 27c (Schmidt, R.R., and Zimmermann, P., Tetrahedron Lett., 1986, 27, 481) under the mixed metal system gave acceptable yields of the Le^y sphingosine 28c as indicated in Figure 18(c).

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The reduction of the azide followed by N-acylation with a palmitic acid side chain proved to be quite challenging. In a model study for the reduction of the azide with subsequent acylation, conditions developed by Corey proved sufficient. (Corey, E.J., et al., Synthesis, 1975, 590) Thus, treatment of galactose azidosphingosine 29c in ethyl acetate with Lindlar's catalyst and palmitic anhydride with a balloon of hydrogen led to galactose ceramide 30c in 91% yield after only 6 hours (Figure 18(d)). As applied to 28c, however, the conditions were less effective. The desired ceramide Le^y conjugate 31c was obtained in only 22% yield (Figure 19(a)). Two other by-products were formed during the reaction both of which lacked the double bond in the side chain ceramide.

As monitored by TLC, the reduction of the azide moiety in 28c was considerably slower than in the model system. One possible reason for this is that the azide now resides near the bulky branching sugar sector leading to inaccessability to the catalyst surface. Thus, the rate of reduction of the azide may be comparable to the rate of reduction of the double bond. Other conditions to reduce the azide remain to be explored. The conjugate 31c was deblocked as shown in Figure 19(b); the ceramide linked Le^y conjugate 32c was evaluated in immunological studies.

Results of immunizations with Le^y conjugates 17c and 32c

The first experiment to characterize the synthetic antigen entailed testing the reactivity of the synthetic conjugate 17c with known antibodies. This is shown in Figure 20(a). 17c showed reactivity with aLe^y (s193), an anti-Lewis^y antibody. As a control, 17c was tested against anti-Lewis^b antibody aLe^b (T218). As indicated in the graph, 17c showed no cross reactivity with this antibody.

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The stage was set for immunizations with conjugates 17c and 32c. The immunizations with the BSA conjugate 17c were carried out on two sets of 5 mice and the immunizations with the ceramide conjugate 32c were carried out on one set of 5 mice. An enzyme-linked immunosorbant assay (ELISA) was used to measure antibody titer (Figure 20(b)). Three different ELISA experiments were performed to measure antibody titer. The first is the measure of total antibody response (aIg), that is the summation of IgM and IgG antibodies, as shown in graphs A, D, G, and J of Figures 21(a) and (b). The other two assays measure IgM and IgG antibodies separately as indicated in the graphs. The ELISA assay (see Figure 20(b)) involves the immobilization of a test antigen in a microtiter well. The serum to be tested for antibodies is placed in the well. Any antibodies in the serum recognizing the test antigen will remain in the well after thorough washing. Next, modified anti-mouse Ig, IgM, or IgG antibodies are placed in the well. These secondary anti-immunoglobulin antibodies have been linked to alkaline phosphatase. After thorough rinsing, the wells are treated with p-nitrophenylphosphate (PNP). The presence of the alkaline phosphatase-linked secondary antibody causes cleavage of the phosphate, yielding the yellow p-nitrophenolate anion, which can be quantified using spectrophotometric techniques.

Graphs A, B, and C in Figure 21(a) show the measured titers of total antibody Ig (A), IgM (B), and IgG (C) type antibodies in five mice immunized with conjugate 17c. The dashed line indicates a control mouse that was not immunized. This serves as the background, since natural antibodies are found in the sera. As seen in these graphs, all mice made antibodies against the immunizing antigen. Remarkably, 4 out of 5 mice produced IgG type antibodies. As discussed previously this is unusual. Graphs D-F represent a control where mice were

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immunized with the BSA carrier alone. This ensures that the response in A-C is due to the conjugate 17c and not just a response to the protein carrier itself, which is a potential immunogen. As indicated in D-F the response seen in A-C is due to 17c and is not an artifact of the immunogenicity of the carrier.

A more important issue was whether the antibodies to the synthetic antigen were capable of recognizing the Le^y mucin structure. Graphs G-I show the measured titer of antibodies with the Le^y mucin as the test antigen. One mouse showed outstanding response to the immunizing antigen by producing both IgG as well as IgM antibodies. All the mice showed an IgM response (graph H) with the mucin structure.

Results of the immunizations with the ceramide conjugate 32c (graphs J-L) were not as exciting. No mice showed any IgG response. Most mice showed only a weak IgM response. The results of these immunization experiments are encouraging and warrant closer investigation into the factors that result in the potency and specificity of the immune response to synthetically generated carbohydrate-based vaccinations.

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Experimental

[(6-O-tert-Butyldiphenylsilyl-3,4-carbonate-b-D-galactopyranosyl)-(1→4))-1,5-anhydro-6-O-tert-butyldimethylsilyl-2-deoxy-D-arabino-hex-1-enopyranose (8c)

To 2.11g (6.84 mmol) of lactal 7c was added 2.80g (41 mmol) of imidazole and 25 mL of dry DMF. The solution was cooled to -10°C and 3.56 mL (13.7 mmol) of tert-butyldiphenylsilyl chloride was added dropwise over 15 minutes. The reaction was allowed to warm gradually to

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room temperature. After stirring 8h the reaction was diluted with 200mL of EtOAc and washed 3 times with 100 mL water and once with saturated brine. The organics were dried over MgSO_4 , filtered, and concentrated in vacuo. The crude material was run through a plug of silica gel with 50% EtOAc/Hexanes and concentrated to give 4.50g (84%) of white foam. This was taken up in 125mL of dry THF and a few crystals of imidazole were added. The solution was cooled to 0°C and 938mg (5.79 mmol) of carbonyl diimidazole was added. After 2 hours most of the starting material was consumed as judged by thin layer chromatography. The reaction was concentrated and chromatographed on silica gel (40% EtOAc/Hexanes) to afford 910 mg (20%) of recovered starting material and 2.72g (58%) of 8c as a white foam: $[\alpha]_D^{25} = -8.7^\circ$ (c 1.35, CHCl_3); IR (thin film) 3450, 2900, 2830, 1785, 1632, 1415, 1225, 1145, 1100, 810, 730, 695; ^1H NMR (400MHz, CDCl_3): δ 7.76-7.69 (m, 8 H), 7.51-7.43 (m, 12 H), 6.38 (d, 1H, $J = 5.9$ Hz), 4.83 (d, 1H, $J = 7.2$ Hz), 7.75 (dd, 1H, $J = 2.0, 6.1$), 4.72 (d, 1H, $J = 6.91$ Hz), 4.64 (app. t, 1H, $J = 5.5$ Hz), 4.40 (d, 1H), 4.08-3.86 (m, 7H), 3.69-3.68 (br, 2H), 2.80 (d, 1H, $J = 3.6$ Hz), 1.13 (s, 9H), 1.12 (s, 9H); HRMS (FAB) calcd for $\text{C}_{45}\text{H}_{54}\text{O}_{10}\text{Si}_2\text{Na}$ 833.3153, found m/z 833.3157 (M+Na).

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[[[(4-O-Benzoyl-2,3-dibenzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate- β -D-galactopyranosyl)-(1-4)]-(4-O-benzoyl-2,3-dibenzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]-1,5-anhydro-6-O-tert-butyldiphenylsilyl-2-deoxy-D-arabino-hex-1-enopyranose (10c)

To 2.00g (2.47 mmol) of lactal carbonate 8c was added 4.44g (9.86 mmol) of fucosyl fluoride 9c. The mixture was azeotroped 5 times with benzene and placed under high vacuum for two hours. Under an argon atmosphere 2.77mL (12.33mmol) of di-tert-butyl pyridine and 16mL of dry

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ether were added. 2.0 g of freshly activated 4A molecular sieves were added and the mixture stirred one hour at room temperature. In an argon glove bag, 2.34g (12.33 mmol) of stannous chloride (SnCl_2) and 2.56g (12.33 mmol) of silver perchlorate (AgClO_4) were added. The flask was equipped with a reflux condensor and the reaction brought to reflux for 72 hours. The reaction was quenched with 5mL of saturated bicarbonate and filtered through a pad of celite. Diluted with 50mL ethyl acetate and washed 2 times with saturated bicarbonate, 2 times with sat. copper sulfate and 2 times with sat. brine. The organics were dried over MgSO_4 and concentrated. Flash chromatography in 20% ethyl acetate/hexanes afforded 2.10g (51%) of a white foam 10c:

[a]_D -78.9° (c 0.555, CHCl_3); IR (thin film) 3040, 3000, 2905, 2860, 2830, 1820, 1800, 1710, 1635, 1585, 1570, 1480, 1460, 1440, 1415, 1370, 1350, 1300, 1260, 1205, 1145, 1100, 950, 735, 695; ^1H NMR (400MHz, CDCl_3) d 8.09 (d, J = 8.12 Hz, 2H), 8.00 (d, J = 8.26 Hz, 2H) 7.66 (m, 4H), 7.59 (d, J = 6.74 Hz, 4H), 7.56 (t, J = 7.27 Hz, 1H), 7.30-7.50 (m, 22H) 7.16-7.26 (m, 10H) 7.09 (m, 2H), 6.99 (t, J = 7.59 Hz, 2H) 6.89 (t, J = 7.97 Hz, 1H), 6.43 (d, J = 6.08 Hz, 1H), 5.46 (bs, 1H), 5.38 (bs, 1H), 5.35 (d, J = 3.42 Hz, 1H), 4.89 (d, J = 11.35 Hz, 1H), 4.75-4.80 (m, 4H), 4.72 (d, J = 5.88 Hz, 2H), 4.69 (d, J = 4.27 Hz, 2H), 4.36-4.55 (m, 5H), 4.28 (q, J = 6.51 Hz, 1H), 4.17 (bd, J = 5.46 Hz, 1H), 3.90-4.00 (m, 6H), 3.85 (d, J = 2.99 Hz, 1H), 3.82 (d, J = 2.89 Hz, 1H), 3.56-3.78 (m, 4H), 1.07 (m, 24H); HRMS (FAB) calcd for $\text{C}_{99}\text{H}_{106}\text{O}_{20}\text{Si}_2\text{Na}$ 1694.6740 found m/z 1694.6787 (M+Na).

[[[(4-O-Benzoyl-2,3-dibenzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate- β -D-galactopyranosyl)-(1-4)]-(4-O-benzoyl-2,3-dibenzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]-6-O-tert-butyldiphenylsilyl-2-deoxy-2-iodo- α -D-manno-pyranosyl benzenesulfonamide (11c)

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To 400 mg (0.239mmol) of tetrasaccharide glycal 10c (azeotroped 3x with 15mL benzene) was added 113 mg (0.718mmol) of benzenesulfonamide and 400 mg of freshly activated 4 Å powdered molecular sieves. This was taken
 5 up in 2.7mL of methylene chloride and cooled to 0°C. Stirred for 30 minutes at 0°C and then added 392 mg (0.837mmol) of iodonium di-sym. collidine perchlorate in one portion. Stirred for 30 minutes and then quenched at 0°C with 3mL of saturated sodium thiosulfate. The
 10 reaction mixture was diluted with EtOAc and filtered through a celite pad into a separatory funnel. Washed once with saturated sodium thiosulfate, once with saturated copper sulfate, and once with saturated brine. The organics were dried over MgSO₄, filtered, and
 15 concentrated. Purification by silica gel chromatography (32% EtOAc/Hexanes) afforded a white foam 11c: $[\alpha]_D^{25} = -95.8^\circ$ (c 0.58, CHCl₃); IR (thin film) 2910, 2835, 1800, 1710, 1440, 1425, 1350, 1260, 1150, 1100, 1040, 695; ¹H NMR (400MHz, CDCl₃): δ 8.09 (d, 2H, $J = 7.1$ Hz), 8.02 (d, 2H, $J = 6.99$ Hz), 7.75 (d, 2H, $J = 7.49$ Hz), 7.67-7.09 (m, 49H), 5.93 (br s, 1H), 5.61 (br s, 1H), 5.46 (br s, 1H), 5.43 (br s, 1H), 5.06-4.54 (m, 10H), 4.47 (br d, 1H, $J = 10.3$ Hz), 4.36-4.30 (m, 2H), 4.18 (br, 2H), 3.98-3.70 (m, 11H), 3.40-3.28 (br, 2H), 1.10 (s, 9H), 1.05 (s, 9H),
 20 1.02 (d, 3H, $J = 6.4$ Hz), .98 (d, 3H, $J = 6.4$ Hz); HRMS (FAB) calcd for C₁₀₅H₁₁₂NO₂₂ISi₂Na 1977.5900, found m/z 1977.6037 (M+Na).

[[[(4-O-Benzoyl-2,3-dibenzyl-6-deoxy-α-L-galactopyranosyl)-(1-2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate-β-D-galactopyranosyl)-(1-4)]-[(4-O-benzoyl-2,3-dibenzyl-6-deoxy-α-L-galactopyranosyl)-(1-3)]-(2-benzenesulfonamido-6-O-tert-butyldiphenylsilyl-2-deoxy-β-D-glucopyranosyl)-(1-3)]]-1,5-anhydro-2-deoxy-6-O-tri-
 30 iso-propylsilyl-D-lyxo-hex-1-enopyranose (13c)

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230 mg (0.12mmol) of iodosulfonamide 11c was azeotroped 5 times with dry benzene and placed under high vacuum for two hours. To this was added 2.4mL of THF solution of 15eq. of tin ether 12c (generated by azeotropic removal of water overnight with a Dean-Stark trap equipped with freshly activated 4 Å mol. sieves from 561 mg (1.80mmol) of 6-TIPS-galactal and 673mL (1.32mmol) bis(tributyltin) oxide in 80mL of benzene). To this solution stirring under an argon atmosphere was added 200 mg of freshly activated 4 Å powdered molecular sieves. Stirred one hour at room temperature. Cooled solution to -78°C and added, via cannula, a solution of 187 mg (0.96mmol) of silver tetrafluoroborate in 2.4mL of THF. Warmed to room temperature over 15 hours and quenched the reaction, which had turned bright yellow, with 2mL. of sat. bicarbonate. The reaction mixture was filtered through a pad of celite into a separatory funnel. The celite pad was washed thoroughly with ethyl acetate. The organics were washed twice with sat. bicarbonate and twice with sat. brine. The organics were dried over MgSO₄. Concentration and chromatography in 25% ethyl acetate/hexanes gave 193 mg (75%) as a white foam 13c: [α]_D -126.4° (c 0.505, CHCl₃); IR (thin film) 3500, 3040, 3000, 2905, 2840, 1820, 1800, 1705, 1635, 1590, 1440, 1410, 1255, 1195, 1100, 1080, 1035, 815, 730, 695; ¹H NMR (400 MHz, CDCl₃) δ 8.09 (app t, 4H), 7.65-7.08 (m, 46H), 6.90 (t, J = 7.65 Hz, 3H), 6.76 (d, J = 6.91 Hz, 2H), 6.12 (d, J = 6.59 Hz, 1H), 5.50 (bs, 1H), 5.45 (bs, 1H), 5.28 (app t, 2H), 4.91-3.03 (m, 36H), 2.82 (br s, 1H) 1.09 (m, 45H); HRMS (FAB) calcd for C₁₂₀H₁₄₁NO₂₆SSi₃Na 2150.8668 found m/z 2150.8765 (M+Na).

[[[(2,3,4-Tri-O-Acetyl-6-deoxy-α-L-galactopyranosyl)-(1-2)]-(3,4,6-tri-O-acetyl-β-D-galactopyranosyl)-(1-4)]-[(2,3,4-tri-O-acetyl-6-deoxy-α-L-galactopyranosyl)-(1-3)]-(2-acetamido-6-O-acetyl-2-deoxy-β-D-glucopyrano-

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syl)-(1-3)]-1,5-anhydro-4,6-di-O-acetyl-2-deoxy-D-lyxo-hex-1-enopyranose (14c)

480 mg (0.225 mmol) of pentasaccharide glycal 13c was
5 taken up in 200 mL of THF and 52 mL (0.902 mmol) of
glacial acetic acid and 1.35 mL of TBAF (1M in THF, 1.35
mmol) were subsequently added at room temperature. After
stirring 8 hours 13c was completely consumed and a new
product appeared as judged by TLC. The reaction mixture
10 was concentrated and chromatographed in 8% MeOH/CHCl₃.
The resulting white solid was taken up in THF and added
to solution of 15mL of about 5:1 NH₃/THF to which a large
excess of solid sodium metal had been added. The dark
blue solution was allowed to reflux at -33°C for 20
15 minutes. The reaction was quenched with 5 mL of MeOH and
stirred overnight. The reaction was partially concen-
trated and then cooled to 0°C. The reaction mixture was
then carefully acidified to pH 8 with Dowex 50-X200 ion
exchange resin. The reaction was filtered and
20 concentrated. The crude solids were taken up in 3mL of
pyridine and 3mL of acetic anhydride and allowed to stir
overnight. Purification by silica gel chromatography
(30% acetone/benzene) gave 110 mg (37%) of a white solid
14c: [α]_D -121.3° (c .635, CHCl₃); IR (thin film) 2930,
25 1735, 1362, 1224, 1060, 1040; ¹H NMR (400 MHz, CDCl₃) δ
6.33 (d, 1H, J = 6.3 Hz), 5.70 (d, 1H, J = 7.2 Hz), 5.30-
5.26 (m, 6H), 5.13 (dd, 1H, J = 3.1, 11.0 Hz), 5.05 (dd,
1H, J = 3.2, 11.1 Hz), 5.00-4.92 (m, 4H), 4.88 (d, 1H, J
= 6.6 Hz), 4.79 (dd, 1H, J = 3.4, 6.1 Hz), 4.53-4.28 (m,
30 7H), 4.22-4.08 (m, 4H), 3.83-3.69 (m, 3H), 3.51 (br dd,
1H, J = 3.7, 9.5 Hz), 2.17 (s, 3H), 2.14 (br s, 6H), 2.13
(br s, 6H), 2.11 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H) 2.00
(s, 3H), 1.98 (s, 3H), 1.96 (s, 3H), 1.95 (s, 3H), 1.91
(s, 3H), 1.17 (d, 3H, J = 6.4 Hz), 1.14 (d, 3H, J = 6.5
35 Hz); HRMS (FAB) calcd for C₅₆H₇₇NO₃₄Na 1330.4220 found m/z
1330.4168 (M+Na).

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O-Allyl-[[[(6-deoxy-a-L-galactopyranosyl)-(1-2)]-(b-D-galactopyranosyl)-(1-4)]-[(6-deoxy-a-L-galactopyranosyl)-(1-3)]-(2-acetamido-2-deoxy-b-D-glucopyranosyl)-(1-3)]]-b-D-galactopyranose (15c)

- 5 To 110 mg (84 mmol) of peracetate glycal 14c was added 100 mL of dry methylene chloride. The solution was cooled to 0°C and 1.44 mL of 3,3-dimethyldioxirane solution (0.07 M in acetone, 100 mmol) was added.
- 10 Stirring continued for 20 minutes at 0°C and then the reaction was concentrated in vacuo. The white solid was taken up in 1 mL of allyl alcohol and then cooled to -78°C. 100 mL of ZnCl₂ solution was added and the reaction was allowed to warm to room temperature overnight. The
- 15 reaction was diluted with EtOAc and washed twice with saturated sodium bicarbonate and once with saturated brine. The organics were dried over MgSO₄, filtered, and concentrated. Purification by silica gel chromatography (35% acetone/benzene) gave a white solid which was
- 20 immediately deacetylated. Added 1 mL of MeOH and then a few drops of 2.5% NaOMe in MeOH (Aldrich 25%, 1mL diluted to 10mL with MeOH) and stirred overnight. The reaction was cooled to 0°C and acidified to pH 7 with Dowex 50-X200 ion exchange resin. The reaction was filtered and
- 25 concentrated. Purification with RP-18 reverse phase silica gel (10% MeOH/H₂O) afforded 55 mg (72%) of a white solid 15c: [a]_D -72.7° (c .1 MeOH); IR (thin film) 3350, 2940, 2900, 2830, 1650, 1550, 1365, 1300, 1155, 1070. 1030; ¹H NMR (400 MHz, CD₃OD) δ 5.95 (m, 1H), 5.32 (d, J
- 30 = 17.3 Hz, 1H), 5.19-5.14 (m, 2H), 5.04 (d, J = 3.8 Hz, 1H), 4.68 (d, J = 8.3 Hz, 2H), 4.51 (d, J = 5.7 Hz, 1H), 4.36 (dd, 1H, J = 5.2, Hz), 4.25 (d, 1H, 7.7 Hz), 4.19-4.10 (m, 2H), 4.04 (d, 1H, J = 2.2 Hz), 3.96-3.33 (m, 34H), 1.96 (s, 3H), 1.23 (m, 6H); HRMS (FAB) calcd for
- 35 C₃₅H₅₆NO₂₄Na 900.3325 found m/z 900.3310 (M + Na).

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Synthesis of Le^y-BSA Neoglycoconjugate 17c: 3.2 mg (3.6 mmol) of allyl glycoside 15c was taken up in 2 mL of MeOH and cooled to -78 °C. Ozone was bubbled through the solution until it appeared to be faintly blue (< 2 min.).

5 The reaction was stirred for an additional 2-3 minutes and then the excess ozone was purged with a vigorous flow of argon until the blue color dissipated. About 2 mL of dimethyl sulfide was added. The reaction was gradually warmed to room temperature over about 4 hours and then

10 was stirred an additional 4 hours. The reaction was concentrated in vacuo and placed under high vacuum for 1 hour. To the crude aldehyde was added 1mg (0.015 mmol) of bovine serum albumin (BSA, Sigma Diagnostics Protein Standard) and 200 mL of pH 8 sodium phosphate buffer. 1

15 mg (14.4 mmol) of sodium cyanoborohydride was then added. The solution was stirred slowly for 3 days after which time it was placed in dialysis tubing (Spectra Por, MWCO 12,400). Exhaustive dialysis against distilled water, followed by lyophilization gave 1.2 mg of a fluffy white

20 cotton 17c. TFA analysis of 17c indicated the Le^y pentasaccharide:protein ratio to be about 15:1. Carbohydrate composition was also determined with a composition of 2 parts L-fucose, 2 parts D-galactose, and 1 part D-glucosamine. Conjugate 17c was recognized by

25 Anti-Le^y (S193).

[[[(4-O-Benzoyl-2,3-di-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1 \rightarrow 2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate-b-D-galactopyranosyl)-(1 \rightarrow 4)]-[(4-O-benzoyl-2,3-di-

30 O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1 \rightarrow 3)]-(2-benzene-sulfonamido-6-O-tert-butyldiphenylsilyl-2-deoxy-b-D-glucopyranosyl)-(1 \rightarrow 3)]-(6-O-tert-butyldiphenylsilyl-b-D-galactopyranosyl)-(1 \rightarrow 4)]-1,5-anhydro-6-O-tert-butyldi-

35 phenylsilyl-2-deoxy-D-arabino-hex-1-enopyranose (19c)

To 110 mg (56 mmol) iodosulfonamide 11c added the tin ether of lactal 18c (formed by azeotroping 704 mg (897

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mmol) of disilyllactal with 233 mL (458 mmol) of bis-tributyltin oxide in 30 mL of benzene) in 1.2 mL of THF. 300 mg of freshly activated 4 Å powdered molecular sieves was added. The reaction mixture was cooled to -78°C and

5 88 mg (450 mmol) of silver tetrafluoroborate in 1.2 mL of THF was added to the reaction. The reaction was allowed to warm to room temperature overnight. Stirring continued at room temperature for another 24 hours, during which time the reaction turned a bright yellow-

10 orange color. The starting iodosulfonamide appeared to be consumed by TLC. The reaction was quenched with saturated sodium bicarbonate and filtered through celite. The filtrate was diluted with EtOAc and washed twice with saturated sodium bicarbonate and twice with saturated

15 brine. The organics were dried over MgSO₄, filtered and concentrated in vacuo. Purification by silica gel chromatography twice (35% EtOAc/hexanes and then 8% EtOAc/benzene) afforded 17 mg (12%) of hexasaccharide **19c** as a white foam: $[\alpha]_D -44.8^\circ$ (c .105, CHCl₃); IR (thin film) 3450, 2910, 2840, 1800, 1710, 1260, 1150, 1100, 1050, 730, 695; ¹H NMR (400 MHz, CDCl₃) δ 8.14 (d, 2H, J = 7.08 Hz), 8.07 (d, 2H, J = 6.92 Hz), 7.82 (m, 4H), 7.74 (m, 4H), 7.67 (m, 8H), 7.56-7.37 (m, 30H), 7.32-7.13 (m, 16H), 6.97 (t, 2H, J = 7.49 Hz), 6.85 (t, 1H), 6.41 (d, 1H, J = 4.19 Hz), 5.52 (d, 1H, J = 2.93 Hz), 5.47 (d, 1H, J = 2.36 Hz), 5.24 (d, 1H, J = 3.58 Hz), 4.90 (d, 1H, J = 11.16 Hz), 4.81-4.74 (m, 5H), 4.67-4.48 (m, 7H), 4.37 (m, 2H), 4.27 (m, 2H), 4.10-3.95 (m, 9H), 3.87-3.70 (m, 10H), 3.58-3.40 (m, 5H), 3.11 (broad d, 2H), 2.60 (broad s, 1H), 1.12-1.03 (m, 39 H), 0.87 (d, 3H, 6.41 Hz); LRMS (FAB) calcd for C₁₄₉H₁₆₇NO₃₁SSi₄Na 2634 found m/z 2634 (M+Na).

[[[(2,3,4-Tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate- β -D-galactopyranosyl)-(1-4)]-(2,3,4-tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]-1,5-anhydro-6-O-tert-butyldi-

35

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phenylsilyl-2-deoxy-D-arabino-hex-1-enopyranose (Le^y-Ceramide) (23c)

To 1.43 g (1.76 mmol) of lactal 8c was added 3.07 g (7.04 mmol) of fluorosugar 20c. The two components were azeotroped three times with benzene and then placed under high vacuum overnight. The mixture was taken up in 36 mL of toluene and 3.2 mL (14.08 mmol) of di-tert-butylpyridine was added. 1.00 g of freshly activate 4Å molecular sieves powder was added and the mixture stirred for 15 minutes. The solution was then cooled to 0°C. In a separate dry flask was added 2.94 g (7.04 mmol) of tin (II) triflate and 3.6 mL of dry THF. The solution of tin triflate was then added via cannula to the cooled reaction mixture. The reaction appeared complete by TLC after 4 hours. The reaction was quenched with saturated sodium bicarbonate solution and then filtered through a pad of celite. The filtrate was washed twice with saturated sodium bicarbonate and twice with saturated brine. The organics were dried over MgSO₄, filtered and concentrated in vacuo. Purification by silica gel chromatography (15% EtOAc/hexanes) gave a ~4.9:1 mixture of 23c and 24c which could be separated by HPLC (Waters RCM, 18% EtOAc/hexanes) to give 1.95 g (67%) of 23c as a white foam: $[\alpha]_D^{25} = -55.5^\circ$ (c 0.780, CHCl₃); IR (thin film): 3050, 3020, 2920, 2850, 1800, 1425, 1350, 1235, 1155, 1105, 1050, 820, 740, 700; ¹H NMR (400 MHz, CDCl₃): d 7.67 (m, 2H, ArH), 7.62 (m, 4H, ArH), 7.56 (m, 2H, ArH), 7.49 (m, 2H, ArH), 7.46-7.25 (m, 32H, ArH), 7.14 (m, 4H, ArH), 7.01 (m, 4H), 6.35 (d, 1H, $J = 5.85$ Hz, glucal H-1), 5.30 (d, 1H, $J = 3.22$ Hz, α-fucose H-1), 4.98 (d, 1H, $J = 11.72$ Hz, PhCH), 4.86 (d, 1H, $J = 10.99$ Hz, PhCH), 4.82 (d, 1H, $J = 11.64$ Hz, PhCH), 4.79-4.70 (m, 5H, included are PhCH, galactose H-1, H-4), 4.67-4.61 (m, 4H, included are PhCH, glucal H-2, α-fucose' H-1, $J = 3.76$ Hz), 4.56 (d, 1H, $J = 12.74$ Hz, PhCH), 4.55 (d, 1H, $J = 12.45$ Hz, PhCH), 4.49 (m, 2H,

PhCH and galactose H-3), 4.35 (m, 2H, glucal H-4 and a-fucose H-5), 4.25 (d, 1H, $J = 10.62$ Hz, PhCH), 4.15-4.07 (m, 4H, included are PhCH, a-fucose H-2, glucal H-3), 3.97-3.87 (m, 5H), 3.82 (t, 1H, $J = 9.38$ Hz), 3.71 (apparent t, 1H, $J = 7.16$ Hz, galactose H-2), 3.66-3.53 (m, 5H), 3.43 (d, 1H, $J = 1.76$ Hz, a-fucose' H-4), 3.36 (d, 1H, $J = 1.80$ Hz, a-fucose' H-4), 1.07 (s, 9H, *t*-butyl), 1.04 (two buried d, 6H, a-fucose' methyls), 1.01 (s, 9H, *t*-butyl); LRMS (FAB) calcd for $C_{99}H_{110}O_{18}Si_2K$ 1682, found m/z 1682 (M+K). The minor isomer 24c 480 mg (16%) containing the b-fucose linkage at the glucal center was obtained as a white foam: 1H NMR (400 MHz, $CDCl_3$): d 7.57 (m, 10H, ArH), 7.40-7.06 (m, 39H, ArH), 6.98 (m, 1H, ArH), 6.13 (d, 1H, $J = 6.22$ Hz, glucal H-1), 5.09 (d, 1H, $J = 3.62$ Hz, a-fucose H-1), 4.95 (d, 1H, $J = 11.72$ Hz, PhCH), 4.86 (d, 1H, $J = 11.92$ Hz, PhCH), 4.86 (m, 1H, buried glucal H-2), 4.81 (d, 1H, $J = 11.35$, PhCH), 4.76 (d, 1H, $J = 12.08$ Hz, PhCH), 4.70 (d, 1H, $J = 11.43$ Hz, PhCH), 4.70 (d, 1H, buried galactose H-1), 4.67-4.44 (m, 8H, galactose H-3, H-4, PhCH), 4.35 (d, 1H, $J = 11.64$ Hz, PhCH), 4.35 (d, 1H, $J = 7.48$ Hz, b-fucose H-1), 4.11-4.00 (m, 5H), 3.95 (q, 1H, $J = 6.31$ Hz, a-fucose H-5), 3.90 (m, 1H), 3.80 (s, 1H), 3.78 (s, 1H), 3.75 (dd, 1H, $J = 2.71$ Hz, $J = 10.32$ Hz, a-fucose H-3), 3.72-3.67 (m, 3H, galactose H-2), 3.53 (dd, 1H, $J = 7.84$ Hz, $J = 9.46$ Hz, b-fucose H-2), 3.49 (d, 1H, $J = 2.48$ Hz, a-fucose H-4), 3.37 (d, 1H, $J = 2.56$ Hz, b-fucose H-4), 3.21 (dd, 1H, $J = 2.93$ Hz, $J = 9.80$ Hz, b-fucose H-3), 3.18 (q, 1H, $J = 6.60$ Hz, b-fucose H-5), 1.05-0.97 (2 s, 18 H, *t*-butyl, and buried fucose methyls 6H).

[[[(2,3,4-Tri-*O*-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-*O*-*tert*-butyldiphenylsilyl-3,4-carbonate-b-D-galactopyranosyl)-(1-4)]-(2,3,4-tri-*O*-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]-6-*O*-*tert*-butyldiphenylsilyl-2-deoxy-2-iodo- α -D-manno-pyranosylbenzene sulfonamide (26c)

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To 1.95 g (1.19 mmol) of glycal 23c was added 561 mg (3.57 mmol) of benzenesulfonamide and 1.50 g of freshly dried 4 Å powdered molecular sieves. This mixture was taken up in 19 mL of dichloromethane and then cooled to 0°C. The reaction was allowed to stir for 15 minutes and then 1.95 g (4.17 mmol) of iodonium-di-sym-collidine perchlorate was added. The reaction was stirred for 30 minutes and then quenched with saturated sodium thiosulfate solution. The crude mixture was filtered through celite and the filtrate washed once with saturated sodium thiosulfate, once with saturated copper sulfate, and twice with saturated brine solution. The organics were dried over sodium sulfate, filtered and concentrated in vacuo. Purification by silica gel chromatography gave iodosulfonamide 26c as a white foam 2.12 g (93%): $[\alpha]_D^{25} = -78.3^\circ$ (c 0.935, CHCl₃); IR (thin film) 2920, 2835, 1800, 1450, 1425, 1350, 1150, 1100, 1050, 735, 700; ¹H NMR (400MHz, CDCl₃): δ 7.74-7.53 (m, 6H), 7.45-7.22 (m, 41H), 7.05 (t, 1H, $J = 7.24$ Hz), 6.90 (t, 2H, $J = 7.61$ Hz), 6.07 (d, 1H, $J = 8.43$ Hz), 5.55 (broad s, 1H), 5.01-4.42 (m, 18H), 4.31 (broad s, 1H), 4.02 (dd, 1H, $J = 3.66$ Hz, $J = 10.17$ Hz), 3.96-3.80 (m, 5H), 3.74 (broad d, $J = 10.26$ Hz), 3.66-3.60 (m, 3H), 3.48 (broad s, 1H), 3.41 (broad s, 1H), 3.19 (broad s, 1H), 1.06 (s, 9H), 0.99 (s, 9H), 0.95 (m, 6H); HRMS (FAB) calcd for C₁₀₅H₁₁₆NO₂₂ISi₂Na 1948.6290, found m/z 1948.6280 (M+Na).

[[[(2,3,4-Tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-O-tert-butyldiphenylsilyl-3,4-carbonate- β -D-galactopyranosyl)-(1-4)]-(2,3,4-tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]--6-O-tert-butyldiphenylsilyl-2-deoxy-2-benzenesulfonamido- α -D-glucopyranosyl fluoride (25c)

To 740 mg (384 mmol) of iodosulfonamide 26c was added 35 mL of 5:2 THF/water. To this solution was added 107 mL (768 mmol) of triethylamine and 106 mg (384 mmol) of

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silver carbonate. The reaction was stirred for one hour after which time TLC indicated complete consumption of starting material. The reaction was partitioned between ethyl acetate and brine and the organics were washed twice with saturated brine solution. The organics were dried over sodium sulfate, filtered and concentrated in vacuo. The crude free reducing sugar was azeotroped once with benzene and then taken up in 2 mL of THF. The solution was cooled to -30°C and 53 mL (403 mmol) of DAST was added. The reaction was warmed to room temperature and stirred for an additional 10 minutes. The reaction was then cooled back down to -30°C and quenched with 1 mL of MeOH. The reaction was concentrated in vacuo. Purification by column chromatography gave fluorosugar 25c as a white foam 593 mg (85%): $[\alpha]_D^{23} = -34.1^\circ$ (c 1.29, CHCl_3); IR (thin film): 2920, 1800, 1450, 1160, 1100, 1045, 915, 735, 700; ^1H NMR (400 MHz, CDCl_3): δ 7.78 (d, 2H, $J = 7.98$ Hz), 7.64-6.96 (m, 48H), 6.62 (broad s, 1H), 5.89 (dd, 1H, $J = 2.11$ Hz, $J = 54.55$ Hz, glucose H-1), 4.99 (d, 1H, $J = 11.44$ Hz), 4.94-4.39 (m, 21H), 4.03-3.97 (m, 4H), 3.84-3.77 (m, 6H), 3.69 (m, 7H), 3.59 (m, 2H), 3.49 (m, 5H), 3.39 (m, 1H), 3.29-3.18 (m, 2H), 1.38 (d, 3H, $J = 6.23$ Hz), 1.02 (s, 9H), 0.97 (s, 9H); LRMS (FAB) calcd for $\text{C}_{105}\text{H}_{116}\text{O}_{20}\text{NSFSi}_2\text{Na}$ 1840.7230, found m/z 1840.7220 (M+Na).

(3R)-O-Benzoyl-(2R)-hexadecanamido-4-octadecenyl-[[[(2,3,4-tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-2)]-(6-O-tert-butylidiphenylsilyl-3,4-carbonate-b-D-galactopyranosyl)-(1-4)]-(2,3,4-tri-O-benzyl-6-deoxy- α -L-galactopyranosyl)-(1-3)]--6-O-tert-butylidiphenylsilyl-2-deoxy-2-benzenesulfonamido-b-D-glucopyranose (31c)

To 97 mg (53 mmol) of fluorosugar 25c was added 34 mg (80 mmol) of azidosphingosine 27c. The mixture was azeotroped twice with benzene and then placed under high vacuum for two hours. 100 mg of freshly activated 4 Å

powdered molecular sieves were added and then 5 mL of dry dichloromethane. In an argon glove box were added 47 mg (160 mmol) of zirconocene dichloride and then 41 mg (160 mmol) of silver triflate. The reaction was stirred overnight. The reaction was quenched with saturated sodium bicarbonate and filtered through a celite pad. The filtrate was washed twice with saturated sodium bicarbonate and twice with saturated brine solution. The organics were dried over sodium sulfate, filtered and concentrated in vacuo. Purification by silica gel chromatography (15-20% EtOAc/hexanes) gave the labile azide 28c 57 mg (50%) as a white foam: IR (thin film): 2920, 2845, 2100, 1810, 1715, 1450, 1350, 1260, 1150, 1100, 1050, 740, 700; ¹H NMR (400 MHz, CDCl₃): δ 8.04 (d, 2H, J = 7.98 Hz), 7.64-7.55 (m, 13H), 7.50-7.10 (m, 40H), 6.97 (m, 2H), 6.87 (m, 2H), 6.80 (d, 1H, J = 3.58 Hz), 5.75 (m, 1H), 5.37 (dd, 1H, J = 7.57 Hz, J = 15.74 Hz), 5.27 (m, 2H), 5.09 (dd, 1H, J = 4.44, J = 8.02 Hz), 4.98 (m, 2H), 4.87-4.40 (m, 18H), 4.31 (d, 1H, J = 4.03 Hz), 4.29 (d, 1H, J = 8.06 Hz), 4.16-4.05 (m, 5H), 3.95 (m, 2H), 3.88 (m, 2H), 3.78 (m, 3H), 3.70-3.53 (m, 6H), 3.46 (broad s, 1H), 3.40 (m, 3H), 3.26 (m, 2H), 2.84 (d, 1H, J = 8.87 Hz), 2.69 (dd, 1H, J = 2.85 Hz, J = 9.52 Hz), 2.57 (m, 1H), 2.04 (m, 2H), 1.21 (broad s, 16H), 1.07 (d, 3H, J = 6.79 Hz), 1.01 (m, 24H) HRMS (FAB) calcd for C₁₃₀H₁₅₄O₂₃N₄SSi₂Na 2251.0190, found m/z 2251.0197 (M+Na). Azidosphingosine 28c 57 mg (26 mmol) was taken up in 1 mL of ethyl acetate. 100 mg of Lindlar's catalyst and 25 mg (51 mmol) of palmitic anhydride were added. The reaction mixture was degassed and placed under an atmosphere of hydrogen. The reaction appeared complete after 36 hours though there appeared to be extensive decomposition. The reaction mixture was degassed and filtered thru a pad of celite. The filtrate was concentrated in vacuo. The crude material was purified first by gel filtration with LH-20 (lipophilic sephadex, MeOH eluant) and then by silica gel chromatography (15%

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- EtOAc/hexanes) to afford 13.6 mg (22%) of the ceramide linked Lewis^y antigen 31c as a white foam: IR (thin film): 2920, 2855, 1830, 1810, 1720, 1650, 1450, 1430, 1320, 1270, 1165, 1110, 1055, 830, 745, 705; ¹H NMR (400 MHz, CDCl₃): δ 7.99 (t, 2H, J = 8.24 Hz), 7.74-7.05 (m, H), 6.94 (m, 3H), 6.76 (broad d, 1H, J = 12.35 Hz), 6.60 (m, 1H), 5.80 (m, 1H), 5.59 (t, 1H, J = 7.89 Hz), 5.41 (dd, 1H, J = 7.85 Hz, J = 15.42 Hz), 5.24 (m, 1H), 5.18 (m, 1H), 5.00-4.21 (m, 18H), 4.06 (broad d, 1H, J = 9.88 Hz), 3.97-3.44 (m, 15H), 3.16 (m, 2H), 2.95 (t, 1H, J = 8.43 Hz), 2.08-1.97 (m, 3H), 1.24 (broad s, 54H), 1.00-.99 (2s, 18H), 0.88 (m, 12H); HRMS (FAB) calcd for C₁₀₅H₁₁₆O₂₀NSFSi₂Na 1840.7230, found m/z 1840.7220 (M+Na).
- 15 (3R)-Hydroxy-(2R)-hexadecanamido-4-octadecenyl-[[[(6-deoxy-α-L-galactopyranosyl)-(1→2)]-(β-D-galactopyranosyl)-(1→4)]-(6-deoxy-α-L-galactopyranosyl)-(1→3)]-2-deoxy-2-acetamido-β-D-glucopyranose (32c)
- 20 To 13.6 mg (5.5 mmol) of blocked Lewis^y ceramide was added 100 mL of THF and 20 mL (20 mmol) of TBAF (1M in THF). The reaction was stirred overnight and then concentrated in vacuo. The crude mixture was dissolved in 200 mL of THF and added to a solution of 100 mg of sodium metal in
- 25 3 mL of liquid ammonia at -78°C. The blue solution was allowed to stir for 45 minutes at -78°C and was then quenched carefully with 1 mL of MeOH. The excess ammonia was removed with a rapid stream of argon and then the crude reaction mixture was acidified to a pH of 7 with
- 30 Dowex 50 X-8 resin. The solution was then filtered and the resin washed with methanol that had been saturated with ammonia. The filtrate was then concentrated in vacuo and the solids dissolved in 200 mL of pyridine and 200 mL of acetic anhydride. The reaction was allowed to
- 35 proceed for 8 hours at room temperature and was then concentrated in vacuo. Purification by silica chromatography (20% acetone/benzene) gave 2.0 mg of the

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peracetylated ceramide antigen. The peracetylated product was taken up in 100 mL of dry MeOH and 3 mg of NaOMe was added and the reaction stirred overnight. The reaction was acidified with Dowex 50 X-8 resin to pH 7 and then filtered and concentrated. The product 32c was purified by gel filtration with LH-20 (lipophilic sephadex, MeOH eluant) giving 1.1 mg (17%) of a white solid: HRMS (FAB) calcd for $C_{60}H_{110}O_2N_2Na$ 1217.7500, found m/z 1217.7534 (M+Na).

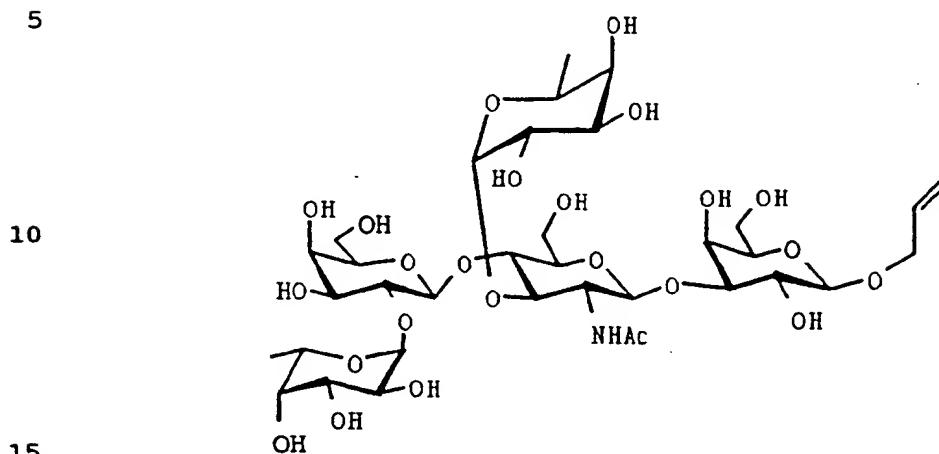
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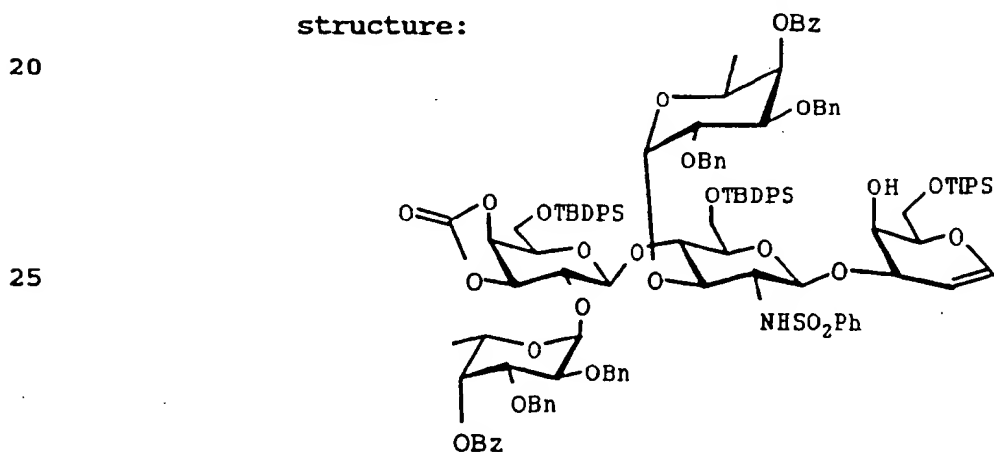
What is claimed is:

1. A method of synthesizing an allyl pentasaccharide having the structure:



which comprises:

- (a) (i) de-silylating a compound having the structure:



with $R^1R^2R^3R^4NF$ wherein R^1 , R^2 , R^3 , and R^4 are independently a linear or branched chain alkyl, aralkyl or aryl, to form an N-sulfonamide pentasaccharide;

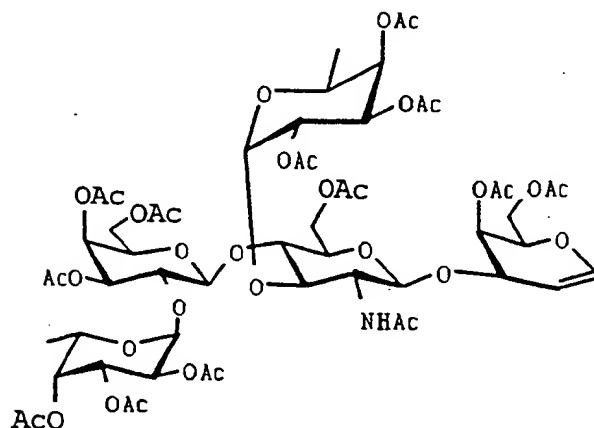
(ii) cleaving the N-sulfonamide pentasaccharide formed in step (a)(i) to form a deprotected pentasaccharide;

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and

(iii) acetylating the deprotected penta-saccharide formed in step (b)(ii) to form a peracetate having the structure:



(b) (i) treating the peracetate formed in step (a)(iii) with an epoxidizing agent to form an epoxide peracetate;

(ii) reacting the epoxide peracetate formed in step (b)(i) with allyl alcohol to form an allylglycoside peracetate; and

(iii) cleaving the allylglycoside peracetate with an alkoxide salt to form the allyl penta-saccharide.

2. The method of claim 1 wherein R^1 , R^2 , R^3 , and R^4 in step (a)(i) are n-butyl.

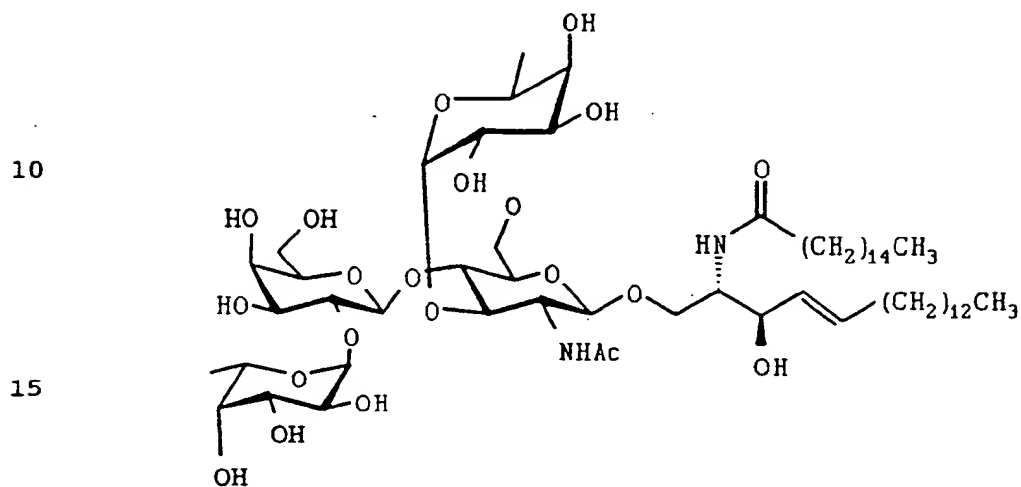
3. The method of claim 1 wherein the cleaving step (a)(ii) is performed with Na/NH_3 .

4. The method of claim 1 wherein the epoxidizing agent of step (b)(i) is 3,3-dimethyldioxirane.

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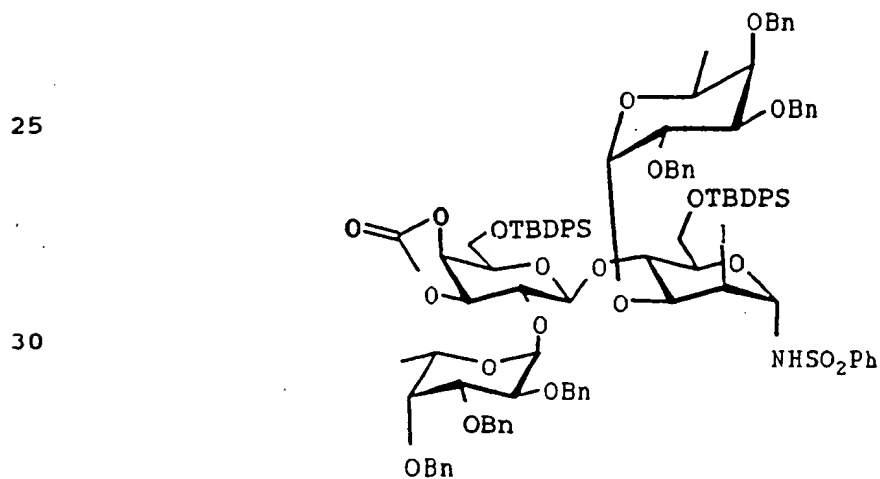
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5. The method of claim 1 wherein the alkoxide salt of step (b)(iii) is NaOMe.
6. A method of synthesizing a ceramide having the structure:



which comprises:

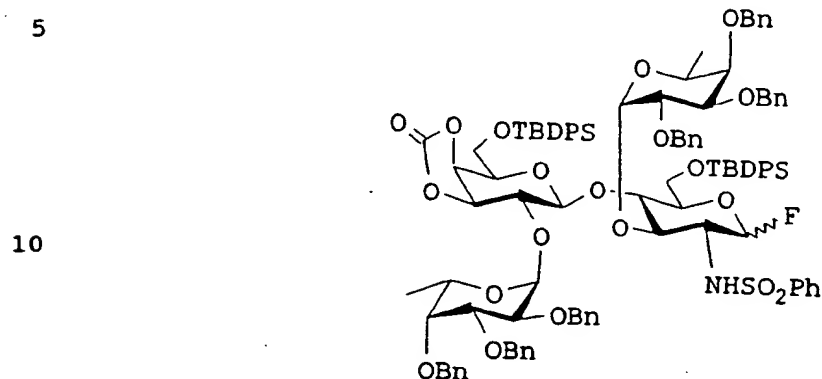
- (a) treating a compound having the structure:



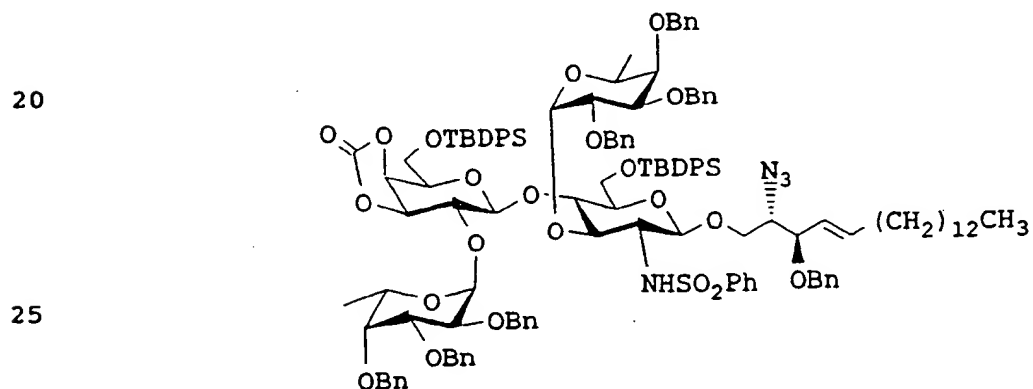
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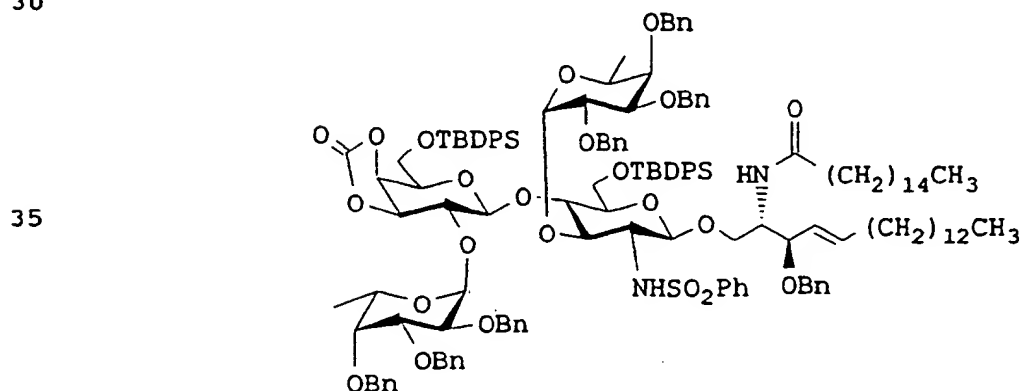
with aqueous silver(I) followed by reacting with diethylaminosulfur trifluoride to form a compound having the structure:



- 15 (b) treating the compound formed in step (a) with azidosphingosine to form a compound having the structure:



- 30 (c) reducing the compound formed in step (b) to form a compound having the structure:



SUBSTITUTE SHEET (RULE 26)

and

5 (d) (i) de-silylating the compound formed in step (c) with $R^1R^2R^3R^4NF$ wherein R^1 , R^2 , R^3 , and R^4 are independently a linear or branched chain alkyl, aralkyl or aryl;

10 (ii) reductively cleaving the compound formed in step (d)(i) to form a polyalcohol tetrasaccharide;

15 (iii) peracetylating the polyalcohol tetrasaccharide to form a peracetate tetrasaccharide; and

(iv) saponifying the peracetate tetrasaccharide with a metal alkoxide to form the ceramide.

20 7. The method of claim 6 wherein silver(I) in step (a) is silver carbonate.

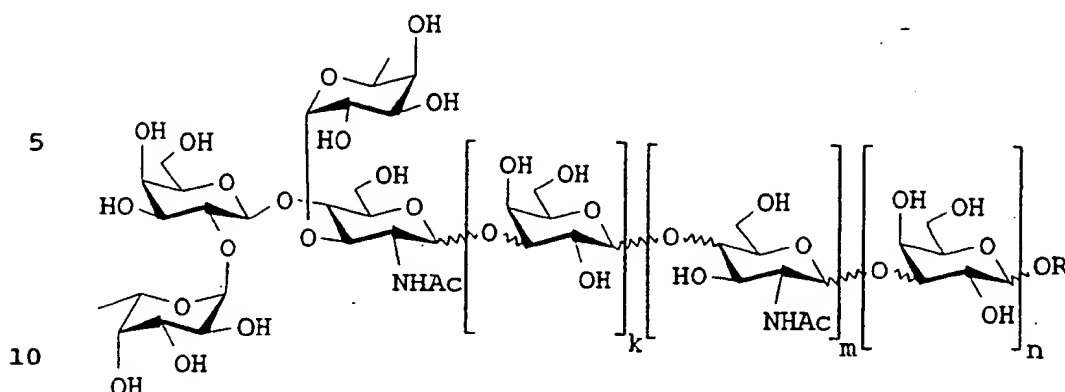
25 8. The method of claim 6 wherein step (b) is performed in the presence of zirconocene dichloride and silver triflate.

9. The method of claim 6 wherein the reducing step (c) is performed using hydrogen gas and palmitic anhydride in the presence of Lindlar's catalyst.

30 10. The method of claim 6 wherein R^1 , R^2 , R^3 , and R^4 in step (d)(i) are n-butyl and step (d)(ii) is performed using Na/NH_3 .

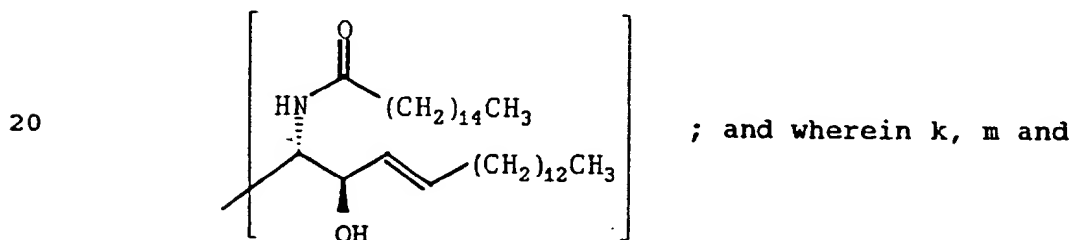
-140-

11. A compound having the structure:



15

wherein R is H, substituted or unsubstituted alkyl, aryl or allyl, or an amino acyl moiety, an amino acyl residue of a peptide, an amino acyl residue of a protein, which amino acyl moiety or residue bears an ω -amino group or an ω -(C=O)- group, which group is linked to O via a polymethylene chain having the structure $-(CH_2)_r-$, where r is an integer between about 1 and 9, or a moiety having the structure:

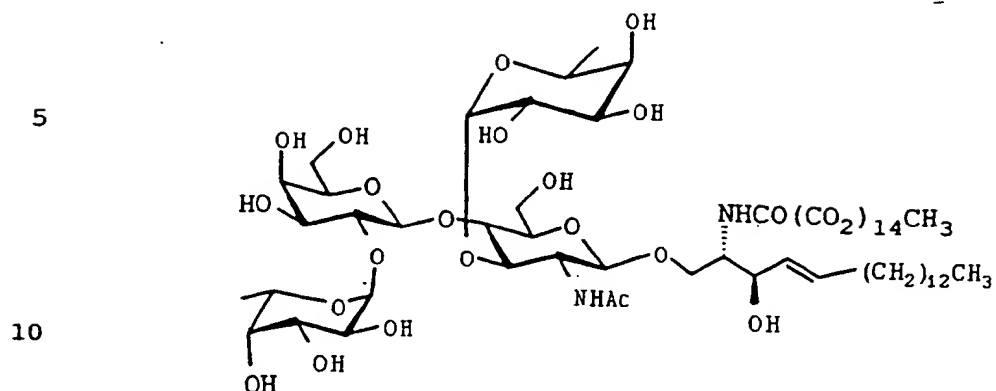


n are independently 0, 1, 2 or 3.

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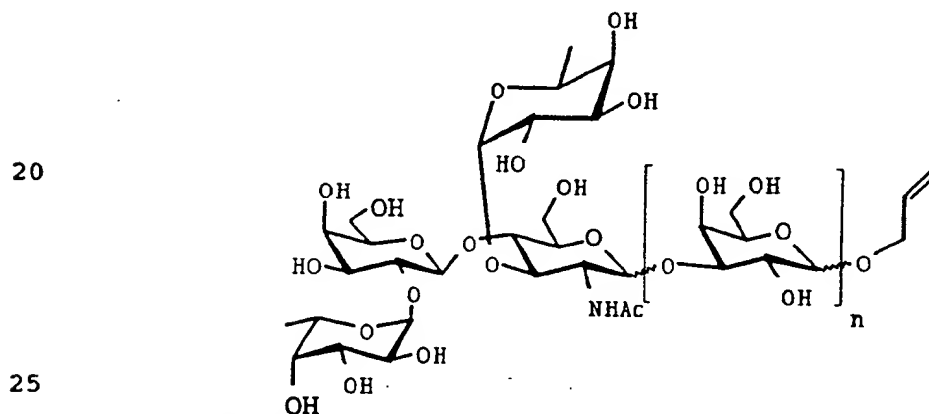
-141-

12. The compound of claim 11 having the structure:



13. The compound of claim 13 wherein the protein is bovine serum albumin or KLH.

14. A compound having the structure:



wherein n is 0, 1, 2, 3 or 4.

15. The compound of claim 13 wherein n is 1 or 2.

SUBSTITUTE SHEET (RULE 26)

-

-

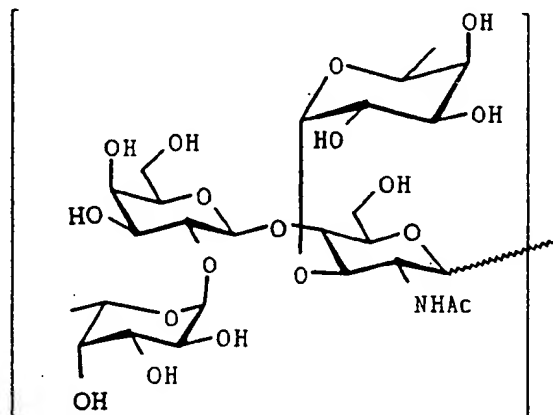
effective to induce the antibodies.

18. The method of claim 17 wherein the compound is bound to a suitable carrier protein.
19. The method of claim 18 wherein the carrier protein is bovine serum albumin, polylysine, or KLH.

BNSDOCID: <WO_____9703995A1 | >

-143-

20. The method of claim 17 which further comprises co-administering an immunological adjuvant.
21. The method of claim 19 wherein the adjuvant is bacteria or liposomes.
22. The method of claim 19 wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21.
23. The method of claim 17 wherein the compound is the compound of claim 11, 13 or 16.
24. The method of claim 17 wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.
25. A method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with colon tumor cells, which comprises administering to the subject an amount of a compound which contains a Le^x tetrasaccharide determinant having the structure:



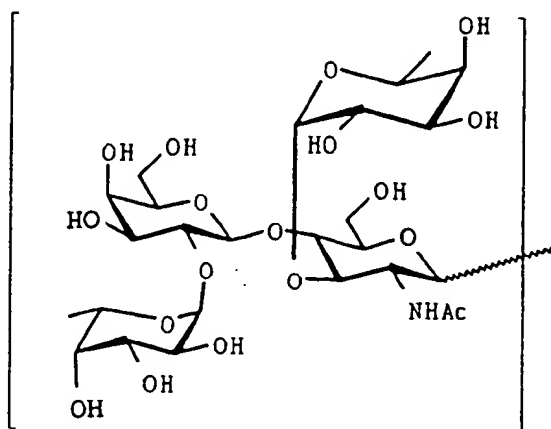
effective to induce the antibodies.

26. The method of claim 25 wherein the compound is bound to a suitable carrier protein.

SUBSTITUTE SHEET (RULE 26)

-144-

27. The method of claim 26 wherein the carrier protein is bovine serum albumin, polylysine, or KLH.
28. The method of claim 25 which further comprises co-administering an immunological adjuvant.
29. The method of claim 28 wherein the adjuvant is bacteria or liposomes.
30. The method of claim 28 wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21.
31. The method of claim 25 wherein the compound is the compound of claim 11, 13 or 16.
32. The method of claim 25 wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.
33. A method of inducing antibodies in a subject, wherein the antibodies are capable of specifically binding with ovarian tumor cells, which comprises administering to the subject an amount of a compound which contains a Le^x tetrasaccharide determinant having the structure:



effective to induce the antibodies.

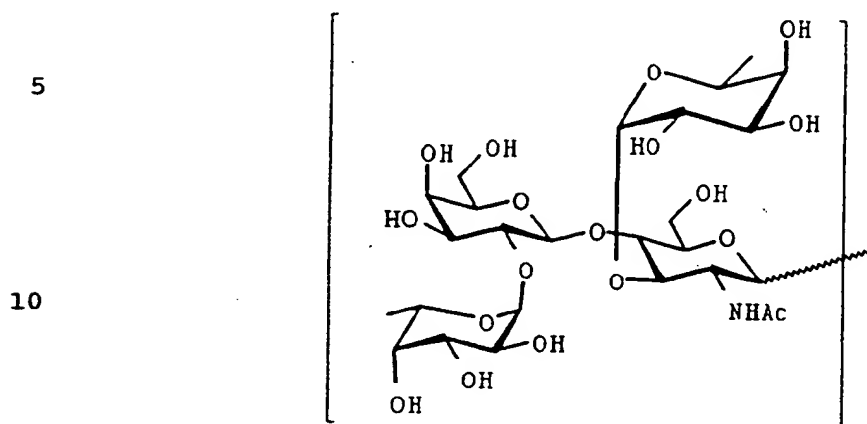
SUBSTITUTE SHEET (RULE 26)

-145-

34. The method of claim 33 wherein the compound is bound to a suitable carrier protein.
- 5 35. The method of claim 34 wherein the carrier protein is bovine serum albumin, polylysine, or KLH.
36. The method of claim 33 which further comprises co-administering an immunological adjuvant.
- 10 37. The method of claim 36 wherein the adjuvant is bacteria or liposomes.
38. The method of claim 36 wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21.
- 15 39. The method of claim 33 wherein the compound is the compound of claim 11, 13 or 16.
- 20 40. The method of claim 33 wherein the subject is in clinical remission or, where the subject has been treated by surgery, has limited unresected disease.
- 25 41. A method of preventing recurrence of epithelial cancer in a subject which comprises vaccinating the

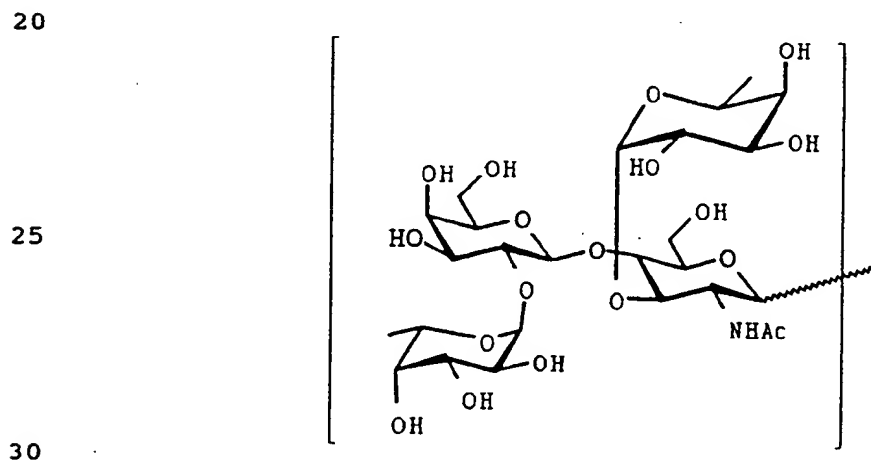
-146-

subject with a compound which contains a Le^x tetrasaccharide determinant having the structure:



effective to induce the antibodies.

- 15
42. A method of preventing recurrence of colon cancer in a subject which comprises vaccinating the subject with a compound which contains a Le^x tetrasaccharide determinant having the structure:



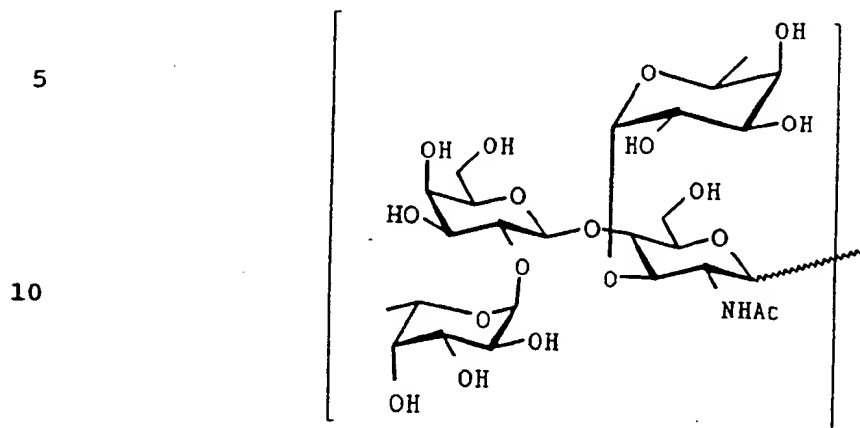
effective to induce the antibodies.

43. A method of preventing recurrence of ovarian cancer in a subject which comprises vaccinating the subject

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with a compound which contains a Le^x tetrasaccharide determinant having the structure:



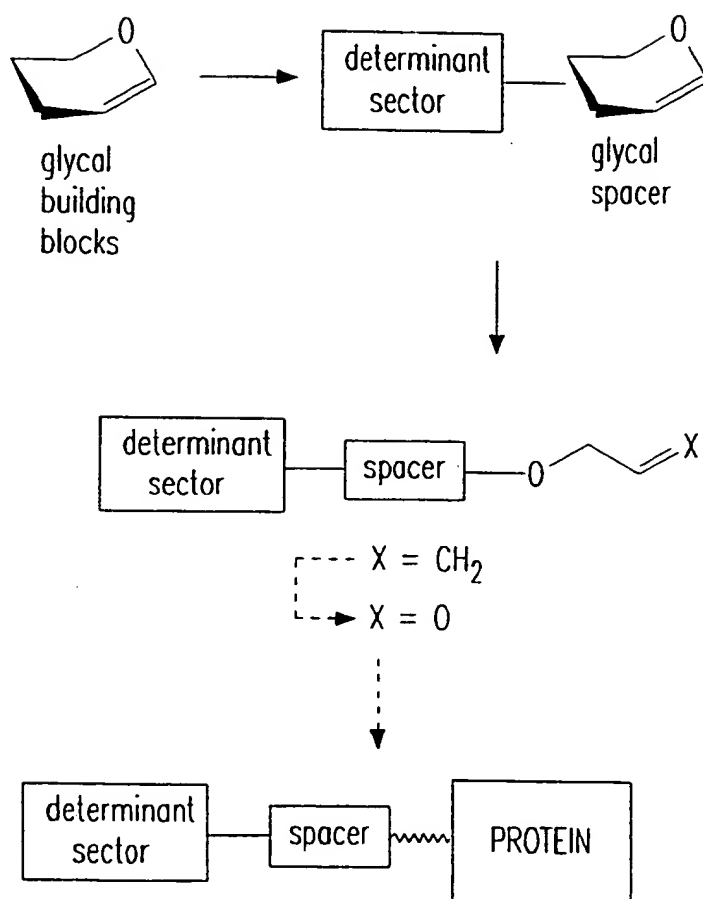
effective to induce the antibodies.

44. The method of claim 41, 42 or 43 wherein the compound is bound to a suitable carrier protein.
45. The method of claim 41, 42 or 43 wherein the carrier protein is bovine serum albumin, polylysine, or KLH.
46. The method of claim 41, 42 or 43 which further comprises coadministering an immunological adjuvant.
47. The method of claim 46 wherein the adjuvant is bacteria or liposomes.
48. The method of claim 46 wherein the adjuvant is *Salmonella minnesota* cells, bacille Calmette-Guerin, or QS21.

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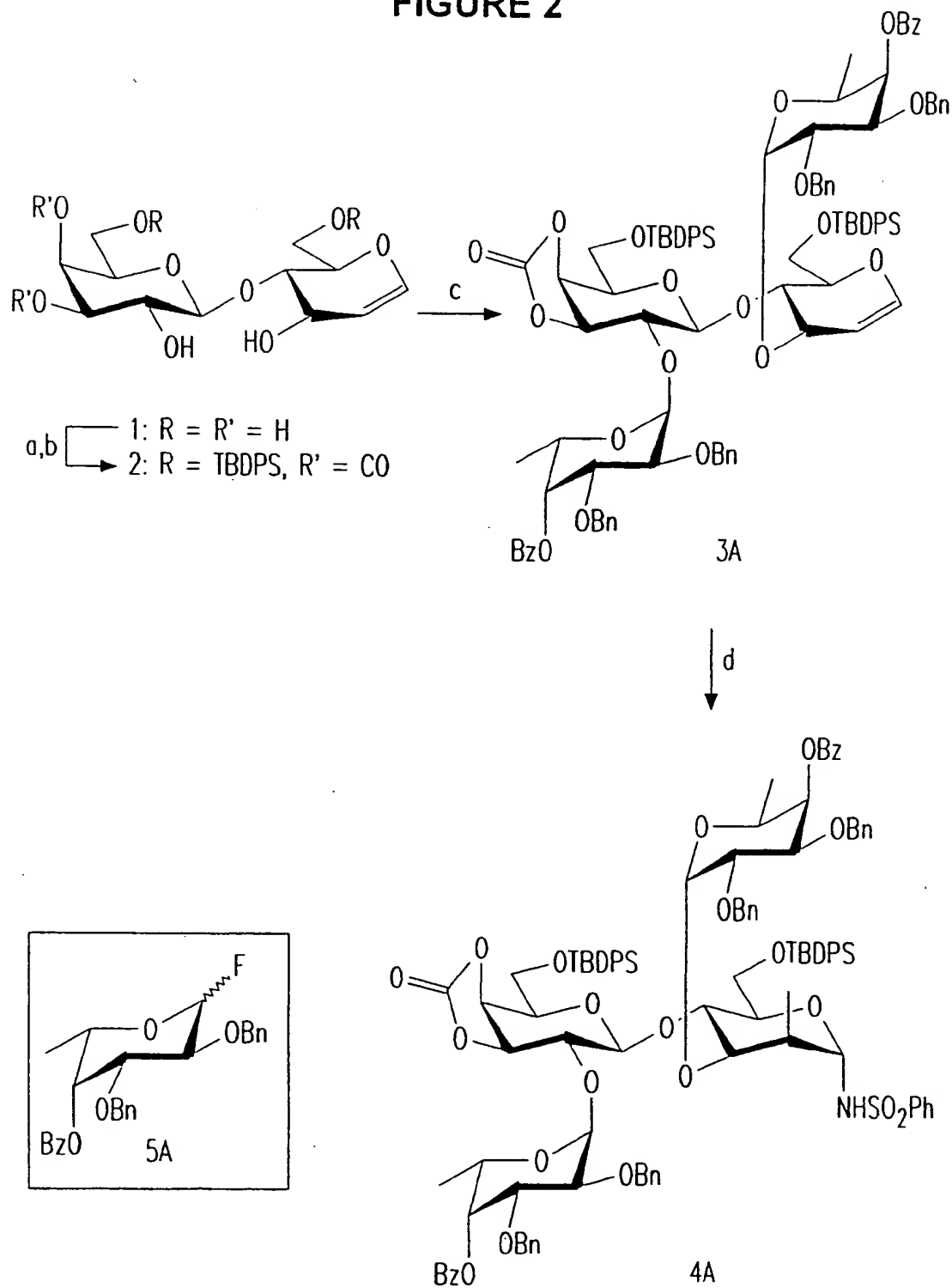
1/34

FIGURE 1



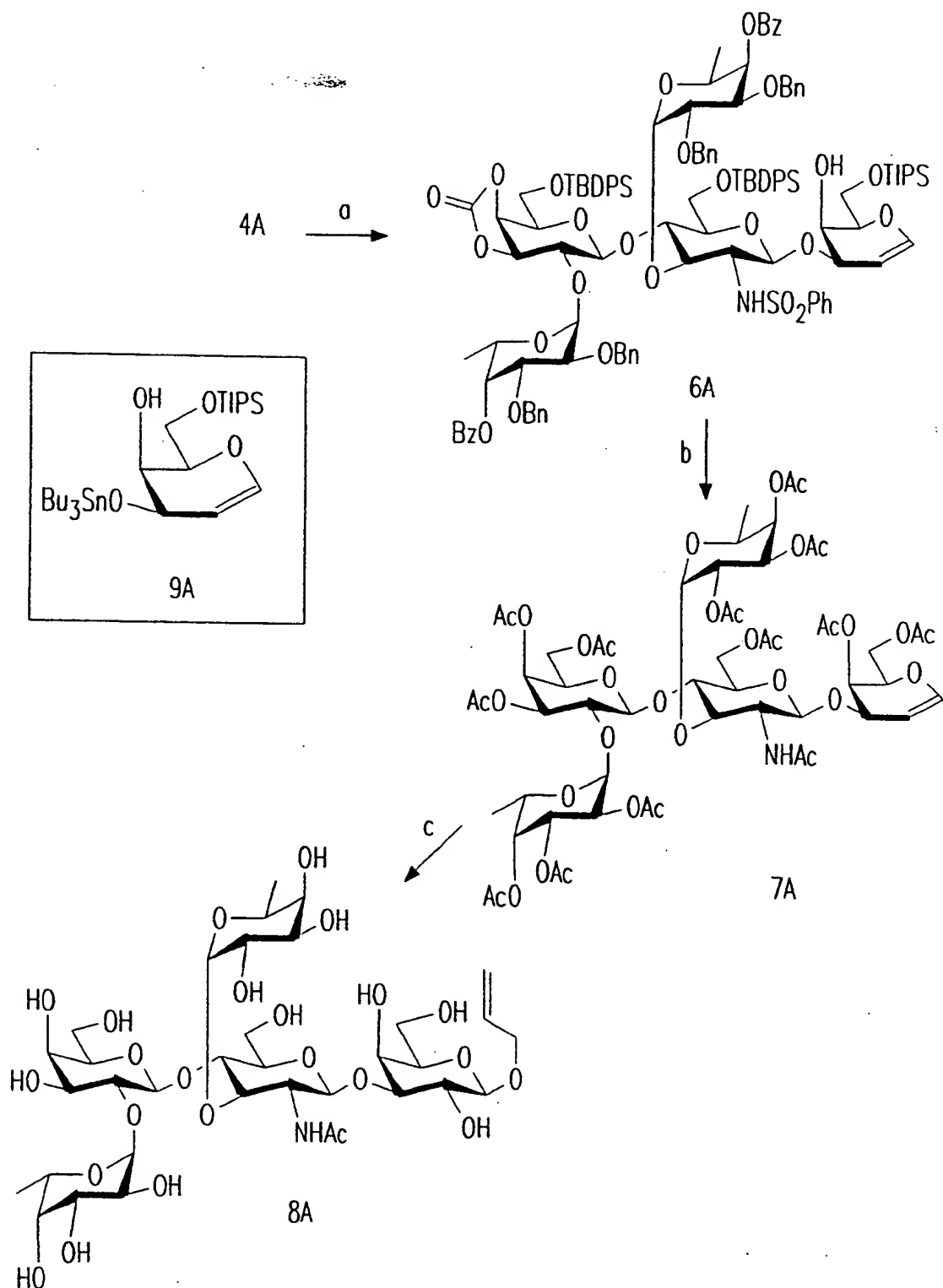
2/34

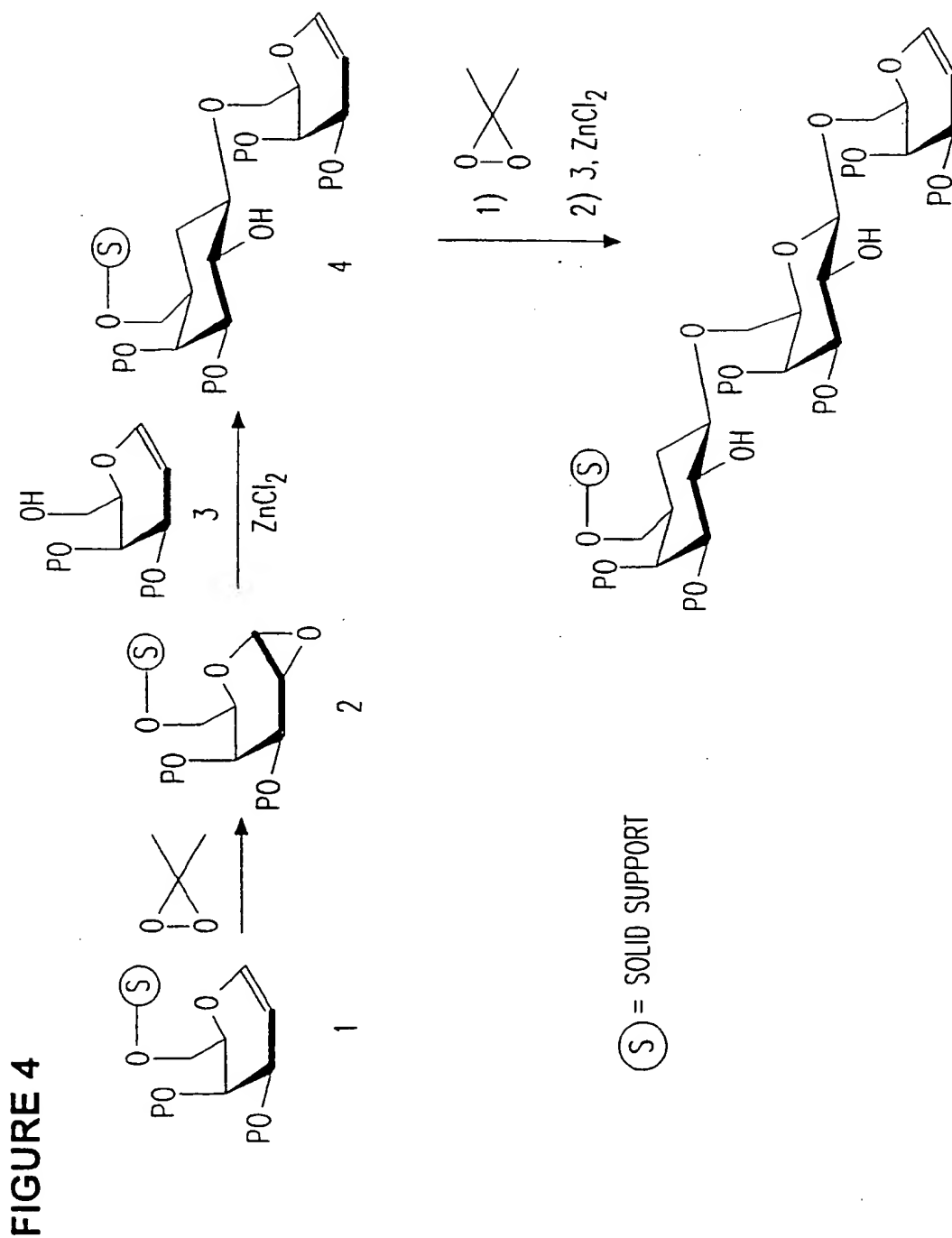
FIGURE 2



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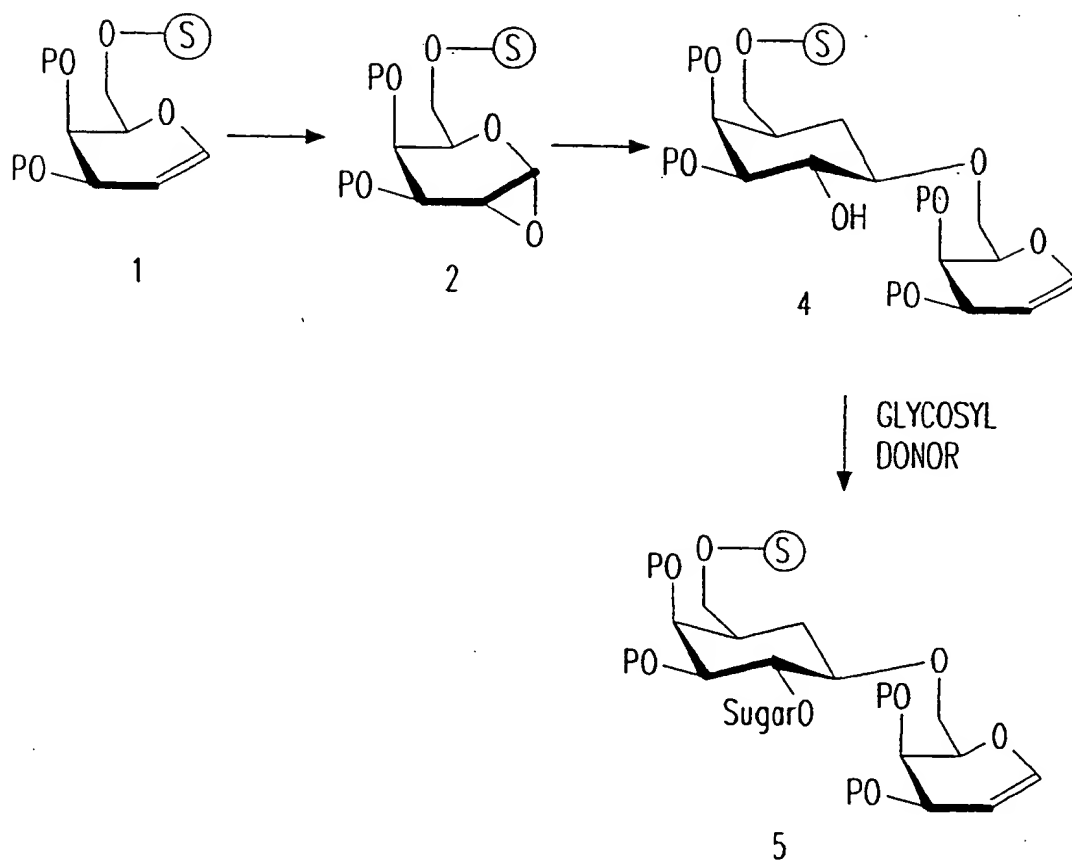
FIGURE 3





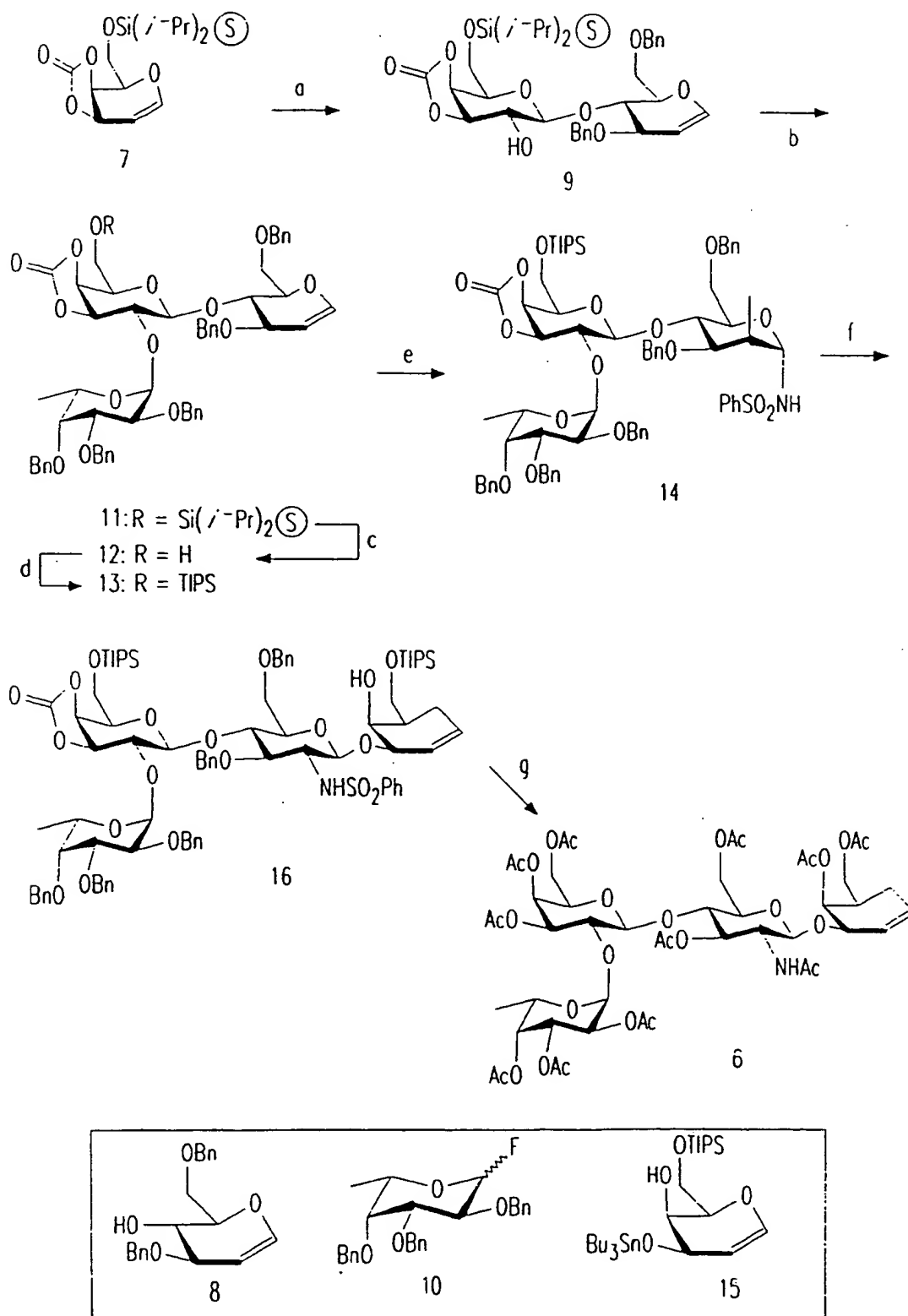
5/34

FIGURE 5



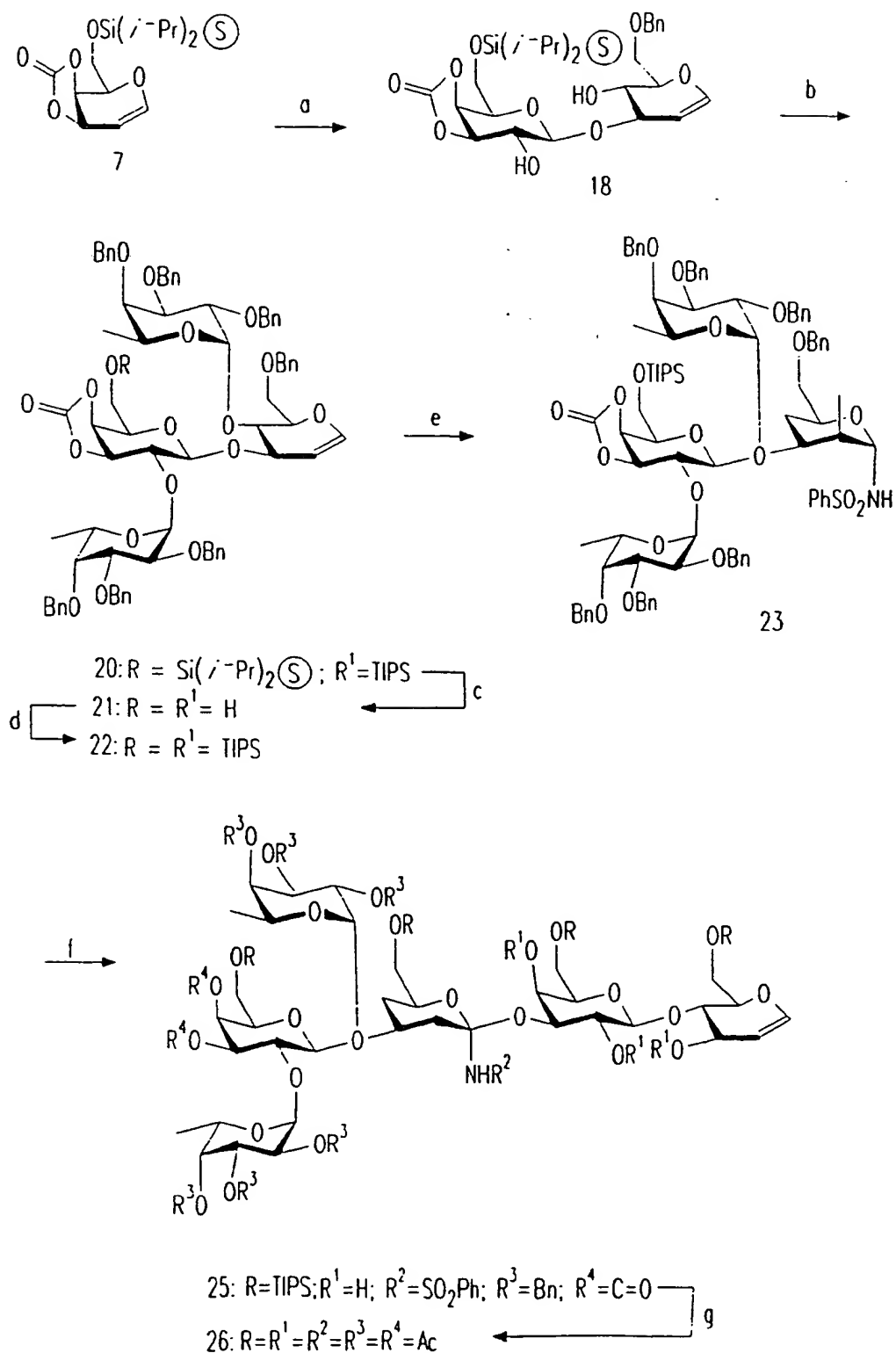
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FIGURE 6



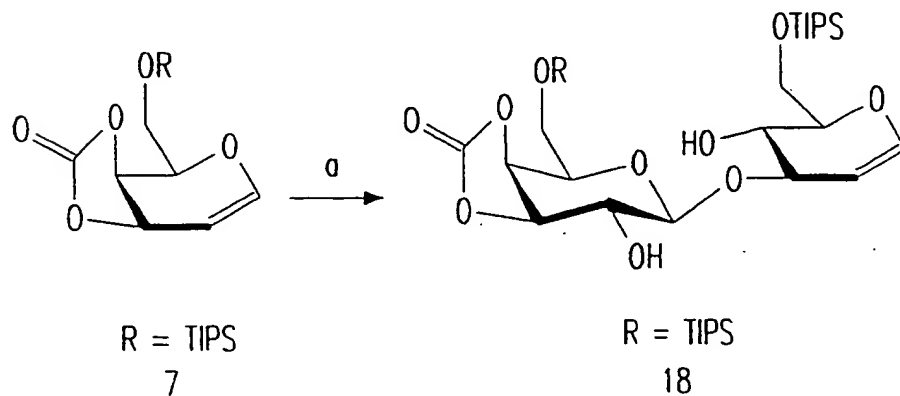
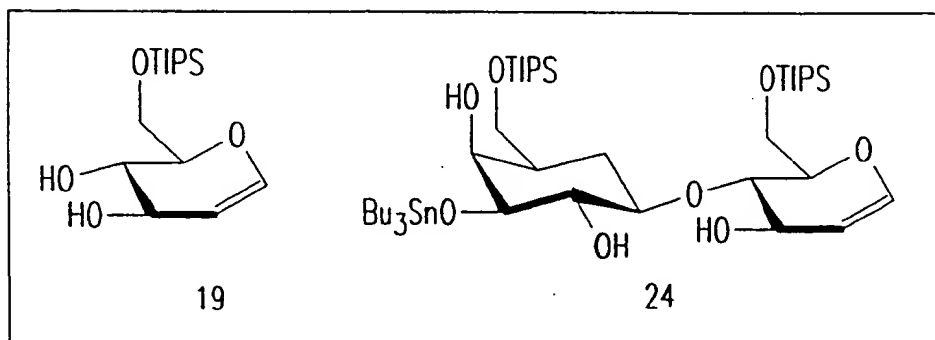
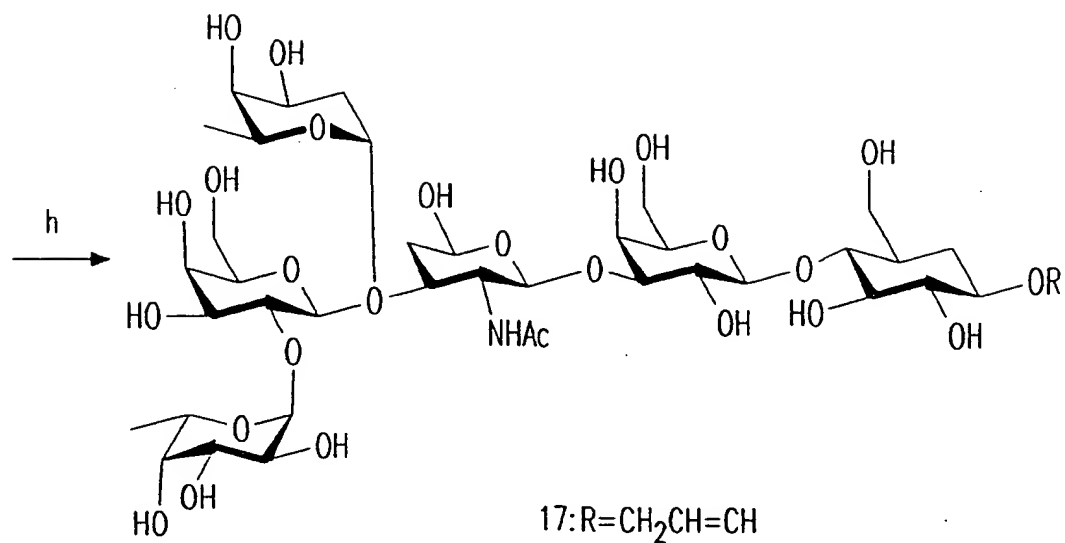
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FIGURE 7A



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FIGURE 7B



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FIGURE 8A

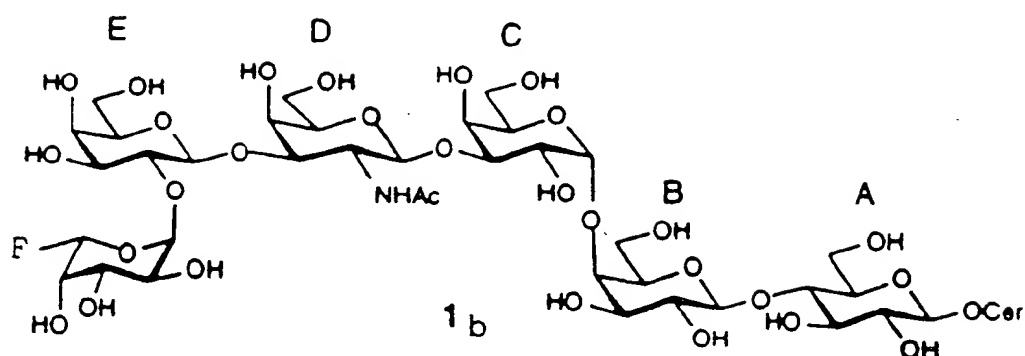
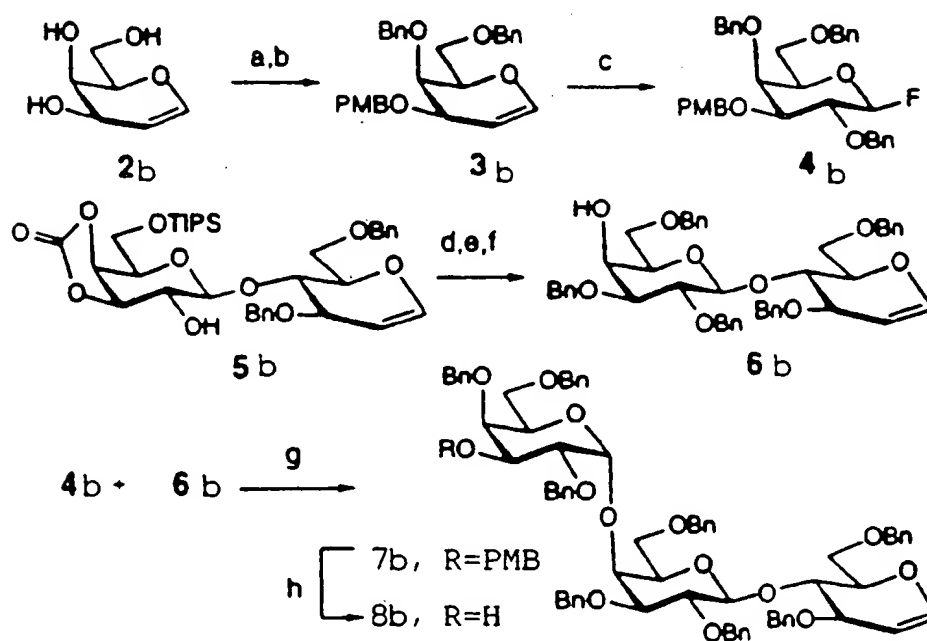
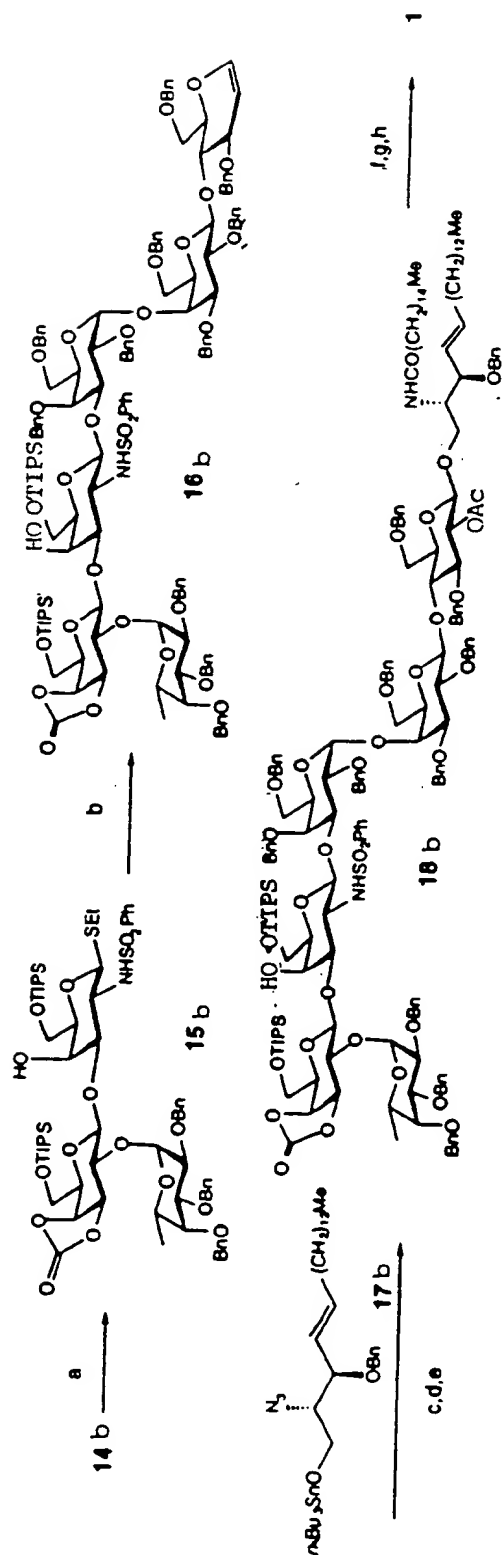


FIGURE 8B



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FIGURE 9



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FIGURE 10A

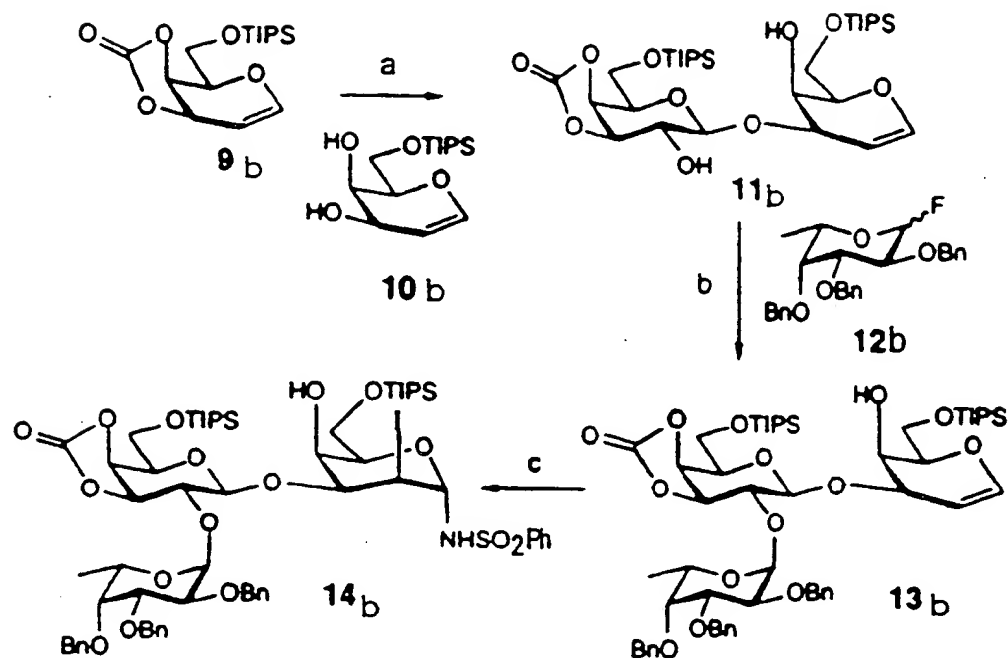
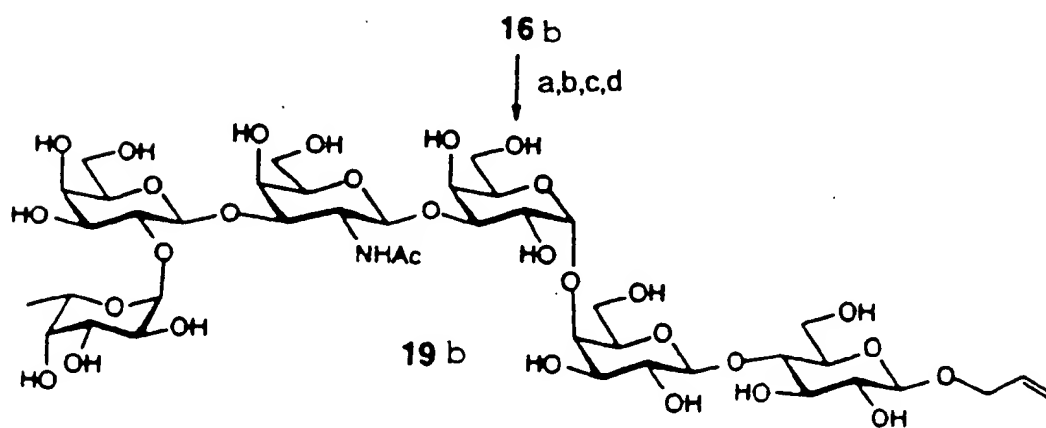
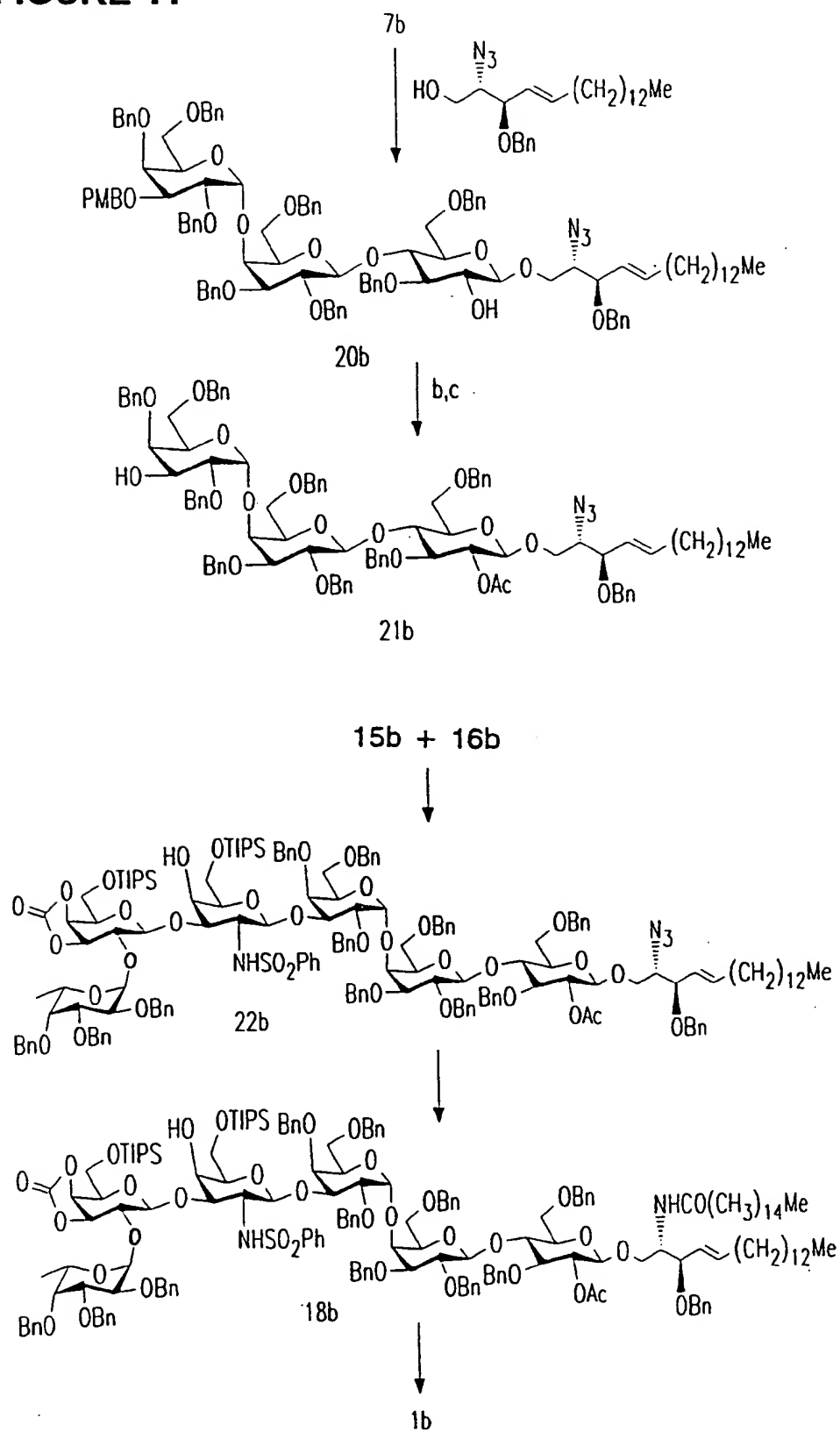


FIGURE 10B

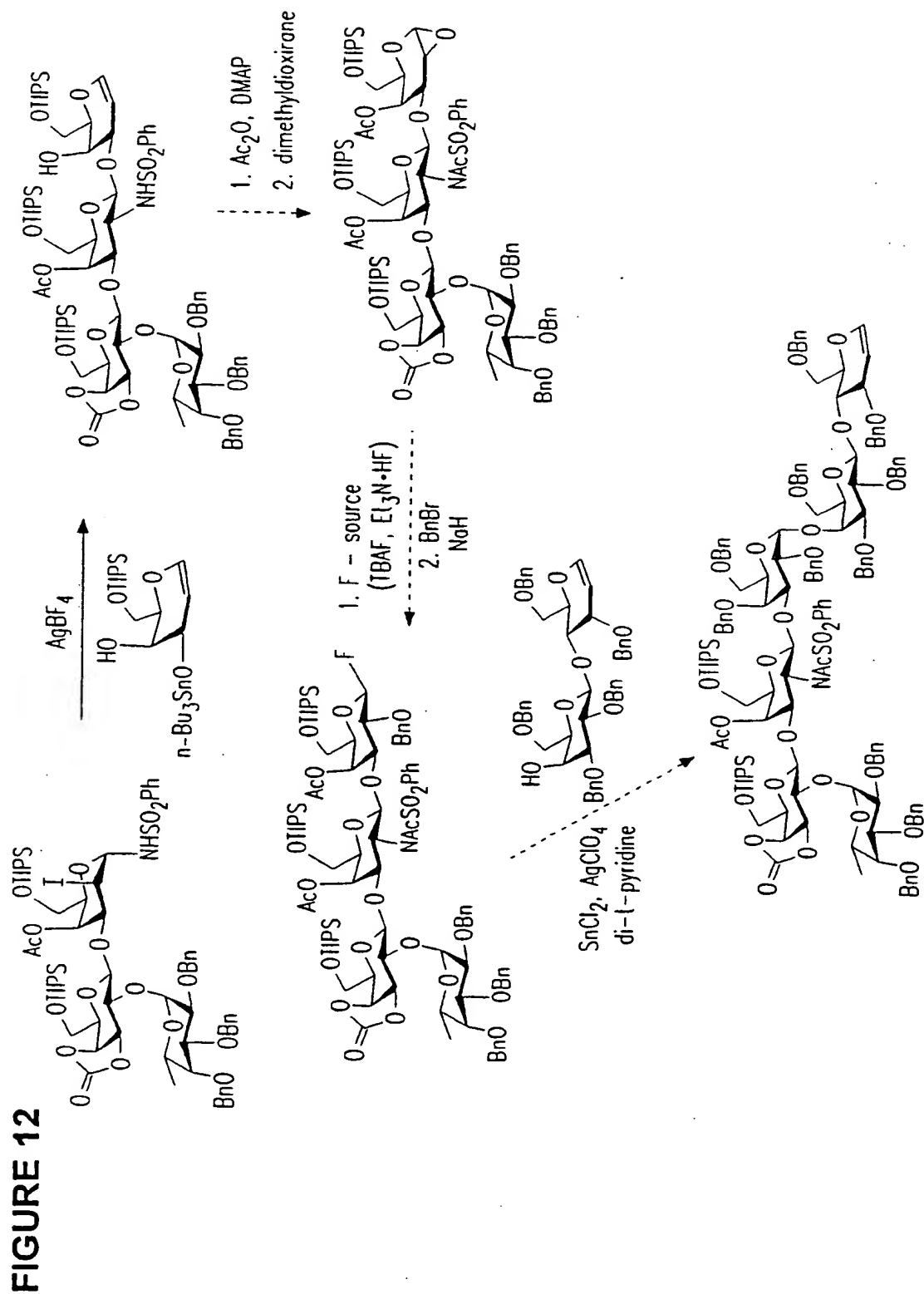


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FIGURE 11

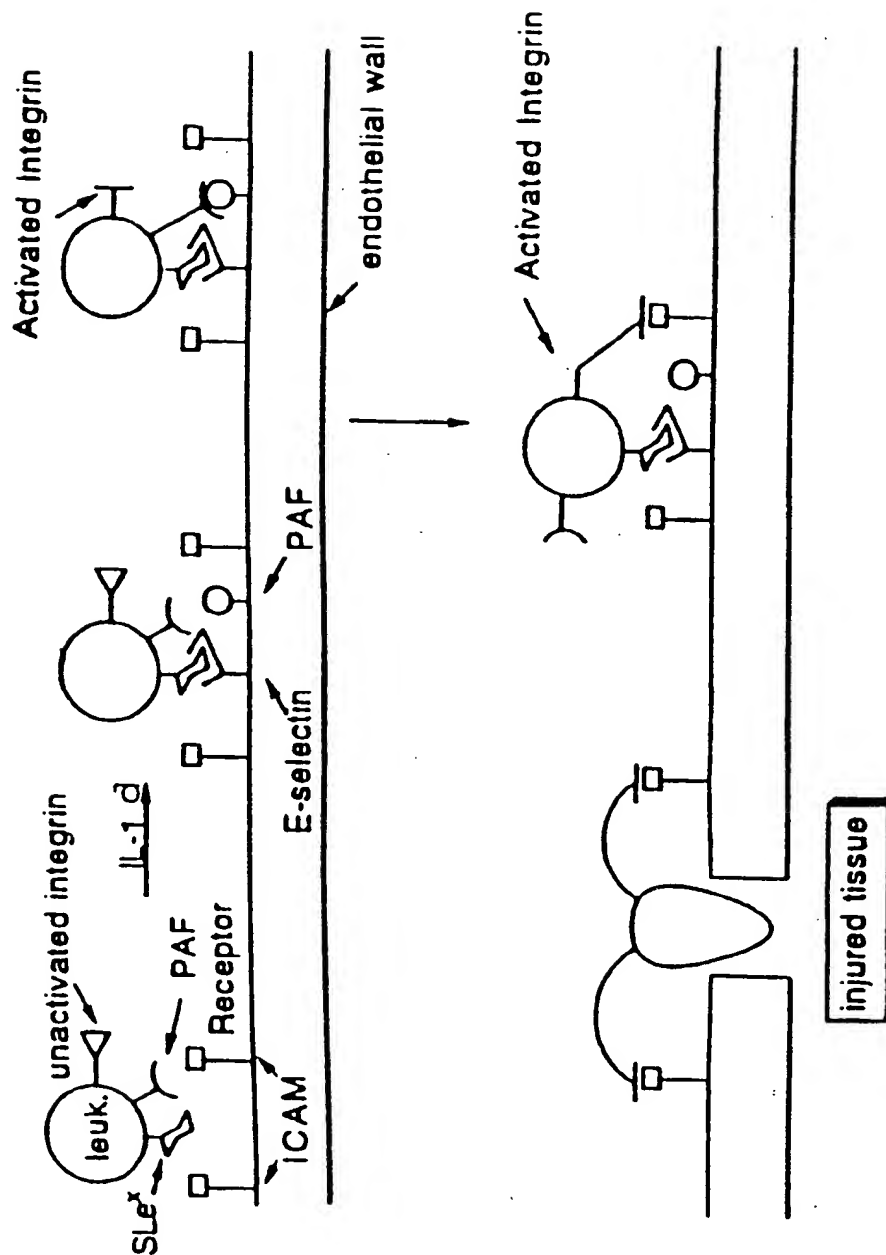


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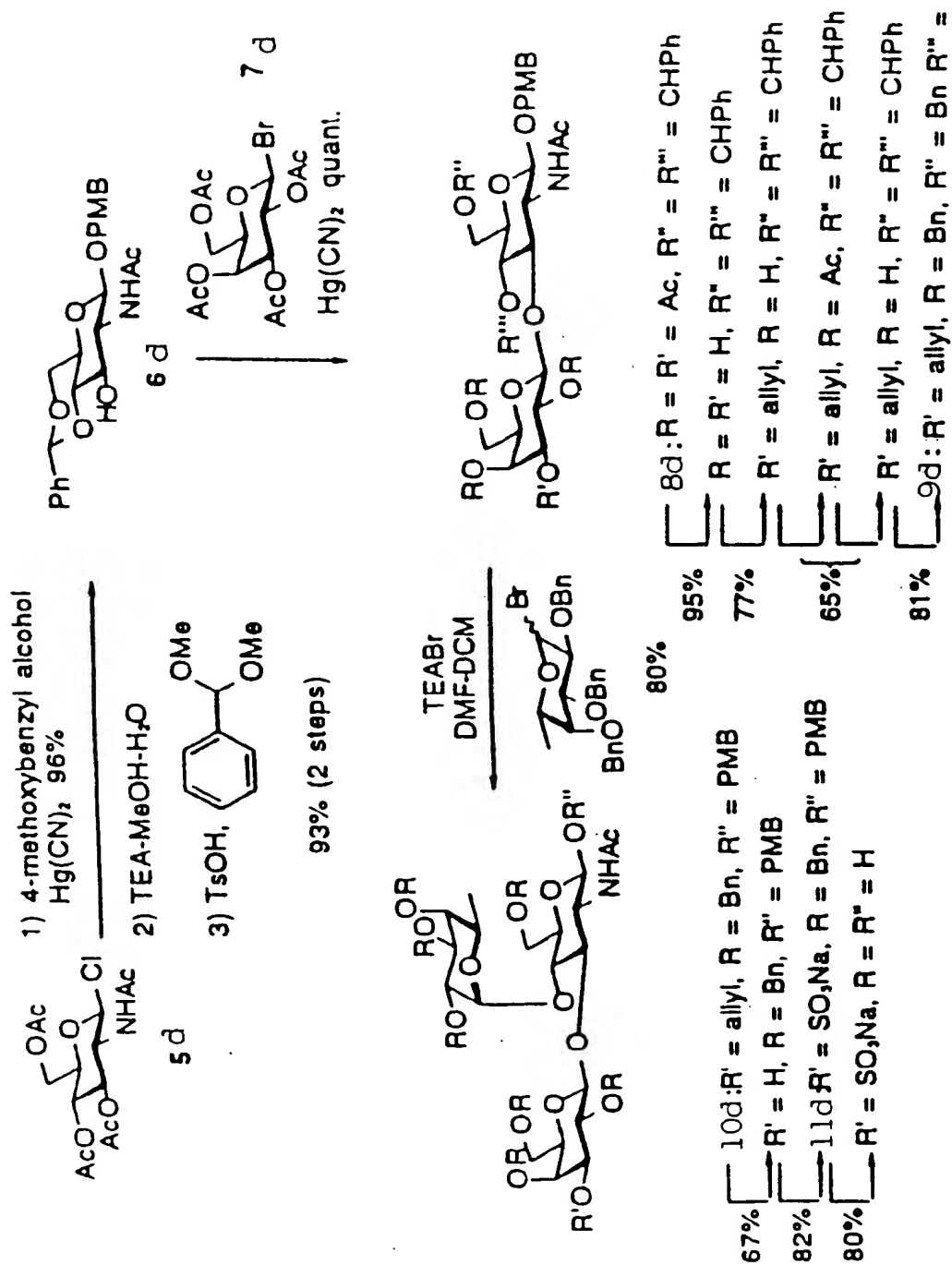
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FIGURE 13A



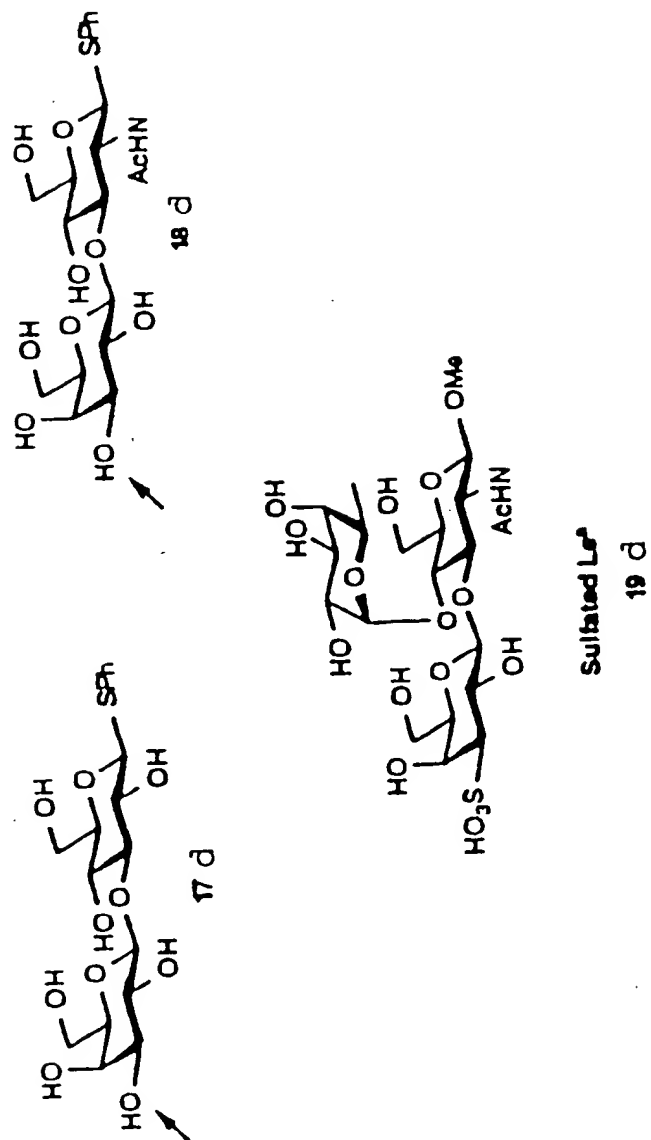
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FIGURE 14A



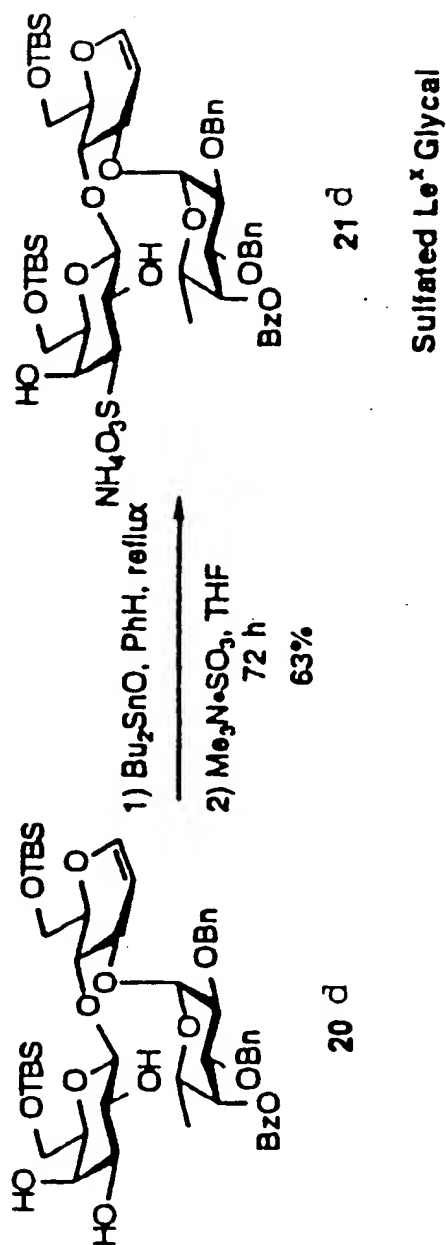
18/34

FIGURE 15A



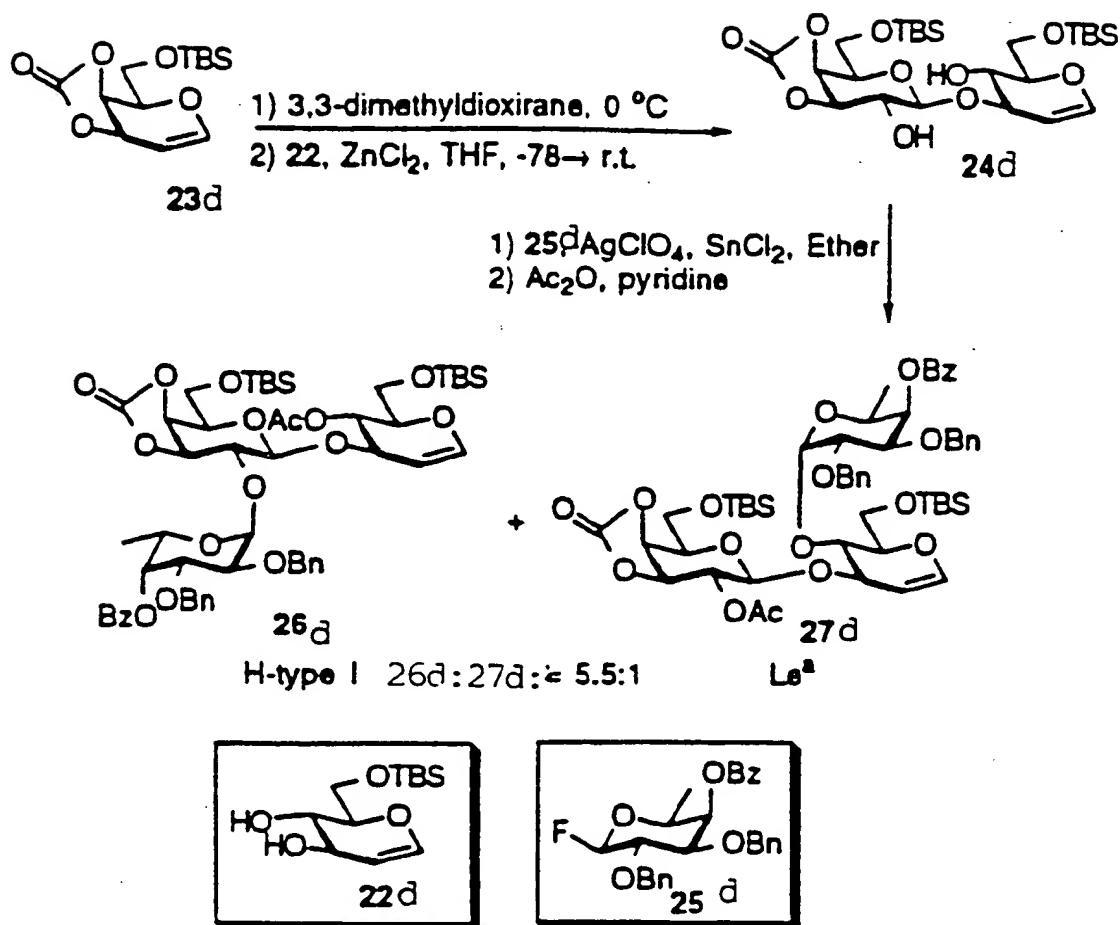
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FIGURE 15B



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FIGURE 15C



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FIGURE 16A

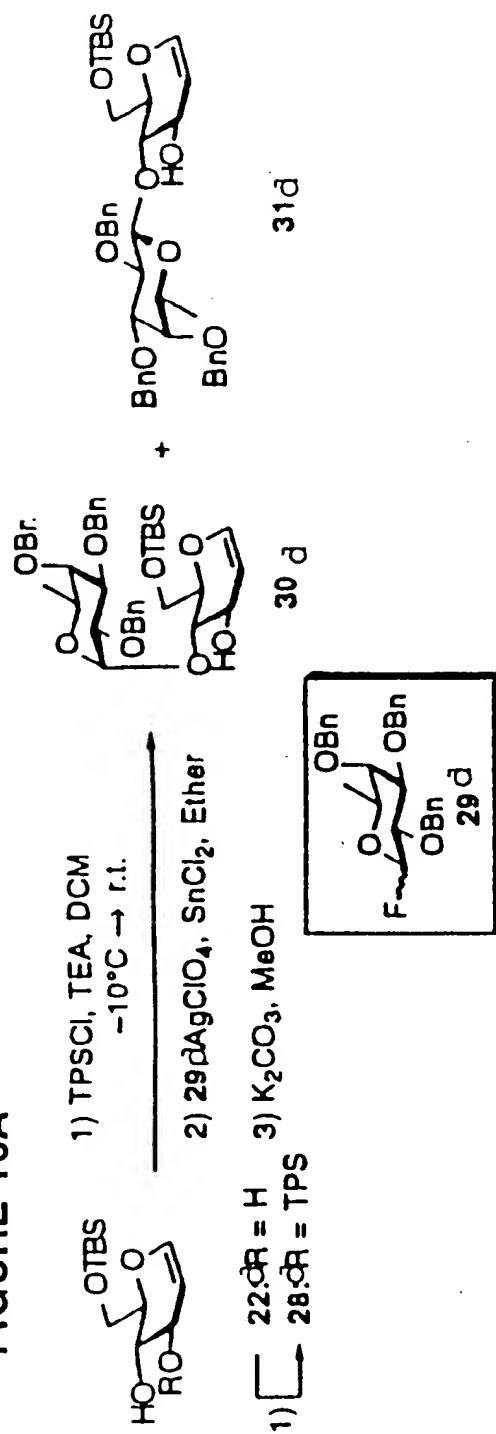
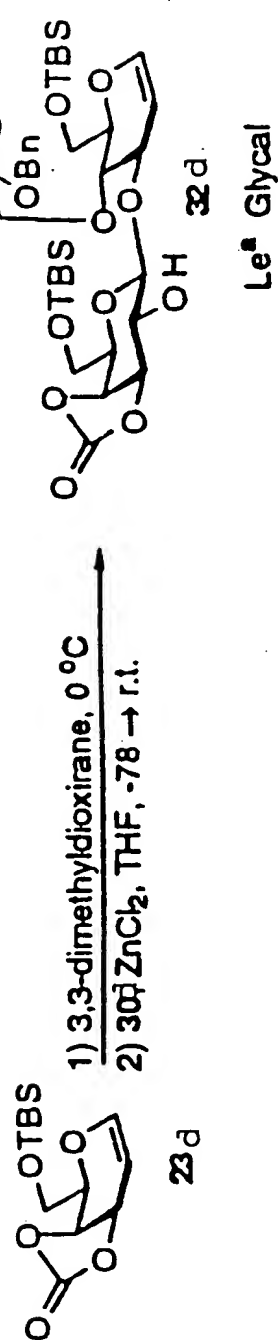
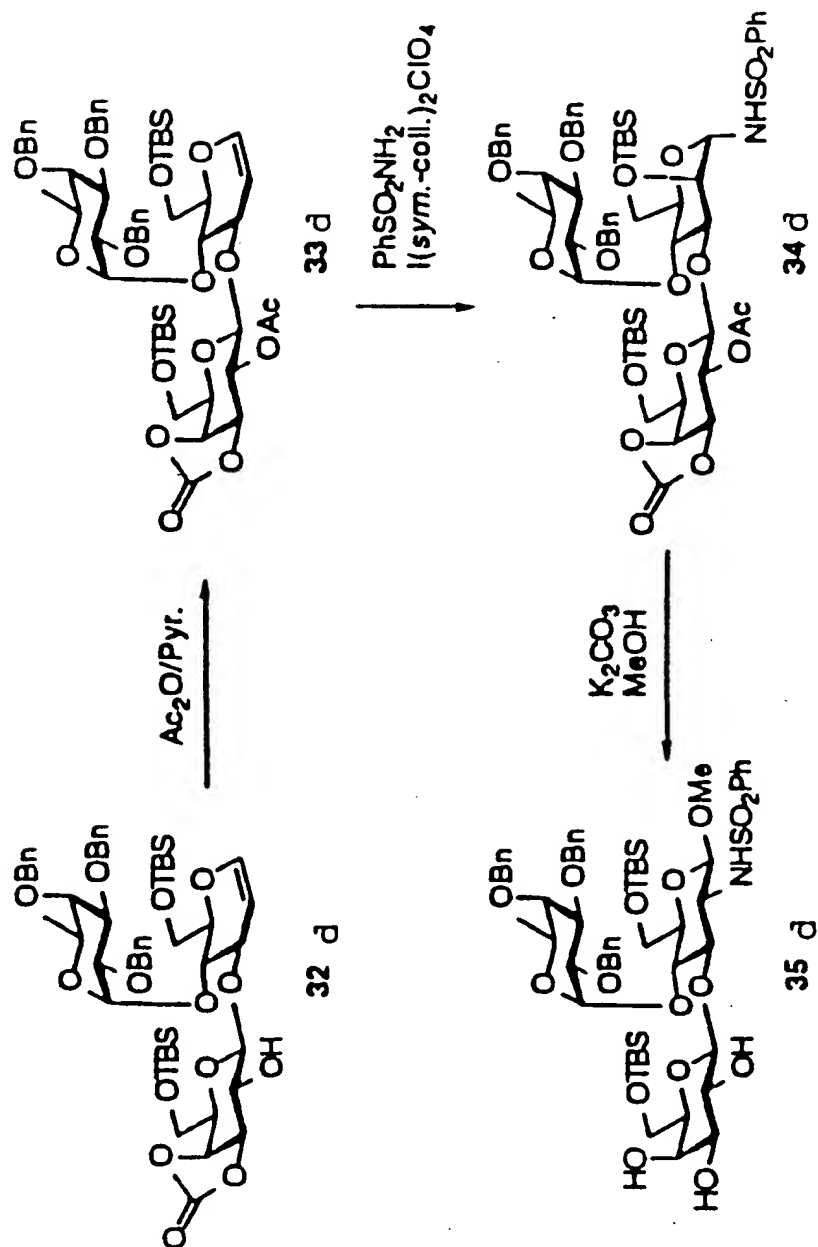


FIGURE 16B



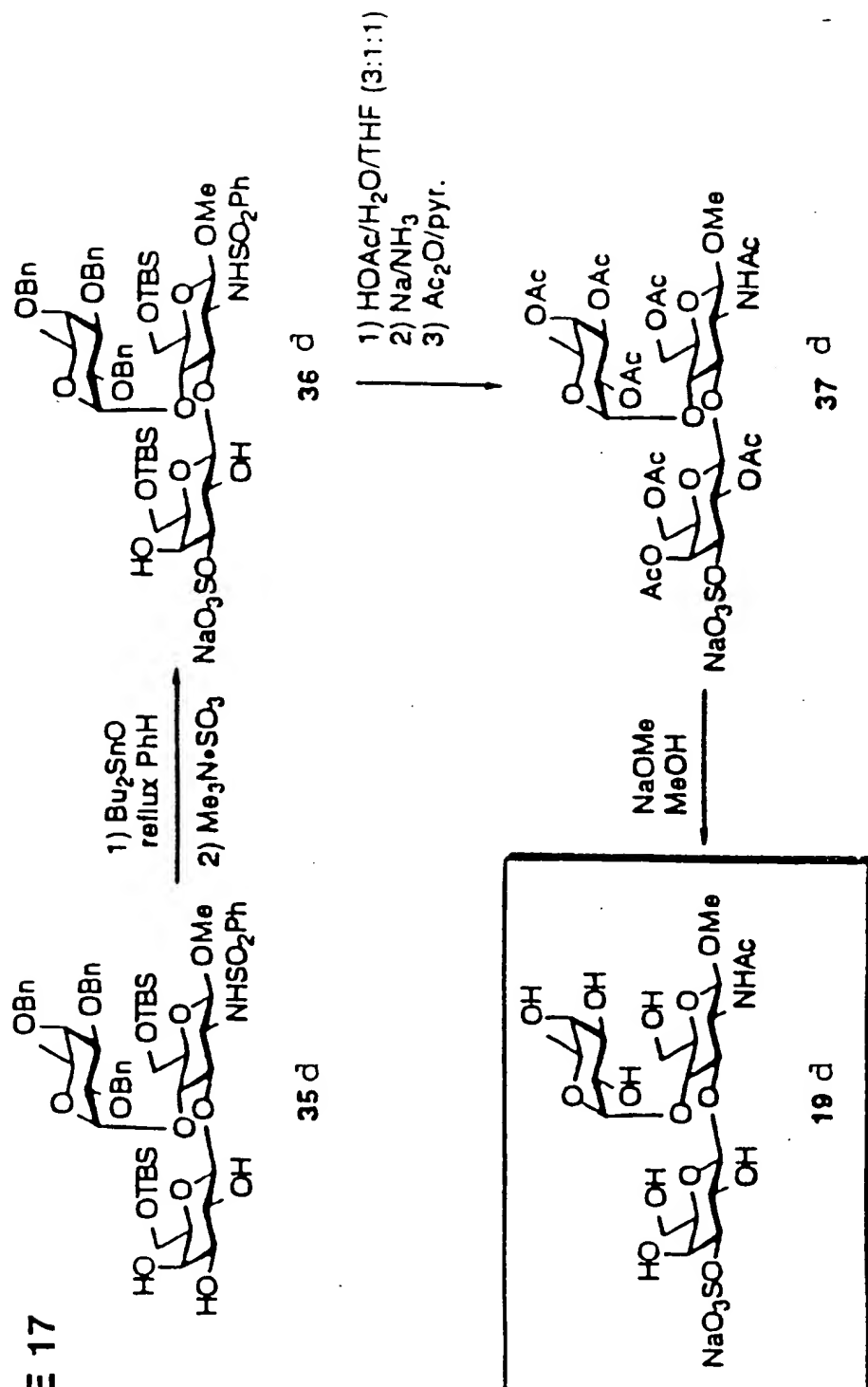
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FIGURE 16C



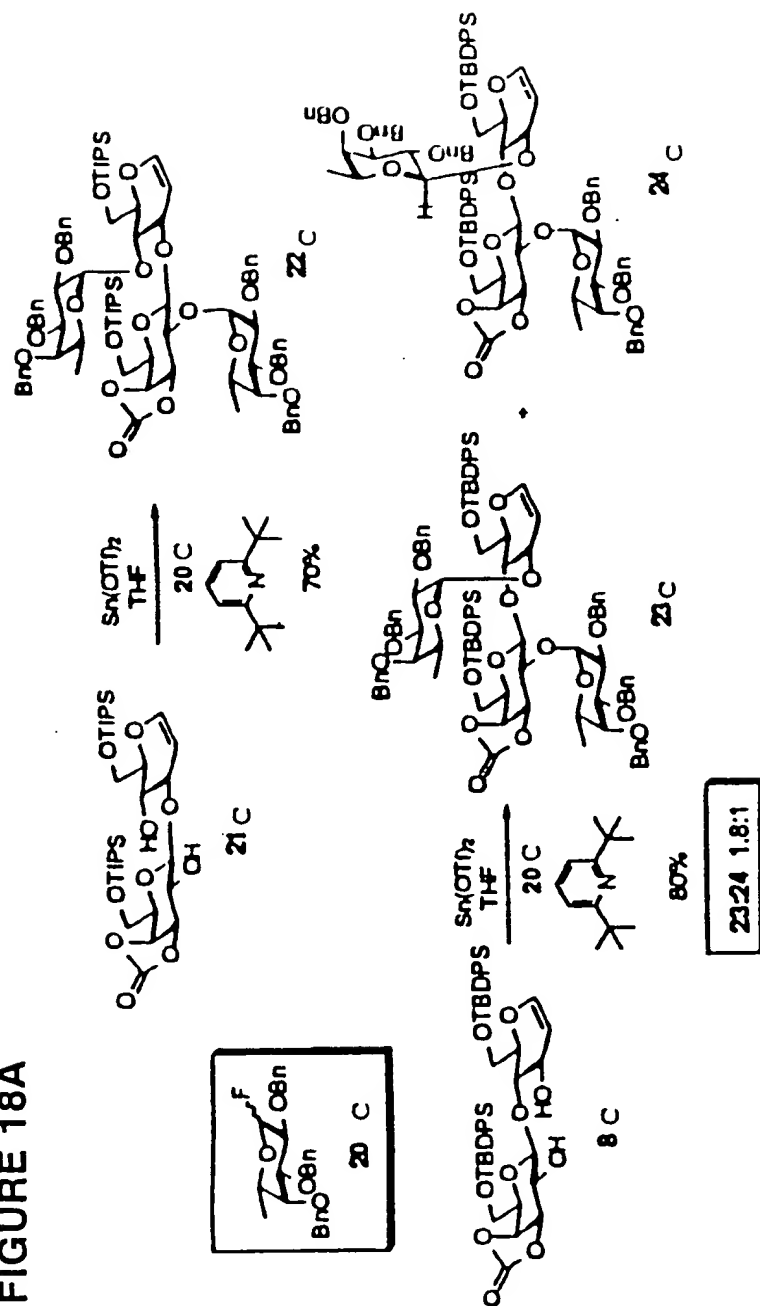
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FIGURE 17



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FIGURE 18A



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FIGURE 18B

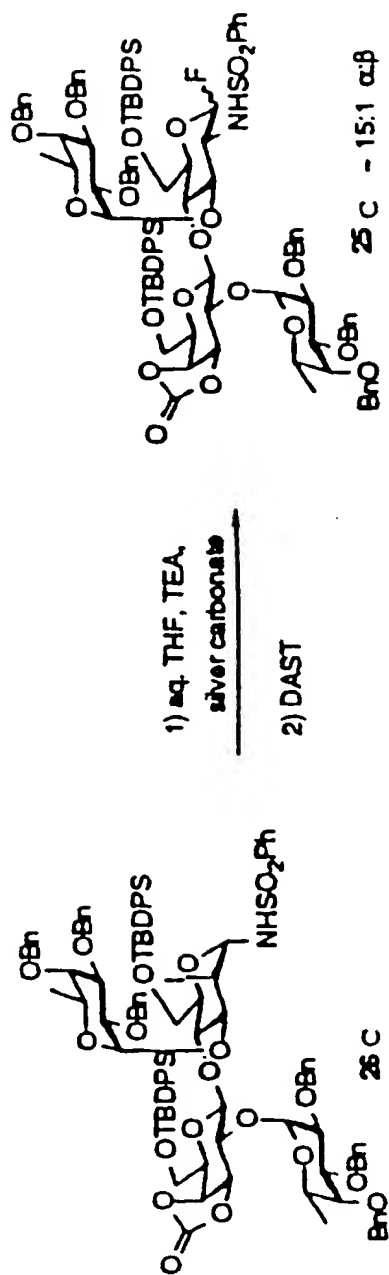
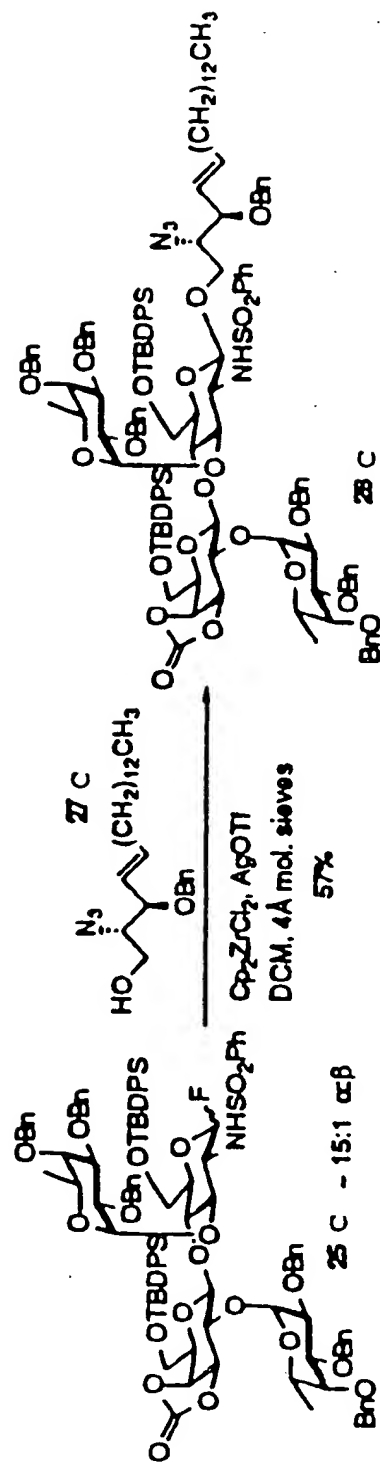
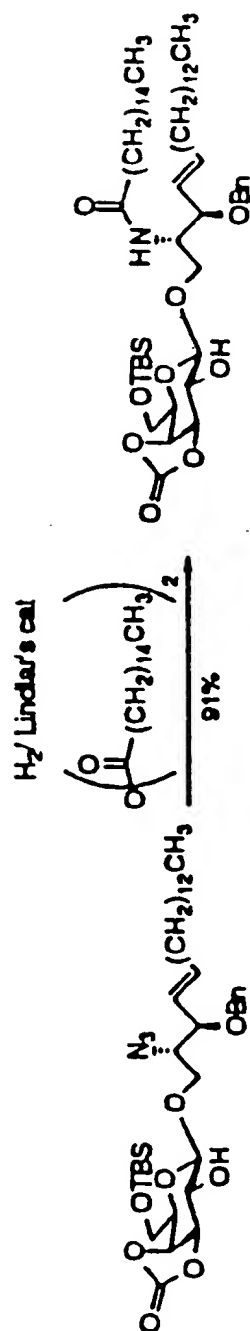


FIGURE 18C



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FIGURE 18D



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FIGURE 19A

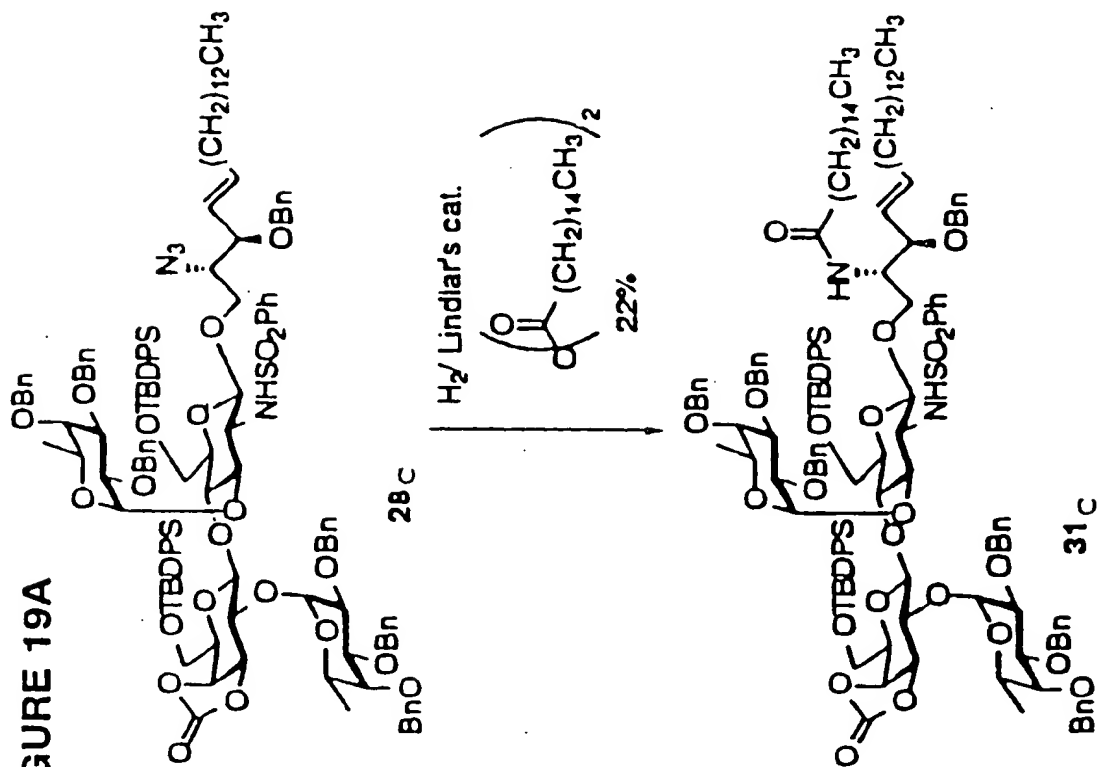
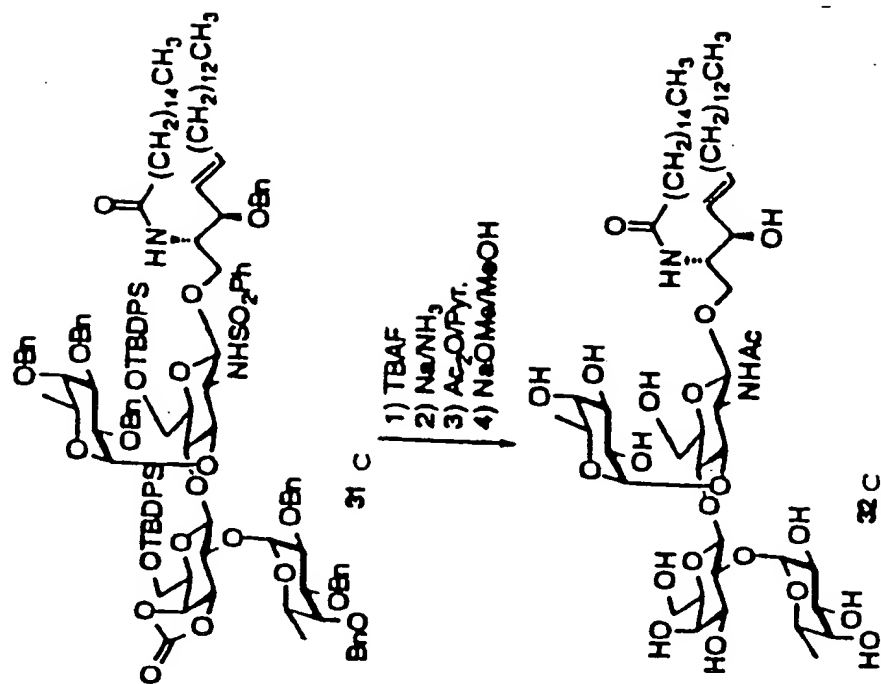


FIGURE 19B



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FIGURE 20A

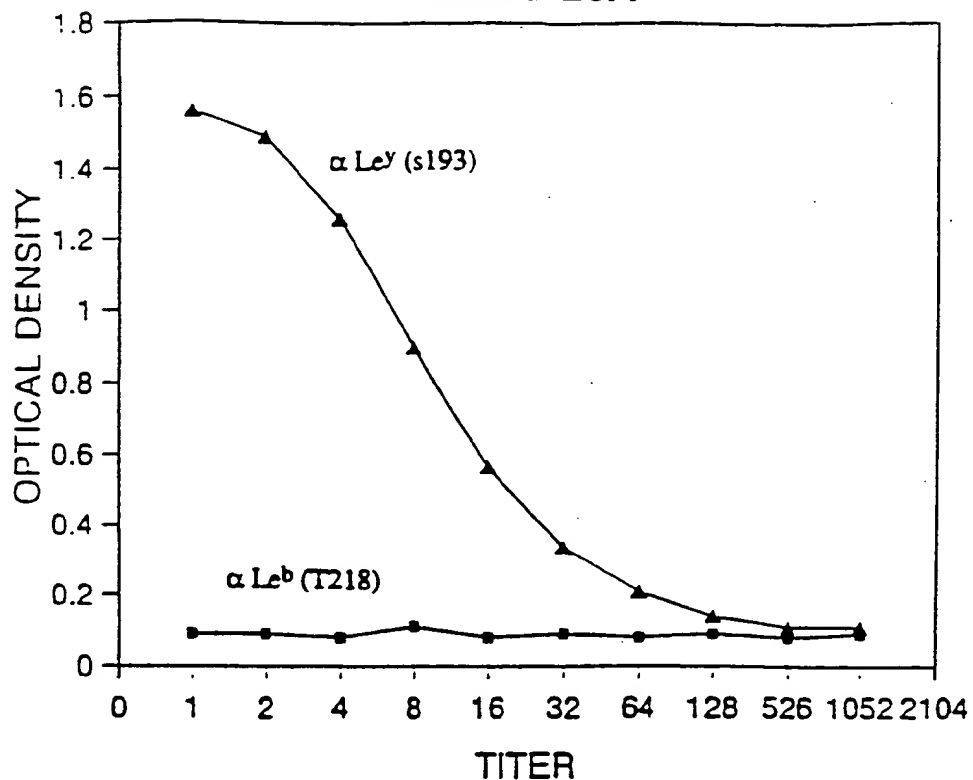
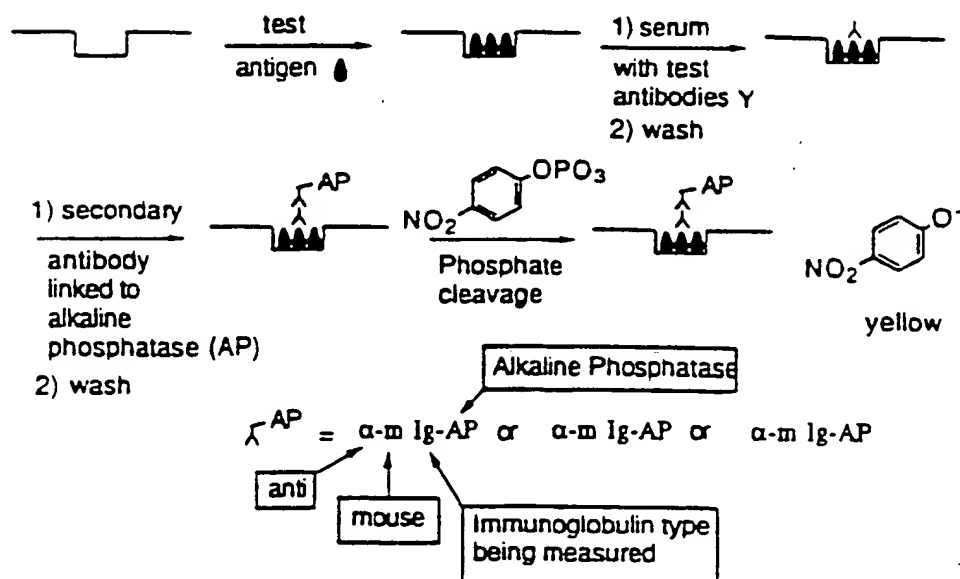
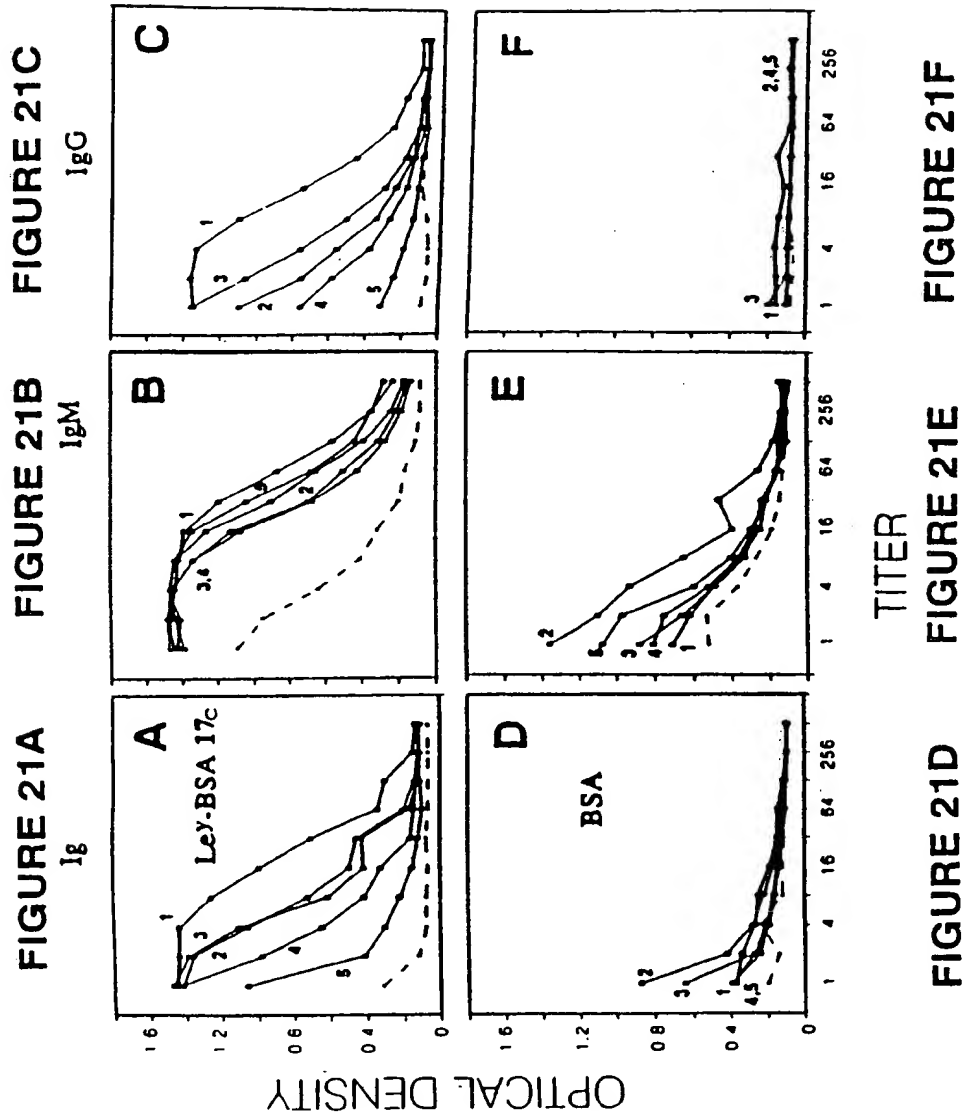


FIGURE 20B



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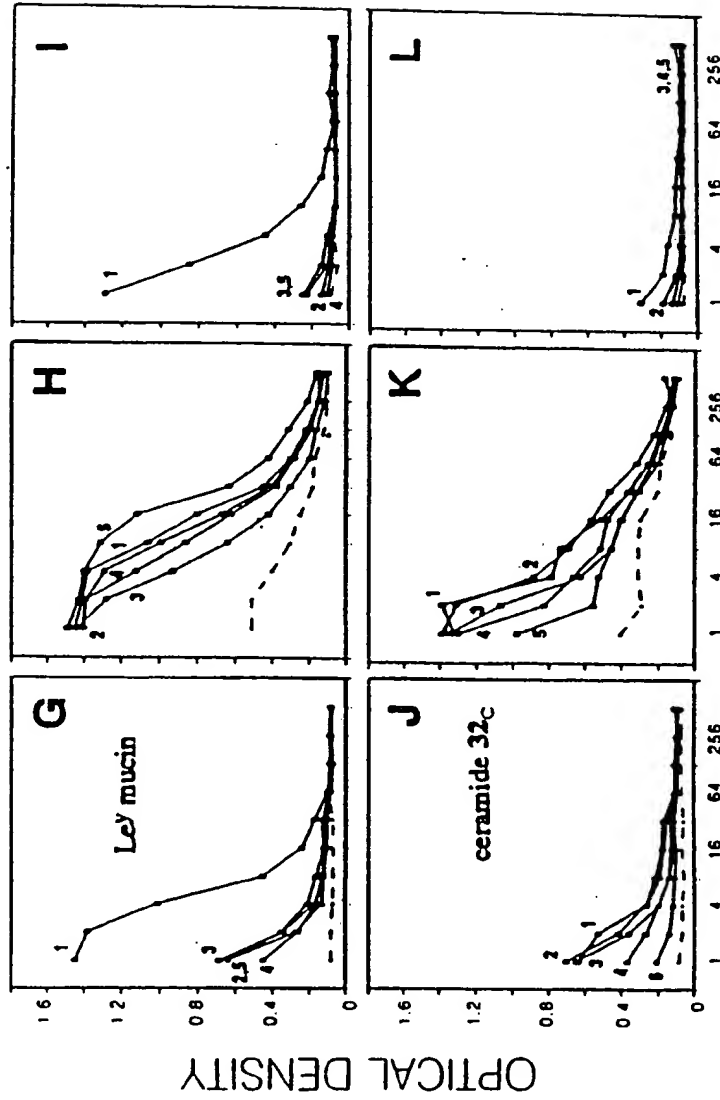


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FIGURE 21G

FIGURE 21H

FIGURE 21I

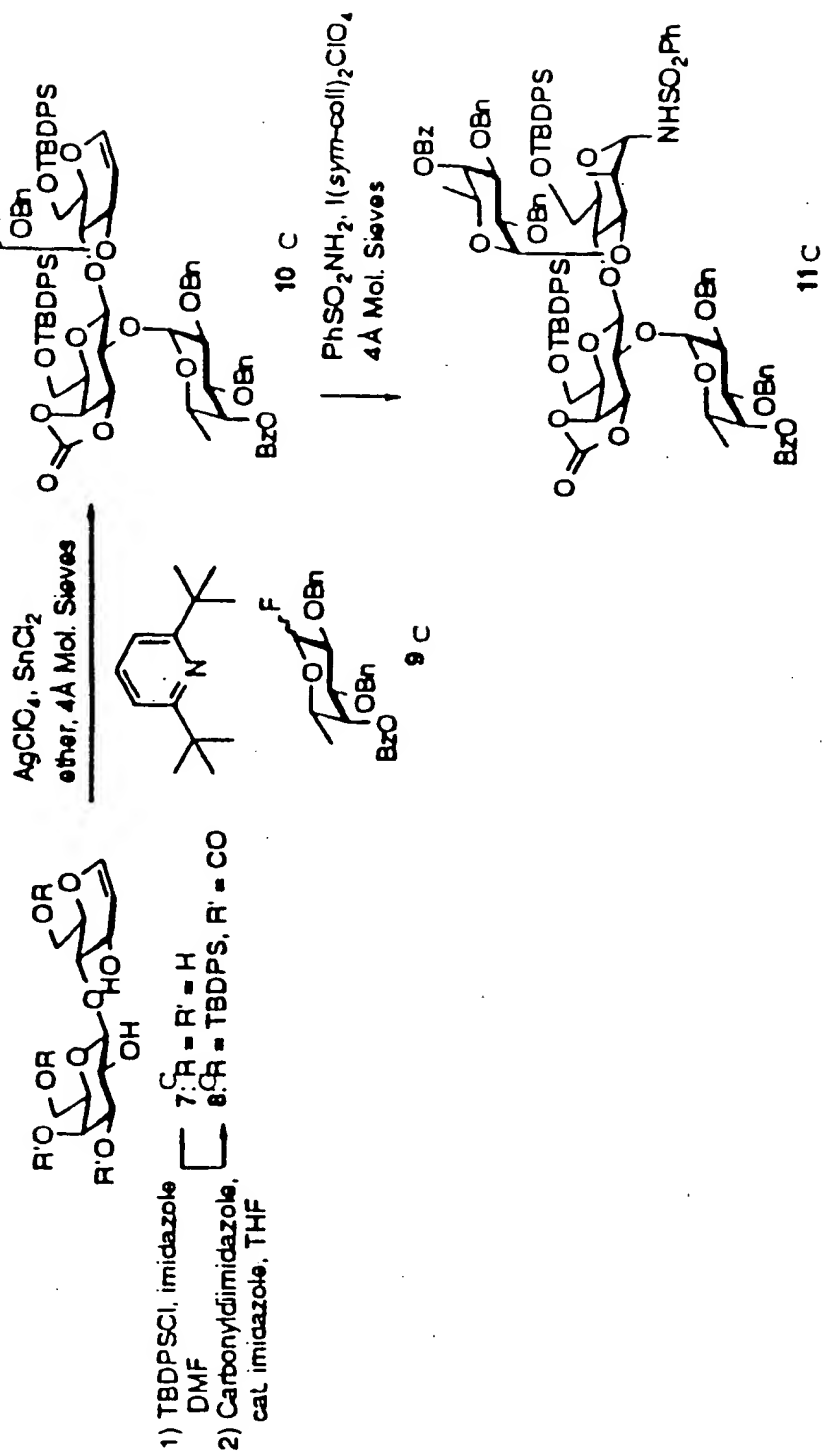


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FIGURE 21J

FIGURE 21K

FIGURE 21L



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FIGURE 22B

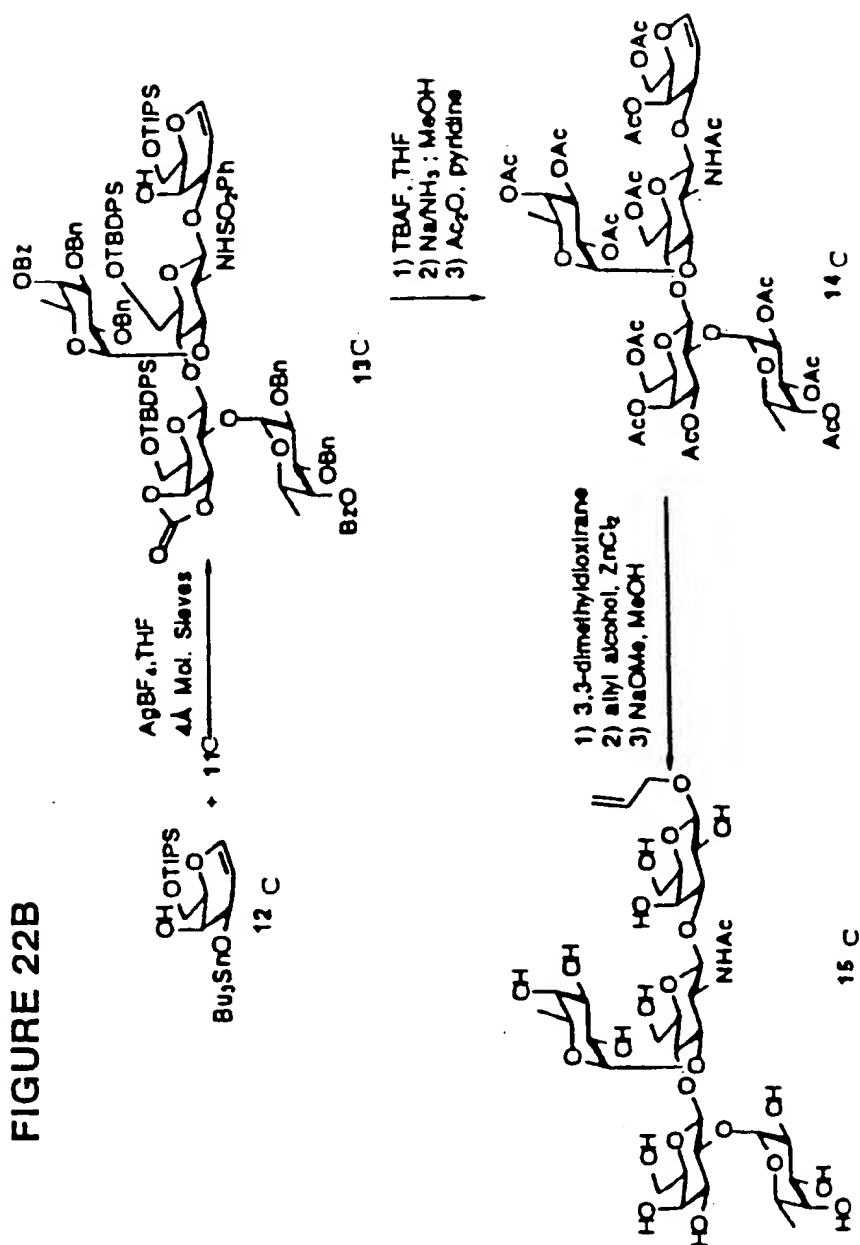
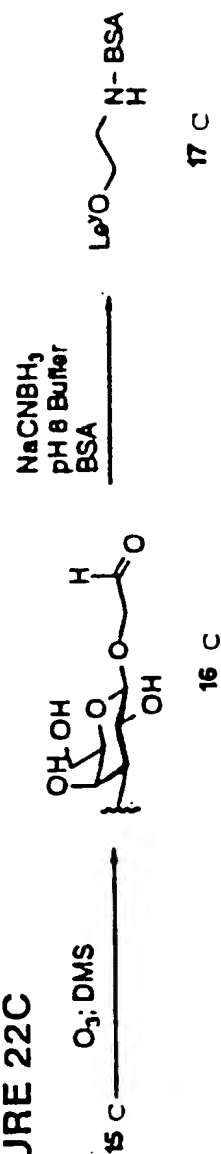


FIGURE 22C



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FIGURE 23A

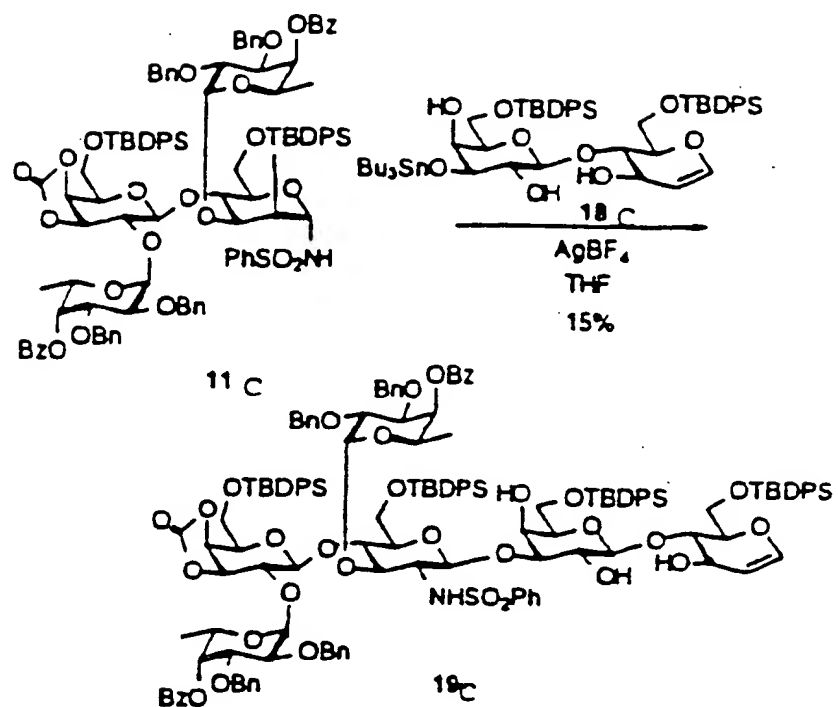
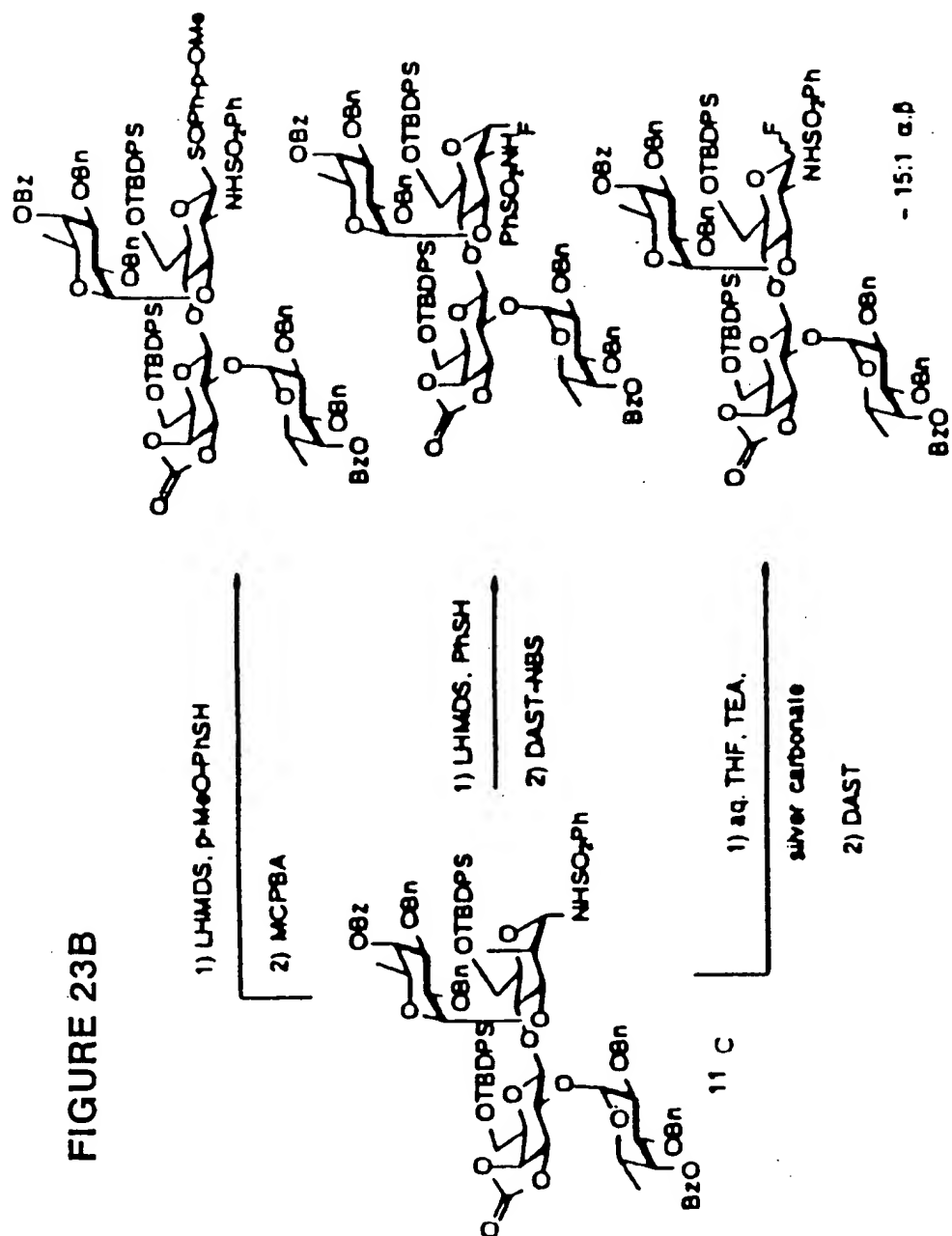


FIGURE 23B



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/12115

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : C07H 15/24, 15/00; A61K 39/395, 39/40

US CL : 536/18.1, 18.6; 424/137.1, 138.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/18.1, 18.6; 424/137.1, 138.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X ---	BEHAR, V. et al. Hochkonvergente Synthese der Blutgruppenderterminante Lewis ^Y in konjugatbildender. Form. Agnew. Chem. 1994, Vol. 106, No. 14, pages 1536-1538, especially pages 1536-1537.	1-5, 11, 12, 14 ---
Y		17-19, 23, 25-27, 31, 33-35, 39, 41-45
X	SALYAN, M.E.K. et al. Differentiation of Type 1 and Type 2 Chain Linkages of Native Glycosphingolipids by Positive-ion Fast-atom Bombardment Mass Spectrometry with Collision-induced Dissociation and Linked Scanning. Rapid Communication in Mass Spectrometry. 1991, Vol. 5, pages 456-462, especially page 459.	16

☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X*	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y*	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*Z*	document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means		
P document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search

03 OCTOBER 1996

Date of mailing of the international search report

24 OCT 1996

Name and mailing address of the ISA/US
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Washington, D.C. 20231

Facsimile No. (703) 305-3230

Authorized officer

SUSAN UNGAR

Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/12115

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	DANISHEFSKY, S.J. et al. Application of the Glycal Assembly Method to the Concise Synthesis of Neoglycoconjugates of Le ^a and Le ^b Blood Group Determinants and of H-Type I and H-Type II Oligosaccharides. J. Am. Chem. Soc. 1995, Vol. 117, pages 5701-5711, especially pages 5701-5706.	16
Y	HELLING, F. et al. G _{D3} Vaccines for Melanoma: Superior Immunogenicity of Keyhole Limpet Hemocyanin Conjugate Vaccines. Cancer Research. 01 January 1994, Vol. 54, pages 197-203, especially pages 197-201.	17-22, 24-30, 32-48
Y	PASTAN, I. et al. Characterization of Monoclonal Antibodies B1 and B3 That React with Mucinous Adenocarcinomas. Cancer Research. 15 July 1991, Vol. 51, pages 3781-3787, especially pages 3781-3786.	17-40
A	LIVINGSTON, P.O. Construction of cancer vaccines with carbohydrate and protein (peptide) tumor antigens. Current Opinion in Immunology. 1992, Vol. 4, pages 624-629.	1-12, 14, 16-48
A	HELLING, G. Ganglioside Conjugate Vaccines. Mol. Chem. Neuropathol. 1994, Vol. 21, pages 299-309.	1-12, 14, 16-48

Form PCT/ISA/210 (continuation of second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/12115

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☒ Claims Nos.: 13 and 15
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

Claim 13 depends upon itself and thus does not give enough information to be able to search. Claim 15 depends upon claim 13 and is unsearchable for that reason.

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:

4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-5, 11-12, 14, 16-48

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

Form PCT/ISA/210 (continuation of first sheet(1))(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US96/12115

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

- I. Claims 1-5, 11-12, 14, and 16-48 are drawn to a method of synthesizing polysaccharides and using the polysaccharides to produce antibodies.
- II. Claims 6-10 are drawn to a method of synthesizing a ceramide.

and it considers that the International Application does not comply with the requirements of unity of invention (Rules 13.1, 13.2 and 13.3) for the reasons indicated below:

Groups I and II are separate methods of manufacture encompassing separate steps.

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